

# *Mycoplasma agalactiae* p40 Gene, a Novel Marker for Diagnosis of Contagious Agalactia in Sheep by Real-Time PCR: Assessment of Analytical Performance and In-House Validation Using Naturally Contaminated Milk Samples<sup>∇†</sup>

Katarína Oravcová,<sup>1,2</sup> Lorena López-Enríquez,<sup>1</sup> David Rodríguez-Lázaro,<sup>3</sup> and Marta Hernández<sup>1\*</sup>

Laboratory of Molecular Biology and Microbiology,<sup>1</sup> and Food Safety and Technology Group,<sup>3</sup> Instituto Tecnológico Agrario de Castilla y León (ITACyL), Valladolid, Spain, and Food Research Institute, Bratislava, Slovakia<sup>2</sup>

Received 28 July 2008/Returned for modification 27 October 2008/Accepted 11 November 2008

We evaluated the capacity of the *Mycoplasma agalactiae* p40 gene as a diagnostic marker for contagious agalactia in sheep by quantitative real-time PCR. The p40 gene encodes an immunodominant adhesin that plays a key role in cytoadhesion of *M. agalactiae*. The assay was 100% specific, with an analytical sensitivity of 1 genome equivalent (GE), a quantification that is highly linear ( $R^2 > 0.992$ ) and efficient (PCR efficiency,  $>0.992$ ) over a 6-log dynamic range, down to 10 GE. We evaluated the capacity of the assay to detect *Mycoplasma agalactiae* in 797 milk samples (373 raw sheep milk samples from refrigerated tanks of different farms and 424 milk samples from individual sheep of a flock positive for *M. agalactiae*). In parallel, we also tested the samples by using microbiological isolation coupled with microscopy identification and by a PCR method recommended by the World Organization for Animal Health. While our assay was able to detect 57 (15.28%) positive samples of the 373 milk samples from different farms, identification by microbiological isolation coupled with microscopy detected only 36 (9.65%) samples, and the conventional PCR detected 31 (8.31%) samples. These findings showed that our assay based on the p40 gene is more specific and sensitive for the detection of *M. agalactiae* in actual natural samples and, thus, can be a promising alternative tool for diagnosis and epidemiological studies of *M. agalactiae* infection.

*Mycoplasma agalactiae* is the principal etiological agent of contagious agalactia in small ruminants. In male and female sheep, this syndrome is produced mostly by *Mycoplasma agalactiae* subsp. *agalactiae*, but other mycoplasmas are also implicated (*M. capricolum* subsp. *capricolum*, *M. mycoides* subsp. *capri*, *M. mycoides* subsp. *mycoides* LC, and *M. putrefaciens*). Contagious agalactia is a serious infectious process characterized by three major signs: mastitis, arthritis, and keratoconjunctivitis (17). It causes the reduction and even the suppression of milk production and occasionally causes abortion and death in up to 70% of young animals, resulting in serious economic loss (16). Nowadays, this syndrome affects animals in most countries that have a high production of sheep and goats. It is found across several continents, including mainly North America, western Asia, North Africa, and Europe, and is endemic in most Mediterranean countries (3). Consequently, this disease represents a very important animal health problem and is included in the list of notifiable diseases listed by the OIE (World Organization for Animal Health) because of its economic impact and widespread distribution. In fact, governments are implementing a compulsory diagnosis scheme for assessing the presence *M. agalactiae* in milk tanks or farms by PCR (7).

The principal sources of *M. agalactiae* infection are the ingestion of contaminated feed, water, or milk and the urine, feces, and nasal or ocular liquids of infected animals. Ewes can be infected through the udder, and lambs can be infected by the consumption of colostrum. Thus, it is recommended that newborn animals be removed from the dam immediately after birth and fed only by using pasteurized colostrum. The period of incubation of this syndrome varies between 7 and 56 days. Most cases of infection occur in the summer, during birth and peak lactation periods. Several mycoplasmas can be isolated from milk and blood for a short period of time during the infectious process (5).

The routine analysis method used by diagnostic laboratories for identifying *M. agalactiae* in clinical samples or milk tanks is based on microbiological culture in selective enrichment media including antibiotics in moist anaerobic chambers (5% CO<sub>2</sub>) for 5 to 7 days. However, this method represents a time-consuming and complicated process, as mycoplasmas grow very slowly. Mycoplasmas form typical “fried-egg”-shaped small colonies that, although visible under the microscope, are very difficult to enumerate. Moreover, these analyses need to be complemented with species identification by biochemical or immunofluorescence tests, making this an expensive procedure. Alternative DNA amplification-based methods for directly detecting this animal pathogen in milk or in other fluids have been devised to overcome this problem. There are several methods that identify *M. agalactiae* by PCR. Some of these methods are based on amplification of the 16S rRNA gene (2, 4, 12, 13). However, 16S rRNA gene sequences in *M. agalactiae* and *M. bovis* share 99.8% similarity, affecting the specificity of

\* Corresponding author. Mailing address: Instituto Tecnológico Agrario de Castilla y León (ITACyL), Carretera de Burgos, km. 119, 47071 Valladolid, Spain. Phone: 34 983 317542. Fax: 34 983 410462. E-mail: ita-herperma@itacyl.es.

† Supplemental material for this article may be found at <http://jcm.asm.org/>.

∇ Published ahead of print on 19 November 2008.

TABLE 1. Oligonucleotides used in this study

Target	Primer	Type	Sequence
<i>M. agalactiae</i> <i>p40</i> gene <sup>a</sup>	MAP40127F	Forward	5'-TCATTTACAGCAGTGCCTTTATTAG-3'
	MAP40235R	Reverse	5'-CACCTAATGCTTGTITTTCAACC-3'
	MAP40160P	TaqMan probe	5'-FAM-TGTGATGATAAGAACGAAAATTCACA AA-BHQ1-3' <sup>b</sup>
IAC chimeric DNA	MAIACP	TaqMan MGB probe	5'-VIC-CCATACACATAGGTCAGG-MGB-NFQ-3'
<i>L. monocytogenes</i> <i>prfA</i> gene	MAIACF	Forward	5'-TCATTTACAGCAGTGCCTTTATTAGGGCTC TATTTGCGGTC-3'
	MAIACR	Reverse	5'-CACCTAATGCTTGTITTTCAACCTCTTGATG CCATCAGGA-3'

<sup>a</sup> The amplicon size is 109 bp, and the melting temperature of the amplicon is 73.0°C ± 0.6°C.

<sup>b</sup> BHQ1, Black Hole Quencher 1.

methods based on the amplification of this gene. Other diagnostic strategies are based on the amplification of unknown sequences (6, 27, 28) or specific genes, like *uvrC* (26) or the *mb-mp81* gene encoding the membrane protein P81 (9). However, the last two methods are based on the PCR-restriction fragment length polymorphism technique, which is more laborious and time-consuming and is not quantitative.

The absolute quantification of contaminant microbiota by real-time quantitative PCR (Q-PCR) is becoming increasingly common for diagnostic purposes in clinical and food microbiology (24). This technique provides a higher specificity and analytical sensitivity and reduces the risk of cross-contamination; at the same time, this technique is faster than conventional PCR and is totally adapted to automation. Lorusso and coworkers (15) have developed a real-time PCR detection method using molecular beacon chemistry. This method targets a 117-bp region of the *M. agalactiae* *mb-mp81* gene encoding the membrane lipoprotein P81, which is also present in the *M. bovis* genome. There is a general consensus that the addition of an internal amplification control (IAC) in each reaction mixture, to assess the potential effects of PCR inhibitors or the malfunction of thermocyclers, must be mandatory. An IAC is a chimeric nontarget DNA fragment that is present in every reaction mixture and can be coamplified with the target sequence (11). However, the method described by Lorusso and coworkers incorporates an alternate template (canine parvovirus type 2 [CPV-2] DNA), which is run in separate PCR wells (15).

It is clearly convenient for diagnostic laboratories to have a large battery of alternative analytical methods for the quantitative detection of *M. agalactiae*, due to the economic impact and public health relevance of this pathogen. An alternative method can be adopted by diagnostics laboratories only if the method has been evaluated and validated with samples collected from animals. Thus, in this study, we report the design and assessment of a real-time PCR assay for the quantitative detection of *M. agalactiae* and its evaluation and in-house validation in the analysis of milk samples. This method includes an IAC that is coamplified in the same reaction mixture to assess the PCR performance of each reaction, thus ensuring the diagnostic efficiency of this method. Finally, we evaluated the performance of the assay with natural samples, i.e., sheep milk samples. We tested 373 natural, raw sheep milk samples from refrigerated tanks from different sheep farms and 424

milk samples from individual sheep from a flock found positive for this pathogen.

**Design and optimization of the *M. agalactiae*-specific duplex *p40*-IAC Q-PCR assay.** The assay targets an *M. agalactiae* species-specific region of the *p40* gene (GenBank accession no. AJ344229) encoding an immunodominant adhesin that plays a key role in cytoadhesion of *M. agalactiae* (8). Bacterial adhesion is a key mechanism of mycoplasma virulence, and the protein P40 displays a strong and persistent signal in response to antibodies; thus, the *M. agalactiae* *p40* gene has been proposed as a good candidate for the development of future diagnostic assays (8). The *p40* gene sequence displayed regions that were 100% specific to *M. agalactiae* by nucleotide sequence comparison using BLAST-N version 2.2.14 (National Centre for Biotechnology Information; www.ncbi.nlm.nih.gov). We selected *M. agalactiae*-specific regions that were identical among various isolates of *M. agalactiae* by aligning all *p40* sequences available in public databases, using a CLUSTALW multiple-alignment tool (EMBL, European Bioinformatics Institute; www.ebi.ac.uk); this was done to ensure no variation in sequences occurred as a result of different strains. Subsequently, we designed the PCR primers MAP40127F, MAP40235R, and MAP40160P by using Primer Express version 3.0 software (Applied Biosystems, Foster City, CA) (Table 1). These PCR oligonucleotides amplify a 109-bp fragment from the coding sequence of the *M. agalactiae* *p40* gene corresponding to positions 127 to 235. The specificity of the oligonucleotides was confirmed in silico using BLAST-N version 2.2.14 software, as none of the selected oligonucleotides or amplicon sequences showed any similarity outside of the *M. agalactiae*-specific region.

One of the major barriers to the systematic introduction of Q-PCR-based methods for routine microbiology diagnostics is the common occurrence of false-negative results. These results occur due to the presence of PCR inhibitors in clinical and environmental samples. This is particularly relevant in analyses performed for detecting *M. agalactiae* cells in raw sheep milk. Raw sheep milk contains various substances that may cause the bacterial counts to be underestimated or the amplification reaction to be totally abolished (1). Our assay was designed to monitor this problem, using an IAC. This IAC consisted of a nontarget nucleic acid fragment that is coamplified with the target sequence. The same primers are used for the test reaction, including a probe labeled differently, for the simultaneous

assessment of PCR performance. The IAC was constructed as previously described (22) and consists of a 118-bp chimeric DNA molecule containing a portion (nucleotide positions 421 to 491) of the *Listeria monocytogenes*-positive regulatory factor A (*prfA*) gene; the gene is flanked by the *M. agalactiae*-specific *p40* gene sequences targeted by the MAP40127F and MAP40235R primers. If a negative signal is obtained for the target *p40* signal, the absence of a positive IAC signal indicates that amplification has failed.

The *M. agalactiae*-specific PCR oligonucleotides were optimized for Q-PCR assays in which 1 ng of DNA from *M. agalactiae* strain ATCC 35890 was used as a template. The assays were carried out in a reaction mixture volume of 20  $\mu$ l containing 1 $\times$  Universal PCR Master Mix (Applied Biosystems). We used a model 7500 real-time PCR system platform (Applied Biosystems) with the following program: 2 min at 50°C, 10 min at 95°C, and 50 cycles of 15 s at 95°C and 1 min at 60°C. Optimal conditions (300 nM for primers MAP40127F and MAP40235R and 200 nM for the 6-carboxyfluorescein [FAM]-labeled MAP40160P probe) were the minimum primer and probe concentrations giving the lowest cycle threshold ( $C_T$ ) value and the highest fluorescence intensity for a normalized reporter value. The optimal IAC probe concentration was determined experimentally by carrying out Q-PCRs in the presence of 10,000 IAC molecules, no *M. agalactiae* DNA, 200 nM FAM-labeled *p40* probe and with various amounts (from 25 to 250 nM) of VIC-labeled IAC probe (Table 1). The minimum probe concentration that did not result in an increase in  $C_T$  values was 100 nM. As excess IAC may inhibit the target-specific reaction, Q-PCRs were also carried out in the presence of various amounts of IAC (10,000, 1,000, 100, and 10 molecules per reaction) and various amounts of *M. agalactiae* DNA (equivalent to approximately  $1 \times 10^4$ ,  $1 \times 10^3$ , 100, 10, and 1 genome equivalents, in which there are 0.88 Mb per genome, according to Sirand-Pugnet and coworkers [25], and one genome equivalent corresponds to 0.972 fg of *M. agalactiae* DNA). The maximum amount of IAC with no inhibitory effect on the *p40*-specific FAM signal was 100 copies of chimeric DNA.

**Selectivity of the *p40*-IAC assay.** The capacity of the *p40*-IAC Q-PCR assay to discriminate between target and nontarget bacteria was assessed using 1 ng of genomic DNA from various sources, as follows: 75 *Mycoplasma* species, including 26 *M. agalactiae* isolates (24 of them were sheep milk isolates) and 49 other *Mycoplasma* species; and 110 strains from 39 non-*Mycoplasma* species, including various genera frequently found in milk and dairy products (see Table S1 and S2 in the supplemental material). Bacterial DNA was isolated using a QIAamp DNA minikit (Qiagen, Hilden, Germany). The *p40*-IAC Q-PCR assay was inclusive for *M. agalactiae* and 100% exclusive for nontarget bacteria: all *M. agalactiae* strains tested gave a positive *p40* signal, whereas none of the 138 nontarget bacteria produced a positive signal. All reactions generated a positive IAC (VIC) signal, excluding the possibility that the absence of a *p40* (FAM) signal observed for non-*M. agalactiae* isolates was due to PCR failure.

**Analytical sensitivity and quantification range of the *p40*-IAC Q-PCR assay.** The achievement of low detection and quantification limits is a critical aspect in the design of molecular diagnostic methods for microbial pathogens in clinical and

TABLE 2. Detection and quantification limits of Q-PCR assay with genomic DNA from *M. agalactiae* strain ATCC 35890<sup>a</sup>

Approximate genome equivalents/reaction	<i>p40</i> Q-PCR		<i>p40</i> -IAC Q-PCR	
	Signal ratio	$C_T$ value $\pm$ SEM	Signal ratio	$C_T$ value $\pm$ SEM
$1 \times 10^7$	9/9	17.13 $\pm$ 0.09	9/9	17.16 $\pm$ 0.06
$1 \times 10^6$	9/9	20.77 $\pm$ 0.06	9/9	20.81 $\pm$ 0.07
$1 \times 10^5$	9/9	24.49 $\pm$ 0.11	9/9	24.31 $\pm$ 0.04
$1 \times 10^4$	9/9	27.98 $\pm$ 0.13	9/9	28.08 $\pm$ 0.04
$1 \times 10^3$	9/9	31.40 $\pm$ 0.12	9/9	31.42 $\pm$ 0.12
$1 \times 10^2$	9/9	33.31 $\pm$ 0.16	9/9	33.42 $\pm$ 0.17
$1 \times 10^1$	30/30	36.15 $\pm$ 0.20	30/30	36.88 $\pm$ 0.40
1	16/30	38.45 $\pm$ 0.40	15/30	38.56 $\pm$ 0.20

<sup>a</sup> Nontemplate controls for both Q-PCR systems were negative ( $C_T$  values of 50 for all the replicates). The overall slopes of the regression curve were  $-3.3216$  and  $-3.3406$  for the TaqMan *p40* and *p40*-IAC Q-PCR TaqMan systems, respectively, indicating PCR efficiency values of 1.000 and 0.992, respectively, and the regression coefficients ( $R^2$ ) were 0.9921 and 0.9931, respectively. Signal ratio indicates the number of positive results out of 9 reactions, 30 reactions for those containing 10 and 1 genome equivalents per reaction.  $C_T$  value  $\pm$  standard error of the mean (SEM) indicates the mean  $C_T$  value at which fluorescence intensity was equal to a fixed threshold. Experimental results were statistically significant ( $P < 0.05$ ), taking into account the unavoidable error associated with serial dilutions.

environmental samples. The detection and quantification limits of the PCR assays were determined by using genomic DNA isolated from *M. agalactiae* strain ATCC 35890. Three independent PCRs were performed, with a range of DNA concentrations equivalent to approximately  $1 \times 10^6$ ,  $1 \times 10^5$ ,  $1 \times 10^4$ ,  $1 \times 10^3$ ,  $1 \times 10^2$ , 10, and 1 target molecules. Table 2 shows the mean  $C_T$  values for a total of 9 PCR replicates (30 replicates for 10 and 1 genome equivalents) from three independent experiments. Positive amplification of all PCR replicates of each DNA dilution was achieved if 10 or more target molecules were present, and 1 target molecule could be detected in at least 16 of the 30 replicates (Table 2). PCR efficiency is related to the slope of the linear regression curve calculated over a 6-log range (down to 10 target molecules) and was similar to that of the theoretical optimum of  $-3.32$  (*p40*,  $-3.3216$ , efficiency = 100%; *p40*-IAC,  $-3.3416$ , efficiency = 99.2%). The linearity of calibration curves represented by the regression coefficient ( $R^2$ ) showed values close to 1 (*p40*, 0.9921; *p40*-IAC, 0.9931), indicating that the assay was highly linear. These results are similar to those for other Q-PCR methods used for other bacterial and eukaryotic organisms (10, 19–21, 23).

**Quantitative detection of *M. agalactiae* in raw sheep milk.** Samples that possess a strong capacity to inhibit PCR amplification pose the risk of false-negative results (1). A complex matrix like raw sheep milk directly transferred from the farm milk tank was selected to illustrate the implementation of the *p40*-IAC PCR method and to evaluate the capacity for quantitative detection of *M. agalactiae*. In three independently replicated experiments, 1-ml portions of phosphate-buffered saline containing approximately  $2.5 \times 10^7$ ,  $2.5 \times 10^6$ ,  $2.5 \times 10^5$ ,  $2.5 \times 10^4$ ,  $2.5 \times 10^3$ , 250, 25, or 3 *M. agalactiae* cells were added to 50-ml centrifuge tubes, each containing 25 ml of raw sheep milk. DNA was purified from milk by the procedure described by López-Enríquez and coworkers (14), with the only modification being homogenization for 10 min in a homogenizer (Pulsifier) prior to DNA extraction. We consistently



TABLE 3. Quantitative detection of *M. agalactiae* in raw sheep milk<sup>a</sup>

Approximate <i>M. agalactiae</i> cells/25 ml	Approximate <i>M. agalactiae</i> genome equivalents/reaction <sup>b</sup>	Signal ratio <sup>c</sup>	$C_T$ value $\pm$ SEM <sup>d</sup>	Relative accuracy <sup>e</sup>
$2.5 \times 10^7$	$1 \times 10^5$	9/9	$24.20 \pm 0.02$	87.17
$2.5 \times 10^6$	$1 \times 10^4$	9/9	$27.12 \pm 0.06$	115.14
$2.5 \times 10^5$	$1 \times 10^3$	9/9	$30.50 \pm 0.05$	110.98
$2.5 \times 10^4$	$1 \times 10^2$	9/9	$34.09 \pm 0.06$	91.81
$2.5 \times 10^3$	10	9/9	$37.33 \pm 0.23$	97.43
$2.5 \times 10^2$	1	9/9	$38.63 \pm 0.32$	NA
25	0.1	0/9	NA <sup>f</sup>	NA
3	0.01	0/9	NA	NA
0 <sup>g</sup>	0	0/9	NA	NA

<sup>a</sup> Raw milk was collected directly from a sheep farm. Data show results from three independent experiments with three PCR replicates used in each. The overall efficiency was 0.996, and the linearity ( $R^2$ ) was 0.9989. NA, not applicable.

<sup>b</sup> Estimated numbers of *M. agalactiae* genome equivalents in each PCR run, assuming 100% DNA extraction efficiency, are shown. Each reaction mixture contained 2  $\mu$ l of a DNA preparation taken from the 1:10 dilution of the initial 50  $\mu$ l extracted from 25 ml of raw sheep milk.

<sup>c</sup> Number of positive results out of nine reactions.

<sup>d</sup> Mean  $C_T$  value  $\pm$  standard error of the mean (SEM) at which fluorescence intensity was equal to a fixed threshold. The experimental results were statistically significant ( $P < 0.05$ ) taking into account unavoidable errors associated with serial dilutions.

<sup>e</sup> Degree of correspondence between results obtained with the standard plating technique (*M. agalactiae* CFU/25 ml) and those obtained with the *p40* Q-PCR method (*M. agalactiae* genome equivalents/25 ml).

<sup>f</sup> Noncontaminated milk.

detected as few as 250 *M. agalactiae* cells per 25 ml of milk (Table 3). This detection limit corresponds to approximately 1 genomic unit per reaction, similar to the limits obtained if genomic DNA isolated from *M. agalactiae* was used as a PCR template. We then evaluated the use of this method for quantifying *M. agalactiae* cells in sheep milk. We constructed regression curves with the  $C_T$  values obtained with artificially contaminated milk samples and the corresponding numbers of *M. agalactiae* cells used for the inoculation. The overall correlation coefficient ( $R^2 = 0.9989$ ) demonstrated that the Q-PCR assay was linear over a range of 5 logs, down to 2,500 cells/25 ml of milk (Table 3). The PCR efficiencies obtained ( $E = 0.996$ ) indicated that the performance of the Q-PCR assay was excellent. Moreover, these values were similar to those obtained with the analysis of purified *M. agalactiae* genomic DNA.

**Validation of the *p40*-IAC Q-PCR assay for the detection of *M. agalactiae* in naturally contaminated sheep milk samples.** The *p40*-IAC Q-PCR assay was performed with 373 natural raw sheep milk samples from refrigerated tanks from different farms and with 424 milk samples from individual sheep from a flock that tested positive for this pathogen. In every experiment, we included negative controls to verify the accuracy of the results. In parallel, we also tested the samples by using two procedures recommended by the World Organization for Animal Health: microbiological isolation coupled with microscopy observation and a PCR for milk samples developed by Tola and coworkers (27). If it was not possible to process samples within 24 h, they were frozen at  $-20^\circ\text{C}$ , after it was determined whether the mycoplasma cells could survive for at least 3 weeks at  $-20^\circ\text{C}$  without the numbers of cells decreasing. All the samples were cultivated in liquid and on solid

selective media before testing, as follows: mycoplasma broth base with mycoplasma supplement G (Oxoid), a medium specific for this genus; and MA1A, a medium specific for *M. agalactiae* (Mycoplasma Experience, Surrey, United Kingdom). Aliquots of samples were cultivated in *Mycoplasma* broth with *Mycoplasma* supplement G at  $37^\circ\text{C}$  in moist anaerobic chambers in a 5%  $\text{CO}_2$  atmosphere for 3 days. The samples were then vortexed, and 1 ml was transferred into a 1.5-ml Eppendorf tube and centrifuged at  $13,000 \times g$  for 15 min. The supernatant was discarded, and the cell pellet was resuspended in 100  $\mu$ l of 10 mM Tris-HCl for further incubation at  $95^\circ\text{C}$  for 20 min. Finally, the tube was centrifuged at maximum speed for 10 min, and the supernatant was transferred into a new tube. We used 2  $\mu$ l of the lysate for PCR. The DNA was then tested by using the conventional PCR (27) and the *p40*-IAC Q-PCR methods. In parallel, enriched samples from mycoplasma broth were also streaked on agar mycoplasma medium containing supplement G and on MA1A agar medium and cultivated at  $37^\circ\text{C}$  in moist anaerobic chambers in a 5%  $\text{CO}_2$  atmosphere for 6 days. The resulting, typical "fried-egg" *Mycoplasma* colonies were examined by light microscopy at  $10\times$  magnification. In order to confirm that the isolated colonies were *M. agalactiae*, we tested the enrichment broth cultures for the presence of strains of *Mycoplasma* species involved in contagious agalactia in small ruminants (i.e., *M. mycoides* cluster and *M. putrefaciens*), using PCR methods previously published (18, 29).

If the sample tested PCR positive, we then assayed DNA extracted directly from milk without culture (Fig. 1). In 373 tank milk samples, a total of 57 samples tested positive by the *p40*-IAC Q-PCR method after culture (Table 4). Of 57 samples, 31 were identified as *Mycoplasma* positive by microscopy observation (none was PCR positive for the *M. mycoides* clus-

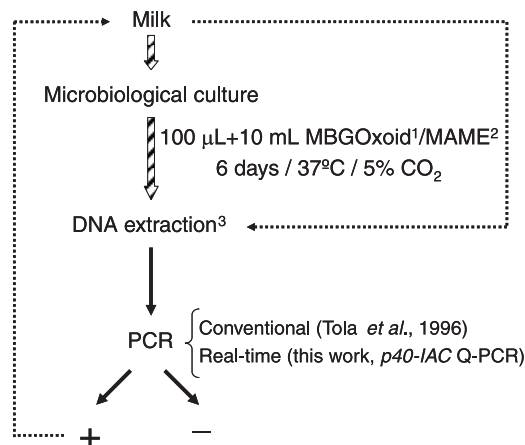


FIG. 1. Raw sheep milk sample analysis scheme. Milk samples (797) were received, and 100  $\mu$ l was cultured in 10 ml of specific microbiological media (1, MBG *Mycoplasma* broth base with supplement G from OXOID; and 2, MA1A medium from Mycoplasma Experience) at  $37^\circ\text{C}$  in 5%  $\text{CO}_2$  chambers for 6 days. DNA was then extracted using thermolysis (3) and analyzed by PCR using the *p40*-IAC Q-PCR assay and the method described by Tola and coworkers (27). If the PCR tested positive, DNA was directly purified from 25 ml of milk to demonstrate direct detection by avoiding cultivation and to improve the analytical performance.

TABLE 4. Detection of *M. agalactiae* in naturally contaminated milk samples<sup>a</sup>

Time	No. of positives samples (%) using <sup>b</sup> :		
	<i>p40</i> -IAC Q-PCR	Conventional PCR <sup>c</sup>	Microscopy <sup>d</sup>
Prior to 6-day culture	44 (11.80)	20 (5.36)	No observation
After 6-day culture	57 (15.28)	36 (9.65)	31 (8.31)

<sup>a</sup> Natural raw sheep milk samples (373) from refrigerated tanks from different farms were investigated using three different methods (*p40*-IAC Q-PCR, conventional PCR [26], and microscopy identification) prior to and after a 6-day culture at 37°C in moist anaerobic chambers in a 5% CO<sub>2</sub> atmosphere.

<sup>b</sup> Data show numbers of positive results obtained per each method for 373 samples tested and percentages of prevalence of *M. agalactiae*, using the three different methods.

<sup>c</sup> Conventional PCR method as published previously by Tola and coworkers (26).

<sup>d</sup> Examination by light microscopy at 10× magnification. Samples were considered positive when typical “fried-egg” colonies were observed.

ter or *M. putrefaciens*, thus confirming that they were only *M. agalactiae*; these samples also tested positive with both of the PCR methods after culture. In addition, five samples, which could not be confirmed by microscopy observation, tested positive with both of the PCR methods. An additional 21 samples tested positive only by our real-time PCR detection system. Thus, the positivity of *M. agalactiae* in 373 milk samples found by the *p40*-IAC Q-PCR method was 15.28% (29 samples had a  $C_T$  value of <25, 19 samples had  $C_T$  values of 25 to 34, and 9 had  $C_T$  values of ≥35) (Table 4). However, the positivity of *M. agalactiae* using microscopy observation or conventional PCR (27) only was 9.65% or 8.31%, respectively. These results indicated that these two procedures (i.e., microscopy observation and the conventional PCR) (27) underestimate the actual presence of *M. agalactiae* in natural samples by almost twofold (1.58 and 1.85 times, respectively) in comparison with our assay. Among the samples extracted directly from milk, 44 samples tested positive by the *p40*-IAC Q-PCR assay developed for this study (all of the samples were also positive after culture) (Table 4). However, the 13 samples not detected prior to incubation exhibited low positive signals when analyzed after culture ( $C_T$  values above 32), suggesting very low *M. agalactiae* numbers in milk after the enrichment and, thus, probably below the limit of detection in the moment of direct bacterial DNA extraction prior to the enrichment. By contrast, conventional PCR (27) detected only 20 positive samples prior to the 6-day cultivation (Table 4). Thus, our *p40*-IAC Q-PCR assay was more sensitive, particularly if DNA was extracted directly from milk. Consequently, *p40*-IAC Q-PCR was capable of detecting realistic levels of *M. agalactiae* contamination in milk samples but was faster and the levels were quantifiable.

We also collected 424 milk samples from individual sheep from a flock for which the corresponding tank tested positive. Only four samples were positive: two exhibited a  $C_T$  value below 25, one sample had  $C_T$  values in the range of 26 to 34, and one sample's  $C_T$  value was above 34. Moreover, all IAC results showed positive amplification in all samples that were negative by the *p40*-IAC Q-PCR method, indicating that the PCRs did not fail. The absence of inhibition in any of the reactions also suggests that the preamplification procedure used is effective in these types of samples. This result provides new insights into the contagious agalactia process in sheep

flocks, since, whereas the refrigerated tank of the sheep farm tested positive, resulting in the belief that the flock was affected by contagious agalactia, only 4 out of 424 animals (0.94%) tested positive.

In conclusion, we describe a reliable and sensitive real-time PCR-based method for quantifying *M. agalactiae* cells in raw sheep milk taken directly from the refrigeration tanks from farms or from animals. Our method incorporates an IAC to assist in the interpretation of the results obtained. This method provides a significant quantification capacity, as defined by its wide dynamic quantification range (at least 6 orders of magnitude), linearity ( $R^2 > 0.9921$ ), PCR efficiency ( $E > 0.992$ ), and quantification limit (down to 10 *M. agalactiae* genome equivalents). If the assay is combined with simple detergent and enzymatic treatments of samples before centrifugation and nucleic acid extraction, it provides a highly effective quantitative analysis for large volumes of milk. It is able to detect 250 cells in 25 ml of raw sheep milk, with an excellent accuracy relative to that of the reference microbiological method. We evaluated and validated the Q-PCR assay's capacity to detect *M. agalactiae* in 797 raw sheep milk samples and showed an average positivity for *M. agalactiae* of 15.28%. This method therefore provides a promising alternative to procedures currently recommended by the World Organization for Animal Health for the rapid, sensitive, and quantitative detection of *M. agalactiae* in milk, which may be easily adopted by animal health laboratories.

This study was supported by research grant RTA2008-00073 and the European Union's Marie-Curie mobility program (contract MERGCT-2007-209050).

K.O. received a National Scholarship of the Slovak Republic Government and a FEMS research fellowship. L.L.-E. received a Ph.D. studentship from the Instituto Nacional de Investigación y Tecnología Agraria y Alimentaria (INIA), and M.H. holds a contract from the INIA, cofunded by the European Social Fund (ESF).

#### REFERENCES

- Al-Soud, W. A., and P. Radstrom. 2001. Purification and characterization of PCR-inhibitory components in blood cells. *J. Clin. Microbiol.* **39**:485–493.
- Bashiruddin, J. B., J. Frey, M. H. Königsson, K. E. Johansson, H. Hotzel, R. Diller, P. de Santis, A. Botelho, R. D. Ayling, R. A. Nicholas, F. Thiaucourt, and K. Sachse. 2005. Evaluation of PCR systems for the identification and differentiation of *Mycoplasma agalactiae* and *Mycoplasma bovis*: a collaborative trial. *Vet. J.* **169**:268–275.
- Bergonier, D., X. Berthelot, and F. Poumarat. 1997. Contagious agalactia of small ruminants: current knowledge concerning epidemiology, diagnosis and control. *Rev. Off. Int. Epizoot.* **16**:848–873.
- Chávez González, Y. R., C. Ros Bascañana, G. Bölske, J. G. Mattsson, C. Fernández Molina, and K. E. Johansson. 1995. In vitro amplification of the 16S rRNA genes from *Mycoplasma bovis* and *Mycoplasma agalactiae* by PCR. *Vet. Microbiol.* **47**:183–190.
- DaMassa, A. J., P. S. Wakenell, and D. L. Brooks. 1992. Mycoplasmas of goats and sheep. *J. Vet. Diagn. Invest.* **4**:101–113.
- Dedieu, L., V. Mady, and P. C. Lefevre. 1995. Development of two PCR assays for the identification of mycoplasmas causing contagious agalactia. *FEMS Microbiol. Lett.* **129**:243–249.
- Director General de Producción Agropecuaria. 2006. Resolución de 24 de mayo de 2007, por la que desarrollan los programas sanitarios a realizar por las Agrupaciones de Defensa Sanitaria Ganaderas, definidos en la Orden AYG/1131/2006, de 30 de junio. Comunidad Autónoma de Castilla y León, Spain.
- Fleury, B., D. Bergonier, X. Berthelot, E. Peterhans, J. Frey, and E. M. Vilei. 2002. Characterization of P40, a cytoadhesin of *Mycoplasma agalactiae*. *Infect. Immun.* **70**:5612–5621.
- Foddai, A., G. Idini, M. Fusco, N. Rosa, C. de la Fe, S. Zinellu, L. Corona, and S. Tola. 2005. Rapid differential diagnosis of *Mycoplasma agalactiae* and *Mycoplasma bovis* based on a multiplex-PCR and a PCR-RFLP. *Mol. Cell. Probes* **19**:207–212.
- Hernández, M., M. Pla, T. Esteve, S. Prat, P. Puigdomènech, and A. Fer-

- rando**. 2003. A specific real-time quantitative PCR detection system for event MON810 in maize YieldGard based on the 3'-transgene integration sequence. *Transgenic Res.* **12**:179–189.
11. **Hoorfar, J., N. Cook, B. Malorny, M. Wagner, D. De Medici, A. Abdulmajid, and P. Fach.** 2004. Diagnostic PCR: making internal amplification control mandatory. *J. Appl. Microbiol.* **96**:221–222.
  12. **Johansson, K. E., M. U. Heldtander, and B. Pettersson.** 1998. Characterization of mycoplasmas by PCR and sequence analysis with universal 16S rDNA primers. *Methods Mol. Biol.* **104**:145–165.
  13. **Königsson, M. H., G. Bölske, and K. E. Johansson.** 2002. Intraspecific variation in the 16S rRNA gene sequences of *Mycoplasma agalactiae* and *Mycoplasma bovis* strains. *Vet. Microbiol.* **85**:209–220.
  14. **López-Enríquez, L., D. Rodríguez-Lázaro, and M. Hernández.** 2007. Quantitative detection of *Clostridium tyrobutyricum* in milk by real-time PCR. *Appl. Environ. Microbiol.* **73**:3747–3751.
  15. **Lorusso, A., N. Decaro, G. Greco, M. Corrente, A. Fasanella, and D. A. Buonavoglia.** 2007. A real-time PCR assay for detection and quantification of *Mycoplasma agalactiae* DNA. *J. Appl. Microbiol.* **103**:918–923.
  16. **Madanat, D., Z. Zendulková, and I. L. Pospí.** 2001. Contagious agalactia of sheep and goats: a review. *Acta Vet. Brno* **70**:403–412.
  17. **Nicolas, R. A. J.** 2004. Contagious agalactia, p. 607–614. *In* OIE manual of diagnostic tests and vaccines for terrestrial animals (mammals, birds and bees), vol. 2. Paris, France.
  18. **Peyraud, A., S. Woubit, J. B. Poveda, C. De la Fe, P. Mercier, and F. Thiaucourt.** 2003. A specific PCR for the detection of *Mycoplasma putrefaciens*, one of the agents of the contagious agalactia syndrome of goats. *Mol. Cell. Probes* **17**:289–294.
  19. **Rodríguez-Lázaro, D., M. Hernández, T. Esteve, J. Hoorfar, and M. Pla.** 2003. A rapid and direct real time PCR-based method for identification of *Salmonella* spp. *J. Microbiol. Methods* **54**:381–390.
  20. **Rodríguez-Lázaro, D., M. Hernández, M. Scortti, T. Esteve, J. A. Vázquez-Boland, and M. Pla.** 2004. Quantitative detection of *Listeria monocytogenes* and *Listeria innocua* by real-time PCR: assessment of *hly*, *iap*, and *lin02483* targets and AmpliFluor technology. *Appl. Environ. Microbiol.* **70**:1366–1377.
  21. **Rodríguez-Lázaro, D., M. D'Agostino, A. Herrewegh, M. Pla, N. Cook, and J. Ikonomopoulos.** 2005. Real-time PCR-based methods for quantitative detection of *Mycobacterium avium* subsp. *paratuberculosis* in water and milk. *Int. J. Food Microbiol.* **101**:93–104.
  22. **Rodríguez-Lázaro, D., M. Pla, M. Scortti, H. J. Monzó, and J. A. Vázquez-Boland.** 2005. A novel real-time PCR for *Listeria monocytogenes* that monitors analytical performance via an internal amplification control. *Appl. Environ. Microbiol.* **71**:9008–9012.
  23. **Rodríguez-Lázaro, D., D. A. Lewis, A. A. Ocampo-Sosa, U. Fogarty, L. Makrai, J. Navas, M. Scortti, M. Hernández, and J. A. Vázquez-Boland.** 2006. An internally controlled real-time PCR method for quantitative species-specific detection and *vapA* genotyping of *Rhodococcus equi*. *Appl. Environ. Microbiol.* **72**:4256–4263.
  24. **Rodríguez-Lázaro, D., B. Lombard, H. V. Smith, A. Rzezutka, M. D'Agostino, R. Helmuth, A. Schroeter, B. Malorny, A. Miko, B. Guerra, J. Davison, A. Kobilinsky, M. Hernández, Y. Bertheau, and N. Cook.** 2007. Trends in analytical methodology in food safety and quality: monitoring microorganisms and genetically modified organisms. *Trends Food Sci. Technol.* **18**:306–319.
  25. **Sirand-Pugnet, P., C. Lartigues, M. Marena, D. Jacob, A. Barre, V. Barbe, C. Schenowitz, S. Mangenot, A. Couloux, B. Segurens, A. de Daruvar, A. Blanchard, and C. Citti.** 2007. Being pathogenic, plastic, and sexual while living with a nearly minimal bacterial genome. *PLoS Genet.* **3**:E75.
  26. **Subramaniam, S., D. Bergonier, F. Poumarat, S. Capaul, Y. Schlatter, J. Nicolet, and J. Frey.** 1998. Species identification of *Mycoplasma bovis* and *Mycoplasma agalactiae* based on the *uvrC* genes by PCR. *Mol. Cell. Probes* **12**:161–169.
  27. **Tola, S., G. Idini, D. Manunta, G. Galleri, A. Angioi, A. M. Rocchigiani, and G. Leori.** 1996. Rapid and specific detection of *Mycoplasma agalactiae* by polymerase chain reaction. *Vet. Microbiol.* **51**:77–84.
  28. **Tola, S., A. Angioi, A. M. Rocchigiani, G. Idini, D. Manunta, G. Galleri, and G. Leori.** 1997. Detection of *Mycoplasma agalactiae* in sheep milk samples by polymerase chain reaction. *Vet. Microbiol.* **54**:17–22.
  29. **Woubit, S., L. Manso-Silván, S. Lorenzon, P. Gaurivau, F. Poumarat, M. P. Pellet, V. P. Singh, and F. Thiaucourt.** 2007. A PCR for the detection of mycoplasmas belonging to the *Mycoplasma mycoides* cluster: application to the diagnosis of contagious agalactia. *Mol. Cell. Probes* **21**:391–399.