MINI REVIEW

Trends in the design and use of elastin-like recombinamers as biomaterials

Arturo Ibáñez-Fonseca, Tatjana Flora, Sergio Acosta, José Carlos Rodríguez-Cabello*

BIOFORGE Lab, CIBER-BBN, University of Valladolid. Paseo de Belén 19, 47011 – Valladolid, Spain

*Corresponding author

Prof. J.C. Rodríguez-Cabello. Edificio LUCIA, Paseo de Belén, 19, 47011 – Valladolid (SPAIN). Phone: +34983184799. E-mail: roca@bioforge.uva.es.

Keywords: biomaterials, elastin-like recombinamers, intrinsically disordered proteins, drug delivery, tissue engineering

Abstract

Elastin-like recombinamers (ELRs), which derive from one of the repetitive domains found in natural elastin, have been intensively studied in the last few years from several points of view. In this mini review, we discuss all the recent works related to the investigation of ELRs, starting with those that define these polypeptides as model intrinsically disordered proteins or regions (IDPs or IDRs) and its relevance for some biomedical applications. Furthermore, we summarize the current knowledge on the development of drug, vaccine and gene delivery systems based on ELRs, while also emphasizing the multiple tissue engineering approaches involving their use. Finally, we show different studies that explore applications in other fields, and several examples that describe biomaterial blends in which ELRs have a key role. This review aims to give an overview of the recent advances regarding ELRs and to encourage further investigation of their properties and applications.

Introduction

As described in previous sections of this special issue, elastin is one of the main components of the extracellular matrix (ECM), and it is involved in conferring elasticity to a variety of organs and tissues, such as lungs, skin and blood vessels, among others, while also contributing to cell signaling [1]. Formed by the lysyl oxidase-mediated crosslinking of lysine residues present in its soluble precursor, tropoelastin, it is composed of highly repetitive and well-conserved domains, including the hydrophobic Val-Pro-Gly-Val-Gly (VPGVG) pentapeptide, first described by Gray et al. [2]. Soon after that, Urry's laboratory became interested in the synthesis of this motif to investigate the features of elastin in a feasible way, in order to shed light on the pathophysiology of several diseases in which elastin is directly involved, hence developing the first elastin-like (poly)pentapeptides (ELPs) [3-5]. These ELPs showed an inverse temperature transition, meaning that they remained soluble below the so-called transition temperature (T_i) , and they aggregated when the temperature was raised above approximately 25°C. Moreover, physicochemical studies concluded that changes in the fourth amino acid of the pentapeptide led to alterations in the T_t , giving flexibility to the ELP design, since the basal monomer became VPGXG, where X (the guest residue) can be any amino acid except proline [6]. However, one of the major limitations at this point was that these primitive ELPs were chemically synthesized, hindering the achievement of long polypeptides. This issue was addressed with the advent of the recombinant DNA technology during the nineteen-eighties, which was easily adapted for the expression of structural protein polymers in heterologous hosts, mainly Escherichia coli [7-10] (Fig.

1). It also increased the versatility of the ELPs, allowing the combination of different structural protein domains [8], or the inclusion of bioactive amino acid sequences [11]. Furthermore, the polymeric and recombinant nature of these ELPs led to a new nomenclature proposed by Rodríguez-Cabello *et al.*, elastin-like recombinamers (ELRs), in order to recapitulate both features in a single term [12].

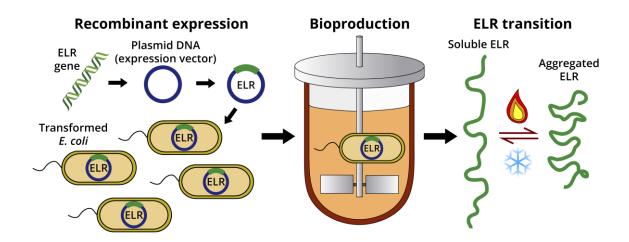


Fig. 1. Schematic representation of the recombinant synthesis of ELRs and their reversible transition above the T_{t} .

In the last years, a significant effort has been made to achieve novel ELR designs to extend the fields of application of this biopolymer. In this regard, several approaches for a comprehensive ELR development have been described, including T_t prediction [13] and the study of the effect of proteins fused to the ELRs in its recombinant expression [14] and self-assembly [15]. Moreover, different computational methods have been validated to elucidate the molecular mechanisms involved in the transition from a soluble to an aggregated state (above the T_t) of ELRs with different compositions, which may help in the anticipation of the physicochemical properties of novel recombinanters [16-19].

In this review, our aim is to give an overview of the most 'trendy' strategies described in the literature regarding the design and use of ELRs as biomaterials in different fields. First, we will discuss works describing how ELRs can be considered model intrinsically disordered proteins (IDPs) and how they can help in the general study of this class of polypeptides. Furthermore, we will comment the diverse applications inferred from the control of their ordered-disordered state. Moreover, while several publications have already reviewed the use of ELRs in drug delivery [20, 21] and in tissue engineering and regenerative medicine (TERM) [1, 22-24], herein we will point out the major advances in the multi-purpose design and use of ELRs in these fields in the past few years. Finally, we will describe other designs and applications that do not completely fit into the aforementioned sections.

ELRs as model intrinsically disordered proteins

Traditionally, protein functionality was associated with the ability of polypeptide chains to fold into 3D structures, following the classical structure-function paradigm [25]. However, during the last decades, the increasing number of structural studies have shed light on the functional key role of intrinsically disordered regions (IDRs) and proteins (IDPs), which represent 40% of the eukaryotic proteome [26]. Furthermore, they are characterized by low complexity sequences, mainly hydrophilic, high content in proline and they are usually found in an unfolded state [27]. However, their flexibility enables them to interact with several different targets and to respond to different stimulus, folding into diverse dynamic molecular assemblies [28, 29], which may undergo phase transitions [30]. Thus, IDRs have been found to play a crucial role in cellular communication and gene regulation [31]. Moreover, mutations in their sequences are involved in the development of several human pathologies [32].

From the structural point of view, some protein polymers, such as ELRs or resilin-like polypeptides, fit into the description of IDRs and IDPs, i.e. they are made of the repetition

of low-complexity sequences with high content in proline and glycine [33], and they undergo phase transitions under selective solution conditions, such as specific temperature, pressure or pH [34]. Therefore, since a few years ago, they have been considered as model IDRs or IDPs, and studied accordingly [35, 36]. In addition, their recombinant nature and the feasibility to tune their sequence make them perfect candidates to mimic complex IDRs and study their function and behavior in solution [36, 37]. For instance, depending on the chemical nature of the guest residue of the elastinlike pentapeptide (VPGXG), we can control the propensity of the sequence to be disordered or, on the contrary, to collapse, thus shedding light on the phase separation process of IDRs.

Regarding the evaluation of the physicochemical properties of ELRs, Zhang *et al.* provided an experimental structural model for the study of the early stages of the phase separation behavior [38]. A 40-pentapeptide ELR suitable for nuclear resonance measurements was developed based on a polyVPGVG that contained a different guest residue every seven pentapeptides (Lys, Thr, Ala, Ile, Ser and Leu). Therefore, it was possible to analyze the influence of every single non-valyl amino acid as guest residue (all the aforementioned except for Ile) on the phase transition. By complementing the chemical characterization with computational simulations, they demonstrated that a monomeric ELR behaved as random coils with a small compaction, even if the structure was folded up to 90% into β -turns. Consistently with previous results [39, 40], they suggested that oligomer coacervation is triggered by the temperature-dependent hydrophobic collapse and that, despite the formation of transient β -turns during this process, the intrinsic disorder of the monomers is maintained in this aggregated state.

Furthermore, many IDRs are involved in the formation of subcellular protein compartments due to their phase separation behavior [41], which implies that ELRs may

be used as a model to form and study membraneless organelles. In this way, ELR collapse and phase separation allowed to develop subcellular compartments in mammal cells, controlling the clathrin-mediated endocytosis with an ELR fused to clathrin light chain [42], and to produce membraneless organelle-like structures in bacteria with a modular design based on an amphiphilic ELR [43]. This ELR was also used to produce artificial subcellular compartments based on nanovesicles that were able to encapsulate cellular reactions and processes such as transcription and translation [44]. The protein membranes of the vesicles were non-permeable to small molecules, allowing the specific separation of both processes. Moreover, their dynamic behavior and their ability to grow and generate protocellular compartments was demonstrated. In addition, even the amphiphilic ELR itself was translated inside the vesicles, and the monomers were incorporated within the vesicular membrane upon synthesis.

Applications derived from the order-disorder balance in ELRs

The control of disorder in ELRs may play a significant role for their use in biomedical applications. For instance, several studies have highlighted the contribution of scaffolds made of disordered proteins in the biomineralization process [45-47]. In this regard, ELRs have demonstrated their promising potential for the development of a great variety of structures that control mineralization due to their flexibility to self-assemble into diverse conformations. Hence, nanoparticles [48], 3D matrices [49, 50], and even nanotopographical hybrid surfaces that enable enzyme-directed mineralization [51] have been achieved, all of them mimicking the mechanical properties of native hard tissues. However, it is important to emphasize how the balance between order and disorder in ELR scaffolds influences the mineralization process. Li *et al.* developed bone fibrils made of ELRs that mimic collagen ones and demonstrated that their ordered structure is crucial for intrafibrillar mineralization [52]. In fact, the incorporation of short charged sequences

within the ELR monomers seemed to prevent their folding into ordered β -spiral structures, predetermining, in this way, the effectiveness of the mineralization via a polymer-induced liquid-precursor (PILP) process, which implied that the decrease in the order of the fibrillar microstructures involved a reduction of the mineral density. Similarly, Elsharkawy *et al.* produced mineralized ELR-based membranes with potential applicability for enamel regeneration [53]. The membranes provided a functional acid-resistant scaffold for the nucleation and growth of hydroxyapatite. In addition, it was possible to control and tune the formation of hierarchical mineralized structures and their mechanical properties through the regulation of the crosslinking degree of the ELR molecules within the membranes, which influenced the order-disorder balance.

The order-disorder equilibrium is also important to adjust the mechanical properties of injectable hydrogel scaffolds intended for tissue regeneration. In a recent work, Roberts *et al.* developed a molecular design based on alternating ordered polyalanine motifs with intrinsically disordered ELR domains to produce injectable porous scaffolds [54]. In this work, the authors showed that the interactions between the polyalanine α -helixes enabled the formation of kinetically stable 3D polypeptide networks, improving their mechanical properties in a similar way to the introduction of other ordered domains that form stable links/bonds and stabilize non-covalently cross-linked hydrogels, such as leucine zippers [55]. Moreover, they studied the modular distribution of the α -helix structures and demonstrated that their introduction within the ELR backbone strongly affects phase separation and hydrogel porosity, observing thermal hysteresis in the phase transition behavior. Therefore, the fine-tuning of the ratio between both domains and their composition provided a way to control the aggregation temperature and the mechanical properties of the ELR-based scaffolds.

The modular design and functional versatility of ELRs open up a range of possibilities for the development of covalent coatings for indwelling biomedical devices. In this way, their biocompatibility can be improved through the addition of different biofunctionalities, in order to avoid the failure of the biomaterial upon implantation, mainly by rejection (foreign body response) or infection (Fig. 2). In this sense, ELR coatings provide an ECM-like environment that helps to elude unspecific protein adsorption, and that can be tuned to increase the cytocompatibility of the devices, for example, through the inclusion of cell adhesion sequences [56, 57]. Furthermore, it could be possible to produce ELR patterns with diverse biofunctional domains to control the adhesion of different types of cells on different areas of the coating [58, 59]. Moreover, biomimetic ELR coatings for biomedical devices can be combined with antimicrobial peptides (AMPs), which are immunomodulatory short cationic peptides with broadspectrum antimicrobial activity and constitute one of the most promising alternatives to overcome bacteria resistance to conventional antibiotics [60]. This strategy has recently been described for the development of ELRs with covalently linked AMPs to produce anti-biofilm coatings for titanium implants, enhancing osteogenic differentiation and preventing the colonization of the devices by pathogenic bacteria [61]. In addition, ELRs provide a scalable method for the production of novel antimicrobial materials by incorporating AMPs into their backbone through recombinant DNA technology, thus overcoming the expensive chemical production of AMPs, which hampers their largescale production. In this regard, ELRs have demonstrated their potential for recombinant production and non-chromatographic purification of a broad range of proteins [62], which may include AMPs, and they can also boost the antimicrobial effect of these peptides when used in the formation of anti-biofilm coatings. In a still unpublished work, we have found that the low-fouling properties of the ELRs, which prevent unspecific protein

adsorption, converge synergistically with the antimicrobial properties of an AMP when both are recombinantly co-produced (Acosta *et al.*, in press). Lastly, ELRs have been used to control the supramolecular assembly of AMPs to enhance their effectiveness. For instance, an AMP-ELR able to self-organize into different structures has been described, allowing the formation of nanoparticles that allow a controlled delivery [63] or films that inhibit bacterial and fungal infection during wound healing [64].

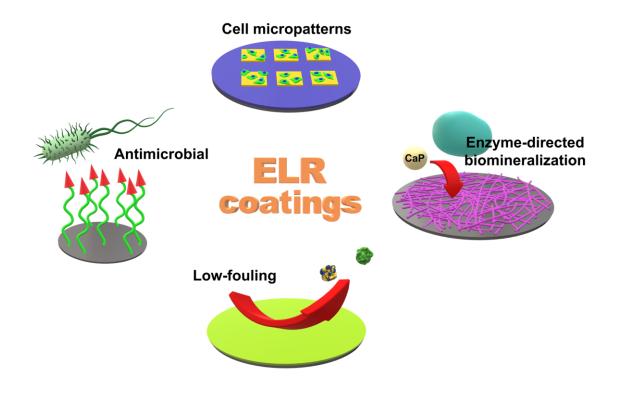


Fig. 2. Schematic illustration of different biomaterial surfaces functionalized with ELR coatings and their properties.

Drug, vaccine and gene delivery systems based on ELRs

ELRs have been used in the last decades for the development of drug delivery systems (DDS) by following different approaches, mainly the recombinant conjugation of pharmacological polypeptides and the use of drugs as cargo of nanoparticles (NPs) and/or

hydrogels. Moreover, within this section we will also consider immunomodulatory ELRs that act as vaccines, and the use of these recombinamers as gene delivery vectors.

ELRs in drug delivery

As aforementioned, recombinant DNA technology permits the genetic fusion of peptides or proteins to the ELRs, giving the opportunity of improving their functionality to almost an infinite extent. By these means, interferon alpha (IFN- α) was fused to an ELR, which not only simplified its purification by inverse transition cycling (ITC), but also improved the pharmacokinetics and tumor accumulation of the drug by increasing its circulating half-life [65]. Similarly, stromal cell-derived growth factor-1 (SDF1), which promotes vascularization, hence enhancing and accelerating re-epithelialization of skin wounds, was fused to an ELR, resulting in a faster wound healing in a diabetic mice model in comparison with the SDF1 by itself [66]. Another example of pharmacologically active ELRs implied the fusion of the fibroblast growth factor 21 (FGF21), a metabolic regulator that enhances insulin sensitivity, thus being a potential treatment for type 2 diabetes [67]. This fusion improved the solubility of the protein drug in E. coli, hence avoiding further refolding steps needed when inclusion bodies are formed during expression of FGF21 in this host. Moreover, the delivery of the ELR-FGF21 increased the half-life of the growth factor, so fewer injections were needed for a sustained pharmacological effect. An alternative to the fusion protein drugs is the use of peptides, whose activity does not usually require a complex folding, so they can be easily expressed in prokaryotic hosts such as E. coli. In this regard, the so-called 'mini cry' peptide, derived from the intravitreal αB crystallin protein, which was found to protect retinal pigment epithelium (RPE) cells from oxidative stress-induced cell death, was genetically conjugated to an ELR to improve the retention of the bioactive peptide from 0.4 to 3.0 days [68]. According to this result, the ELR-mini cry was proposed as a potential DDS to prevent RPE atrophy and progressive age-related macular degeneration. To further increase the circulation time in plasma of ELRs and ELR particles, novel zwitterionic polypeptides (ZIPP) were designed by introducing cationic and anionic amino acids within the same elastin-like pentapeptide [69]. The results showed that the combination of lysine and glutamic acid improved pharmacokinetics, an effect that was successfully leveraged to obtain a more sustainable delivery of glucagon-like peptide-1, used for the treatment of type 2 diabetes, when fused to the ZIPP.

Another different but classical approach, which is also more versatile because other than protein drugs can be delivered, is the loading of nanoparticles or hydrogels with pharmacologically active compounds. In this last regard, it has been reported the formation of elastin-like (EL) and silk-elastin-like (SEL) hydrogels loaded with timolol maleate (TM), a drug used in the treatment of glaucoma to reduce the intraocular pressure [70]. The retention of TM was improved without eye irritation, an effect that was even more marked for SEL hydrogels, due to the higher stability obtained by the cross-linking of silk-like domains [71]. Therefore, both EL and SEL hydrogels were proposed as delivery systems in ophthalmic applications. As concerns the use of ELR particles as DDS, some design improvements have been achieved in the past few years to overcome different limitations, such as low plasma half-life, similarly to the aforementioned ZIPPs. For instance, an albumin-binding domain (ABD) was fused to an ELR, and the wellknown antitumor drug doxorubicin (DOX) was conjugated to the resulting ABD-ELR [72]. The whole system was found to self-assemble into spherical micelles, and the delivery of DOX with the ABD-containing particles resulted in a 3-fold increase in plasma half-life compared to the naked ELR micelles. Moreover, the ABD-ELR-DOX nanoparticles demonstrated higher uptake by the tumor, hence reducing toxicity in other organs, like liver or spleen, and achieving a therapeutic effect with lower DOX doses.

Another issue when targeting tumors is the low uptake of drug-loaded nanoparticles by cells, thus limiting the activity of chemotherapeutic agents. In order to address this limitation, van Oppen *et al.* described the fusion of an octa-arginine peptide (R8) to an amphiphilic ELR, which promoted the uptake of the micelles by the cells *in vitro* [73]. This result sets the basis for the use of this R8-ELR for the delivery of antitumor drugs, although this work lacks evidence of *in vivo* performance. On the other hand, retention of delivery systems in mucus is a major challenge that was recently overcome by genetic conjugation of a hydrogel-forming ELR with a mucoadhesive peptide able to bind to transferrin receptors in the epithelial cell layer [74]. This DDS can be potentially used towards different *mucosae*, as demonstrated by *in vitro* and *in vivo* results.

ELR-based vaccines

Vaccination has also been explored as an opportunity to extend the applications of ELRs in the biomedical field. Specifically, García-Arévalo *et al.* described the fusion of an antigenic sequence from a major membrane protein of *Mycobacterium tuberculosis* to an amphiphilic ELR able to form stable nanoparticles [75]. *In vivo* immune-challenge experiments in mice showed the induction of a response similar to the one found upon vaccination, also highlighting the adjuvant effect of the ELR by itself. Likewise, Ingrole *et al.* designed an ELR fused to the M2e peptide, derived from the highly conserved extracellular domain of the influenza virus transmembrane protein [76]. This construction induced the production of a higher amount of M2e-specific serum antibodies in mice in comparison with the single M2e, thus enhancing the immunogenicity of the antigen. Another strategy has been recently reported, involving the fusion of an ELR to cytotoxic T lymphocyte (CTL) epitopes, which induce the activation of T cells and can be used for the treatment of many different diseases, including cancer. This fusion polypeptide allowed the binding of CTL epitopes to dendritic cells (DCs) directly *in vivo*, avoiding

the steps required for the extraction and CTL-loading of DCs *ex vivo* [77]. By also fusing a matrix metalloproteinase (MMP)-sensitive peptide and an albumin-binding domain, the accumulation of the carrier in lymph nodes was increased by 4-fold and the immune response towards the CTL was improved by 1.5-fold.

ELR-mediated gene delivery

Regarding gene delivery, few examples have arisen involving the use of ELRs as transfection agents to substitute other chemical methods and viral vectors, which raise some safety concerns. In a first work, Piña *et al.* described the use of a cationic ELR fused to the LAEL fusogenic peptide, which improves cellular uptake, to form polyplexes with plasmid DNA (pDNA) [78], resulting in an enhanced transfection efficiency compared to the pDNA by itself. In a subsequent study, they conjugated the ELR to an aptamer (short oligonucleotides that can bind molecules with high affinity and selectivity) towards MUC1, a membrane glycoprotein overexpressed in epithelial tumors, such as breast cancer [79]. Therefore, this strategy permitted the specific targeting of breast cancer cells for transfection with toxic agents able to kill tumor cells. Another recent example showed the generation of induced pluripotent stem cells by delivering the four factors described by Yamanaka for such purpose, i.e. Oct-4, Klf4, c-myc and Sox2, with an ELR-based transfection method, thus avoiding viral vectors [80]. Nevertheless, the aforementioned examples need to overcome some issues, such as lower transfection efficiency and higher cost than the widely used polyethylenimine (PEI).

ELRs as biomaterials for tissue engineering and regenerative medicine (TERM)

ELR hydrogels

In the last years, ELRs have been used as biomaterials for the development of hydrogels formed through different cross-linking methods. Specifically, our group has described the covalent cross-linking of ELRs by catalyst-free (i.e. copper-free) click chemistry [81], termed 1,3-dipolar cycloaddition, which is basically an azide-alkyne cycloaddition, taking advantage of the modification of amine groups present in the side chains of lysines. These hydrogels are very promising, since they have shown good *in vitro* and *in vivo* biocompatibility [82]. Macroporous scaffolds have been also achieved with this strategy, expanding the use of these 'click' hydrogels to applications where a highly porous structure is needed [83]. Similarly, other covalent cross-linking methods have been developed, using residues other than lysine for the cross-linking, such as tyrosine or cysteine [84, 85]. On the other hand, non-covalently cross-linked ELR hydrogels have also been developed [55, 71, 86]. All these hydrogels are good candidates as implants for enhanced tissue regeneration, considering the properties of the ELRs, and some of them have already been used for this purpose, as described below.

ELRs for enhancing implant vascularization

One of the major challenges concerning hydrogels and other tissue-engineered constructs is that they need to be vascularized to achieve a successful grafting upon implantation. The lack of vascularization implies an inability to provide adequate nutrient transport during the initial phase after implantation, due to a lack of blood flow between the construct and the host tissue, thus precluding the survival of implanted and host cells inside the scaffold [87, 88]. Therefore, novel strategies for enhancing vascularization in engineered templates are essential to ensure their clinical translation [89]. Particularly, ELR-based hydrogels, whose application in tissue engineering and regenerative medicine has significantly increased in the past few years, are characterized by potential angiogenic properties, which is an essential step for their integration with the host tissue when implanted or injected. Recently, Marsano and co-workers have studied the angiogenic activity of ELR-based hydrogels that included cell adhesion sequences such as the Arg-Gly-Asp (RGD) tri-peptide, which promotes the attachment of different cell types [90], and Arg-Glu-Asp-Val (REDV), a selective cell adhesion motif for endothelial cells [91]. These hydrogels also comprised proteolytic sites, namely the Val-Gly-Val-Ala-Pro-Gly (VGVAPG) sequence, sensitive to elastolytic enzymes [92]. Overall, this study demonstrated the control of angiogenesis by modulating the presence of bioactive sequences within ELR-based hydrogels, as they favor the integration with the host tissue, as well as the beginning of the vascularization process [93]. Similarly, Alagoz et al. coated poly(3-hydroxybutyrate-co-3-hydroxyvalerate) (PHBV) scaffolds with ELRs containing REDV sequence, aiming for bone regeneration. This work shows that the presence of REDV motifs improves endothelial cell adhesion and growth in vitro, with a consequent enhancement of vascularization, which is an important event when facing bone regeneration [94]. As described above, Roberts et al. developed a complex system that consists in the combination of stimuli-responsive disordered ELRs and structurally stable polyalanine helices [54]. When injected subcutaneously in C57BL/6 mice, this system formed a stable porous scaffold that was invaded by a high quantity of cells. This was identified as a mild inflammatory response at the first timepoints, but it was followed by a uniform vascularization, as observed by the formation of capillaries and some large vessels after 21 days post-injection. Hence, their ability to promote angiogenesis makes them optimal candidates for diverse applications in regenerative medicine.

An alternative approach to enhance vascularization in tissue engineering constructs relies on the use of growth factors that induce vascular organization and remodeling. Among them, the vascular endothelial growth factor (VEGF) plays a key role in the majority of angiogenic processes, namely by promoting endothelial cell proliferation and migration [95]. However, the use of VEGF is limited by the need of recombinant expression in eukaryotic cells, mainly mammalian, for its production, and by its low stability in solution. Thus, to overcome these limitations, new growth factor-mimetic peptides have been identified, which can be easily synthesized and chemically tethered to the constructs. For instance, Cai et al. conjugated the so-called QK peptide (a VEGF-mimetic peptide) within ELR hydrogels, without affecting their mechanical properties [96]. Different concentrations of this peptide, i.e. 10 nM, 1 µM and 100 µM, were studied using human umbilical vein endothelial cells (HUVECs) in vitro, demonstrating that, at low concentrations, the QK peptide promoted better cell adhesion and proliferation than higher concentrations, which inhibited their outgrowth. In a subsequent experiment, the QK peptide was tethered to ELR molecules at a concentration of 1 µM and they were injected intramuscularly in a hind limb zone in mice to form hydrogels in situ [97]. The results showed that the QK peptide enhanced the *de novo* formation of functional capillaries within the ELR construct, favoring cell survival and tissue growth in vivo. In summary, these studies provide an alternative approach to the use of growth factor proteins, such as VEGF, in angiogenesis signalling.

Another important feature of engineered constructs that also affects vascularization is their biodegradation, which is known to regulate many cellular behaviors. Biomaterials with a controlled and predictable biodegradation are in high demand, and they should simultaneously provide mechanical support, biological signals and resist physiological loads during the early stages of implantation. Madl *et al.* demonstrated that the formation of vascular-like structures depends on the number of cleavage sites in a polymer sequence, as well as on the changes of the construct architecture upon biodegradation [98]. In addition, protease-mediated degradation of biomaterials plays an important role in the development of 3D systems intended for tissue engineering, as it provides a more precise control on cell infiltration (Fig. 3). Recently, Straley et al. demonstrated that the kinetics and sensitivity to proteolytic cleavage of urokinase plasminogen activator (uPA) epitopes are different depending on the amino acid sequence [99]. For instance, the Gly-Thr-Ala-Arg (GTAR) sequence shows a high sensitivity to proteolytic cleavage, giving a fast response, while the Asp-Arg-Ile-Arg (DRIR) sequence confers a low cleavage efficiency, resulting in slower degradation kinetics. In another related work, Flora et al. evaluated the spatiotemporal control of cell infiltration in a 3D hydrogel that consists of a sandwich-like three-layer disc made of two different protease-sensitive ELRs that included the aforementioned proteolytic sequences (Flora et al., in press). This 3D construct was implanted subcutaneously in mice and the spatiotemporal progression of cell invasion was studied for twelve weeks. It was observed that cell infiltration progressed through an inside-to-outside pattern, meaning that the central layer of the 3D system, which is formed by the GTAR-ELR (characterized by a fast degradation rate), was first colonized, degraded and vascularized. Subsequently, the external layers made of DRIR-ELR (slow degradation rate) were invaded and degraded mainly by the cells that migrated from the inner part of the construct. This study offers new opportunities for the generation of tunable biodegradable systems that closely mimic complex biological structures, such as organoids or organs, and that can be implemented in tissue engineering and regenerative medicine.

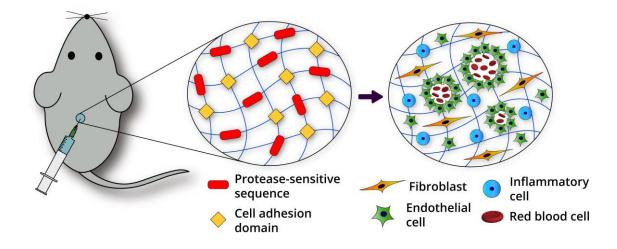


Fig. 3. Graphical representation of ELR-based hydrogels with protease-sensitive and cell adhesion domains injected subcutaneously in mice. Due to their bioactivity, cells would be able to infiltrate over time, even forming vessel-like structures, which is indicative of angiogenesis.

TERM applications of ELR hydrogels

ELR-based hydrogels have been used in cardiovascular applications, taking advantage of their features, such as stable mechanical properties under physiological pressure and flow conditions, elasticity, biocompatibility and hemocompatibility, which make them good candidates in this field [100]. For instance, Gonzalez de Torre *et al.* covered metal stents with ELR-catalyst-free click gels that presented different bioactive sequences, promoting endothelialisation (formation of native endothelium) in less than 2 weeks *in vitro* [101]. When exposed to blood flow in dynamic conditions, minimal platelet adhesion and fibrinogen adsorption were detected on the surface of the ELR-covered stents, showing high hemocompatibility. Thus, this method represents an effective approach to obtain biostents that prevent the formation of an atherosclerotic plaque. Moreover, different ELRs have been used to fabricate vascular grafts. Specifically, Mahara *et al.* developed a small-calibre blood vessel made of ELR-based hydrogels reinforced with poly(lactic acid)

nanofibers as a new therapeutic strategy for reconstructive surgery [102]. When implanted *in vivo*, the tubular scaffold showed a rapid tissue regeneration with a high patency, maintaining a physiologic blood flow without thrombogenicity. Furthermore, Inostroza-Brito *et al.* fabricated geometrically complex structures, namely tubes, through spatiotemporally controlled self-assembly. For this purpose, peptide amphiphiles (PAs) were employed to guide the assembly of a larger protein, i.e. an ELR, into self-growing tubes. These structures promoted mouse-adipose-derived stem cells (mADSCs) and HUVECs adhesion and proliferation, due to the presence of bioactive RGD motifs coded within the ELR backbone. Therefore, these biomimetic tubes could be implemented in cardiovascular tissue engineering for the achievement of vascular implants [103].

Musculoskeletal tissues have also been proposed as targets for ELR hydrogels-mediated regeneration. In this regard, some *in vitro* tests have suggested the potential application of cartilage with a silk-elastin-like recombinamer [104], and bone with pro-mineralizing ELRs [105, 106]. Moreover, a non-covalently cross-linked bioactive ELR-based hydrogel, in combination with human mesenchymal stem cells, has been used to successfully regenerate an osteochondral defect in rabbits, showing even the formation of hyaline-like cartilage [107]. On the other hand, bone defects have been also treated with bioactive and biodegradable ELR-based hydrogels formed with an ELR fused to the bone morphogenetic protein-2 (BMP-2), which is a very powerful osteogenic factor. In this case, fully repaired defects were found after 3 months, showing the good performance of the hydrogels to promote this regeneration [108].

ELRs have also aimed for the regeneration of neural tissue, although short steps have been taken towards it up until now. In one work, Johnson *et al.* genetically fused two different neurotrophins to an ELR, separately: the nerve growth factor (NGF) and the brain-derived neurotrophic factor (BDNF), which could be potentially used for the stimulation of neural regeneration both *in vitro* and *in vivo* [109]. In another work, S.C. Heilshorn and co-workers found that the maintenance of stemness in neural progenitor cells needs a scaffold (i.e. a hydrogel) that is able to be biodegraded over time, simulating matrix remodeling [110].

Although several elastin-based materials, composed of tropoelastin or α -elastin, among other versions of elastin, have been designed for wound healing applications, not so many ELRs have been postulated in this regard [24]. Recently, our group obtained oriented electrospun clickable ELR fibers, which have proven good cytocompatibility with fibroblasts and keratinocytes *in vitro*, suggesting their potential application in the formation of artificial skin or to promote skin regeneration [111]. Contemporarily, Fernández-Colino *et al.* achieved similar fibers, which highlights their ease of attainment and their potential application in different tissue engineering fields [112].

Other designs and applications of ELRs

In addition to the aforementioned designs and applications of ELRs, other interesting uses have been explored in the last years. For instance, there are different works describing the design of ELRs for antibody precipitation as an alternative to chromatography [113, 114]. Moreover, hydrogels containing ELRs have been developed to study and modulate cell behavior in 3D cultures through the incorporation of different ligand chemistries or by tuning the stiffness of the scaffolds [115, 116]. On the other hand, fluorescent proteins have been fused to ELRs, enabling the study and prediction of self-assembly of diverse ELR constructs when genetically conjugated to large proteins [117], or even being proposed as potential Förster resonance energy transfer (FRET)-paired biosensors [118]. Another novel design proved the use of an ELR as a cytocompatible underwater adhesive intended for biomedical applications by the incorporation of the non-canonical amino

20

acid 3,4-dihydroxyphenylalanine (DOPA), which mimics the adhesion of mussels to surfaces [119]. Additionally, Yang *et al.* described how the genetic incorporation of the self-assembling peptide RADA-16 led to the obtaining of a hemostatic sponge able to stop the bleeding of wounds in mice [120].

Furthermore, it is also noteworthy the increasing number of works describing the combination of ELRs with other polymers or structural proteins, giving blends or biohybrids that recapitulate the properties of all the components. The first, and probably the most exploited blend, came from the recombinant fusion of ELRs with silk-like domains to give silk-elastin-like recombinamers (SELRs) that are able to form fibers and hydrogels, among other structures, whose stability relies on the β -sheet cross-linking between silk-like motifs [8, 71, 121-123]. Collagen-like peptides have also been explored as partners of ELRs in a recombinant way [124], even achieving nanoplatelets [125]. Furthermore, other non-polypeptide polymers have been combined with ELRs, including the widely used polyethylene glycol (PEG) [126, 127], inorganic bioglass [128], hyaluronic acid (HA) [129], self-assembling peptides [130], and lipids [131]. All these examples highlight the interest of using ELRs in combination with other biomaterials to obtain superior structures, such as hydrogels, that recapitulate the properties of their components, hence getting closer to the complexity of biological tissues and matrices.

Final considerations

Elastin-like recombinamers have gained an increasing interest during the last decades, with a great boost in the past few years, as highlighted by data: there are more PUBMEDindexed publications regarding ELRs issued during the last five years than in all the previous years since their discovery. This fact has enormous implications, since it means that ELRs are currently being thoroughly investigated from different points of view,

21

elucidating their molecular mechanisms of self-assembly and shedding further light on their physicochemical and mechanical properties. Furthermore, it also implies that many groups are designing and using novel or already existing ELRs for still unexplored applications, broadening their scientific impact, and probably getting them closer to be translated into clinics as biomaterials. In this review, we have gathered multiple recent examples that reflect this interest in different fields, from the basic research of the physicochemical properties of ELRs and their consideration as IDPs/IDRs, to the several applications where they have found uses, including drug, vaccine and gene delivery, and TERM. All these examples provide convincing evidences of the high potential of ELRs as biomaterials, and encourage further investigation that may lead to consider them as a benchmark in the biomedical field.

Acknowledgements

The authors are grateful for the funding from the European Commission (NMP-2014-646075), the Spanish Government (PCIN-2015-010, MAT2016-78903-R, BES-2014-069763), Junta de Castilla y León (VA317P18) and Centro en Red de Medicina Regenerativa y Terapia Celular de Castilla y León.

Abbreviations

3,4-dihydroxyphenylalanine (DOPA); albumin-binding domain (ABD); antimicrobial peptide (AMP); bone morphogenetic protein-2 (BMP-2); brain-derived neurotrophic factor (BDNF); cytotoxic T lymphocyte (CTL); dendritic cell (DC); doxorubicin (DOX) drug delivery system (DDS); elastin-like polypentapeptide (ELP); elastin-like recombinamer (ELR); extracellular matrix (ECM); fibroblast growth factor 21 (FGF21); Förster resonance energy transfer (FRET); human umbilical vein endothelial cell (HUVEC); hyaluronic acid (HA); interferon alpha (IFN-α); intrinsically disordered

protein (IDP); intrinsically disordered region (IDR); inverse transition cycling (ITC); matrix metallo-proteinase (MMP); mouse-adipose-derived stem cell (mADSC); nanoparticle (NP); nerve growth factor (NGF); peptide amphiphile (PA); poly(3hydroxybutyrate-co-3-hydroxyvalerate) (PHBV); polyethylene glycol (PEG); polyethylenimine (PEI); polymer-induced liquid-precursor (PILP); retinal pigment epithelium (RPE); silk-elastin-like (SEL); stromal cell-derived growth factor-1 (SDF1); timolol maleate (TM); tissue engineering and regenerative medicine (TERM); transition temperature (T_t); urokinase plasminogen activator (uPA); vascular endothelial growth factor (VEGF); zwitterionic polypeptides (ZIPP).

References

[1] G.C. Yeo, B. Aghaei-Ghareh-Bolagh, E.P. Brackenreg, M.A. Hiob, P. Lee, A.S. Weiss, Fabricated Elastin, Advanced Healthcare Materials 4(16) (2015) 2530-2556.

[2] W.R. Gray, L.B. Sandberg, J.A. Foster, Molecular Model for Elastin Structure and Function, Nature 246(5434) (1973) 461-466.

[3] D.W. Urry, W.D. Cunningham, T. Ohnishi, Studies on the conformation and interactions of elastin. Proton magnetic resonance of the repeating pentapeptide, Biochemistry 13(3) (1974) 609-16.

[4] D.W. Urry, K. Okamoto, R.D. Harris, C.F. Hendrix, M.M. Long, Synthetic, crosslinked polypentapeptide of tropoelastin: an anisotropic, fibrillar elastomer, Biochemistry 15(18) (1976) 4083-9.

[5] V. Renugopalakrishnan, M.A. Khaled, R.S. Rapaka, D.W. Urry, Proton magnetic resonance and conformational energy calculations of repeat peptides of tropoelastin. A permutation of the hexapeptide, Biochimica et biophysica acta 536(2) (1978) 421-8.

[6] D.W. Urry, What Sustains Life? Consilient Mechanisms for Protein-Based Machines and Materials, Birkhäuser Boston, Boston (USA), 2006.

[7] F.A. Ferrari, C. Richardson, J. Chambers, S.C. Causey, T.J. Pollock, J. Cappello, J.W. Crissman, Construction of synthetic DNA and its use in large polypeptide synthesis, Protein Polymer Technologies, Inc., United States, 1987.

[8] J. Cappello, J. Crissman, M. Dorman, M. Mikolajczak, G. Textor, M. Marquet, F.
Ferrari, Genetic Engineering of Structural Protein Polymers, Biotechnology Progress 6(3)
(1990) 198-202.

[9] D.A. Tirrell, M.J. Fournier, T.L. Mason, Genetic Engineering of Polymeric Materials, MRS Bulletin 16(7) (1991) 23-28.

[10] D.T. McPherson, C. Morrow, D.S. Minehan, J. Wu, E. Hunter, D.W. Urry, Production and purification of a recombinant elastomeric polypeptide, G-(VPGVG)19-VPGV, from Escherichia coli, Biotechnology Progress 8(4) (1992) 347-52.

[11] A. Nicol, D. Channe Gowda, D.W. Urry, Cell adhesion and growth on synthetic elastomeric matrices containing ARG-GLY–ASP–SER–3, Journal of Biomedical Materials Research 26(3) (1992) 393-413.

[12] J.C. Rodríguez-Cabello, L. Martín, M. Alonso, F.J. Arias, A.M. Testera, "Recombinamers" as advanced materials for the post-oil age, Polymer 50(22) (2009) 5159-5169.

[13] J.R. McDaniel, D.C. Radford, A. Chilkoti, A Unified Model for De Novo Design of Elastin-like Polypeptides with Tunable Inverse Transition Temperatures, Biomacromolecules 14(8) (2013) 2866-2872. [14] T. Christensen, M. Amiram, S. Dagher, K. Trabbic-Carlson, M.F. Shamji, L.A. Setton, A. Chilkoti, Fusion order controls expression level and activity of elastin-like polypeptide fusion proteins, Protein Science 18(7) (2009) 1377-1387.

[15] G. Qin, P.M. Perez, C.E. Mills, B.D. Olsen, Effect of ELP Sequence and FusionProtein Design on Concentrated Solution Self-Assembly, Biomacromolecules 17(3)(2016) 928-934.

[16] A. Tarakanova, W. Huang, A.S. Weiss, D.L. Kaplan, M.J. Buehler, Computational smart polymer design based on elastin protein mutability, Biomaterials 127 (2017) 49-60.

[17] A. Prhashanna, P.A. Taylor, J. Qin, K.L. Kiick, A. Jayaraman, Effect of Peptide Sequence on the LCST-Like Transition of Elastin-Like Peptides and Elastin-Like Peptide–Collagen-Like Peptide Conjugates: Simulations and Experiments, Biomacromolecules 20(3) (2019) 1178-1189.

[18] N.K. Li, S. Roberts, F.G. Quiroz, A. Chilkoti, Y.G. Yingling, Sequence Directionality Dramatically Affects LCST Behavior of Elastin-Like Polypeptides, Biomacromolecules 19(7) (2018) 2496-2505.

[19] J. Yeo, W. Huang, A. Tarakanova, Y.-W. Zhang, D.L. Kaplan, M.J. Buehler, Unraveling the molecular mechanisms of thermo-responsive properties of silk-elastinlike proteins by integrating multiscale modeling and experiment, Journal of Materials Chemistry B 6(22) (2018) 3727-3734.

[20] J.C. Rodríguez-Cabello, F.J. Arias, M.A. Rodrigo, A. Girotti, Elastin-like polypeptides in drug delivery, Advanced Drug Delivery Reviews 97 (2016) 85-100.

[21] J.C. Rodríguez-Cabello, M.J. Piña, A. Ibáñez-Fonseca, A. Fernández-Colino, F.J. Arias, Nanotechnological Approaches to Therapeutic Delivery Using Elastin-Like Recombinamers, Bioconjugate Chemistry 26(7) (2015) 1252-1265.

[22] A. Girotti, D. Orbanic, A. Ibáñez-Fonseca, C. Gonzalez-Obeso, J.C. Rodríguez-Cabello, Recombinant Technology in the Development of Materials and Systems for Soft-Tissue Repair, Advanced Healthcare Materials 4(16) (2015) 2423-2455.

[23] D.L. Nettles, A. Chilkoti, L.A. Setton, Applications of elastin-like polypeptides in tissue engineering, Advanced Drug Delivery Reviews 62(15) (2010) 1479-1485.

[24] J.C. Rodríguez-Cabello, I. González de Torre, A. Ibañez-Fonseca, M. Alonso, Bioactive scaffolds based on elastin-like materials for wound healing, Advanced Drug Delivery Reviews 129 (2018) 118-133.

[25] P. Tompa, Structure and function of intrinsically disordered proteins, Chapman & Hall/CRC Press2010.

[26] M.E. Oates, P. Romero, T. Ishida, M. Ghalwash, M.J. Mizianty, B. Xue, Z. Dosztányi, V.N. Uversky, Z. Obradovic, L. Kurgan, A.K. Dunker, J. Gough, D2P2: database of disordered protein predictions, Nucleic Acids Research 41(D1) (2012) D508-D516.

[27] F.-X. Theillet, L. Kalmar, P. Tompa, K.-H. Han, P. Selenko, A.K. Dunker, G.W. Daughdrill, V.N. Uversky, The alphabet of intrinsic disorder: I. Act like a Pro: On the abundance and roles of proline residues in intrinsically disordered proteins, Intrinsically disordered proteins 1(1) (2013) e24360-e24360.

[28] V.N. Uversky, Natively unfolded proteins: A point where biology waits for physics,Protein Science 11(4) (2002) 739-756.

26

[29] M.M. Babu, The contribution of intrinsically disordered regions to protein function, cellular complexity, and human disease, Biochemical Society Transactions 44(5) (2016) 1185-1200.

[30] Timothy J. Nott, E. Petsalaki, P. Farber, D. Jervis, E. Fussner, A. Plochowietz, T.D. Craggs, David P. Bazett-Jones, T. Pawson, Julie D. Forman-Kay, Andrew J. Baldwin, Phase Transition of a Disordered Nuage Protein Generates Environmentally Responsive Membraneless Organelles, Molecular Cell 57(5) (2015) 936-947.

[31] P.E. Wright, H.J. Dyson, Intrinsically disordered proteins in cellular signalling and regulation, Nature Reviews Molecular Cell Biology 16(1) (2015) 18-29.

[32] V.N. Uversky, Intrinsically disordered proteins and their (disordered) proteomes in neurodegenerative disorders, Frontiers in aging neuroscience 7 (2015) 18-18.

[33] S. Rauscher, S. Baud, M. Miao, Fred W. Keeley, R. Pomès, Proline and Glycine Control Protein Self-Organization into Elastomeric or Amyloid Fibrils, Structure 14(11) (2006) 1667-1676.

[34] K.M. Ruff, S. Roberts, A. Chilkoti, R.V. Pappu, Advances in Understanding Stimulus-Responsive Phase Behavior of Intrinsically Disordered Protein Polymers, Journal of Molecular Biology 430(23) (2018) 4619-4635.

[35] R. Balu, R. Knott, N.P. Cowieson, C.M. Elvin, A.J. Hill, N.R. Choudhury, N.K. Dutta, Structural ensembles reveal intrinsic disorder for the multi-stimuli responsive biomimetic protein Rec1-resilin, Scientific Reports 5(1) (2015) 10896-10896.

[36] S. Roberts, M. Dzuricky, A. Chilkoti, Elastin-like polypeptides as models of intrinsically disordered proteins, FEBS Letters 589(19PartA) (2015) 2477-2486.

[37] M. Dzuricky, S. Roberts, A. Chilkoti, Convergence of Artificial Protein Polymers and Intrinsically Disordered Proteins, Biochemistry 57(17) (2018) 2405-2414.

27

[38] Y. Zhang, V. Zai-Rose, C.J. Price, N.A. Ezzell, G.L. Bidwell, J.J. Correia, N.C.Fitzkee, Modeling the Early Stages of Phase Separation in Disordered Elastin-likeProteins, Biophysical Journal 114(7) (2018) 1563-1578.

[39] S.E. Reichheld, L.D. Muiznieks, F.W. Keeley, S. Sharpe, Direct observation of structure and dynamics during phase separation of an elastomeric protein, Proceedings of the National Academy of Sciences of the United States of America 114(22) (2017) E4408-E4415.

[40] N.K. Li, F.G.a. Quiroz, C.K. Hall, A. Chilkoti, Y.G. Yingling, Molecular description of the lcst behavior of an elastin-like polypeptide, Biomacromolecules 15(10) (2014) 3522-3530.

[41] M. Feric, N. Vaidya, T.S. Harmon, D.M. Mitrea, L. Zhu, T.M. Richardson, R.W. Kriwacki, R.V. Pappu, C.P. Brangwynne, Coexisting Liquid Phases Underlie Nucleolar Subcompartments, Cell 165(7) (2016) 1686-1697.

[42] M.K. Pastuszka, C.T. Okamoto, S.F. Hamm-Alvarez, J.A. MacKay, Flipping the Switch on Clathrin-Mediated Endocytosis using Thermally Responsive Protein Microdomains, Advanced functional materials 24(34) (2014) 5340-5347.

[43] M.C. Huber, A. Schreiber, P. von Olshausen, B.R. Varga, O. Kretz, B. Joch, S. Barnert, R. Schubert, S. Eimer, P. Kele, S.M. Schiller, Designer amphiphilic proteins as building blocks for the intracellular formation of organelle-like compartments, Nature Materials 14(1) (2015) 125-132.

[44] K. Vogele, T. Frank, L. Gasser, M.A. Goetzfried, M.W. Hackl, S.A. Sieber, F.C. Simmel, T. Pirzer, Towards synthetic cells using peptide-based reaction compartments, Nature Communications 9(1) (2018) 3862.

[45] E. Beniash, J.P. Simmer, H.C. Margolis, Structural changes in amelogenin upon selfassembly and mineral interactions, Journal of dental research 91(10) (2012) 967-72.

[46] A.L. Boskey, E. Villarreal-Ramirez, Intrinsically disordered proteins and biomineralization, Matrix Biology 52-54 (2016) 43-59.

[47] T. Wald, F. Spoutil, A. Osickova, M. Prochazkova, O. Benada, P. Kasparek, L. Bumba, O.D. Klein, R. Sedlacek, P. Sebo, J. Prochazka, R. Osicka, Intrinsically disordered proteins drive enamel formation via an evolutionarily conserved self-assembly motif, Proceedings of the National Academy of Sciences of the United States of America 114(9) (2017) E1641-E1650.

[48] M.H. Misbah, M. Espanol, L. Quintanilla, M.P. Ginebra, J.C. Rodríguez-Cabello, Formation of calcium phosphate nanostructures under the influence of self-assembling hybrid elastin-like-statherin recombinamers, RSC Advances 6(37) (2016) 31225-31234.

[49] Y. Li, X. Chen, A. Fok, J.C. Rodriguez-Cabello, C. Aparicio, Biomimetic Mineralization of Recombinamer-Based Hydrogels toward Controlled Morphologies and High Mineral Density, ACS Applied Materials & Interfaces 7(46) (2015) 25784-25792.

[50] M.H. Misbah, M. Santos, L. Quintanilla, C. Günter, M. Alonso, A. Taubert, J.C. Rodríguez-Cabello, Recombinant DNA technology and click chemistry: a powerful combination for generating a hybrid elastin-like-statherin hydrogel to control calcium phosphate mineralization, Beilstein journal of nanotechnology 8 (2017) 772-783.

[51] Y. Li, X. Chen, A.J. Ribeiro, E.D. Jensen, K.V. Holmberg, J.C. Rodriguez-Cabello,
C. Aparicio, Hybrid Nanotopographical Surfaces Obtained by Biomimetic Mineralization
of Statherin-Inspired Elastin-Like Recombinamers, Advanced Healthcare Materials 3(10)
(2014) 1638-1647.

[52] Y. Li, J.C. Rodriguez-Cabello, C. Aparicio, Intrafibrillar Mineralization of Self-Assembled Elastin-Like Recombinamer Fibrils, ACS Applied Materials & Interfaces 9(7)
(2017) 5838-5846.

[53] S. Elsharkawy, M. Al-Jawad, M.F. Pantano, E. Tejeda-Montes, K. Mehta, H. Jamal, S. Agarwal, K. Shuturminska, A. Rice, N.V. Tarakina, R.M. Wilson, A.J. Bushby, M. Alonso, J.C. Rodriguez-Cabello, E. Barbieri, A. del Río Hernández, M.M. Stevens, N.M. Pugno, P. Anderson, A. Mata, Protein disorder–order interplay to guide the growth of hierarchical mineralized structures, Nature Communications 9(1) (2018) 2145-2145.

[54] S. Roberts, T.S. Harmon, J.L. Schaal, V. Miao, K. Li, A. Hunt, Y. Wen, T.G. Oas, J.H. Collier, R.V. Pappu, A. Chilkoti, Injectable tissue integrating networks from recombinant polypeptides with tunable order, Nature Materials 17(12) (2018) 1154-1163.

[55] A. Fernández-Colino, F.J. Arias, M. Alonso, J.C. Rodríguez-Cabello, Amphiphilic Elastin-Like Block Co-Recombinamers Containing Leucine Zippers: Cooperative Interplay between Both Domains Results in Injectable and Stable Hydrogels, Biomacromolecules 16(10) (2015) 3389-3398.

[56] E. Salvagni, G. Berguig, E. Engel, J.C. Rodriguez-Cabello, G. Coullerez, M. Textor, J.A. Planell, F.J. Gil, C. Aparicio, A bioactive elastin-like recombinamer reduces unspecific protein adsorption and enhances cell response on titanium surfaces, Colloids and Surfaces B: Biointerfaces 114 (2014) 225-233.

[57] M. Pierna, M. Santos, F.J. Arias, M. Alonso, J.C. Rodríguez-Cabello, Efficient Cell and Cell-Sheet Harvesting Based on Smart Surfaces Coated with a Multifunctional and Self-Organizing Elastin-Like Recombinamer, Biomacromolecules 14(6) (2013) 1893-1903. [58] L. Li, C.-K. Mo, A. Chilkoti, G.P. Lopez, N.J. Carroll, Creating cellular patterns using genetically engineered, gold- and cell-binding polypeptides, Biointerphases 11(2) (2016) 021009-021009.

[59] T. Flora, I.G. de Torre, L. Quintanilla, M. Alonso, J.C. Rodríguez-Cabello, Spatial control and cell adhesion selectivity on model gold surfaces grafted with elastin-like recombinamers, European Polymer Journal 106 (2018) 19-29.

[60] R.E.W. Hancock, H.-G. Sahl, Antimicrobial and host-defense peptides as new antiinfective therapeutic strategies, Nature Biotechnology 24 (2006) 1551.

[61] S. Atefyekta, M. Pihl, C. Lindsay, S.C. Heilshorn, M. Andersson, Antibiofilm elastin-like polypeptide coatings: functionality, stability, and selectivity, Acta Biomaterialia (2018).

[62] K. Trabbic-Carlson, L. Liu, B. Kim, A. Chilkoti, Expression and purification of recombinant proteins from Escherichia coli: Comparison of an elastin-like polypeptide fusion with an oligohistidine fusion, Protein Science 13(12) (2009) 3274-3284.

[63] A. da Costa, A.M. Pereira, A.C. Gomes, J.C. Rodriguez-Cabello, M. Casal, R. Machado, Production of bioactive hepcidin by recombinant DNA tagging with an elastinlike recombinamer, New Biotechnology 46 (2018) 45-53.

[64] A. Da Costa, R. Machado, A. Ribeiro, T. Collins, V. Thiagarajan, M.T. Neves-Petersen, J.C. Rodriguez-Cabello, A.C. Gomes, M. Casal, Development of elastin-like recombinamer films with antimicrobial activity, Biomacromolecules 16(2) (2015).

[65] J. Hu, G. Wang, X. Liu, W. Gao, Enhancing Pharmacokinetics, Tumor Accumulation, and Antitumor Efficacy by Elastin-Like Polypeptide Fusion of Interferon Alpha, Advanced Materials 27(45) (2015) 7320-7324.

[66] A. Yeboah, R.I. Cohen, R. Faulknor, R. Schloss, M.L. Yarmush, F. Berthiaume, The development and characterization of SDF1 α -elastin-like-peptide nanoparticles for wound healing, Journal of Controlled Release 232 (2016) 238-247.

[67] C.A. Gilroy, S. Roberts, A. Chilkoti, Fusion of fibroblast growth factor 21 to a thermally responsive biopolymer forms an injectable depot with sustained anti-diabetic action, Journal of Controlled Release 277 (2018) 154-164.

[68] P.G. Sreekumar, Z. Li, W. Wang, C. Spee, D.R. Hinton, R. Kannan, J.A. MacKay, Intra-vitreal αB crystallin fused to elastin-like polypeptide provides neuroprotection in a mouse model of age-related macular degeneration, Journal of Controlled Release 283 (2018) 94-104.

[69] S. Banskota, P. Yousefpour, N. Kirmani, X. Li, A. Chilkoti, Long circulating genetically encoded intrinsically disordered zwitterionic polypeptides for drug delivery, Biomaterials 192 (2019) 475-485.

[70] A. Fernández-Colino, D.A. Quinteros, D.A. Allemandi, A. Girotti, S.D. Palma, F.J. Arias, Self-Assembling Elastin-Like Hydrogels for Timolol Delivery: Development of an Ophthalmic Formulation Against Glaucoma, Molecular Pharmaceutics 14(12) (2017) 4498-4508.

[71] A. Fernández-Colino, F.J. Arias, M. Alonso, J.C. Rodríguez-Cabello, Self-Organized ECM-Mimetic Model Based on an Amphiphilic Multiblock Silk-Elastin-Like Corecombinamer with a Concomitant Dual Physical Gelation Process, Biomacromolecules 15(10) (2014) 3781-3793.

[72] P. Yousefpour, J.R. McDaniel, V. Prasad, L. Ahn, X. Li, R. Subrahmanyan, I. Weitzhandler, S. Suter, A. Chilkoti, Genetically Encoding Albumin Binding into

32

Chemotherapeutic-loaded Polypeptide Nanoparticles Enhances Their Antitumor Efficacy, Nano Letters 18(12) (2018) 7784-7793.

[73] L.M.P.E. van Oppen, J. Pille, C. Stuut, M. van Stevendaal, L.N. van der Vorm, J.A.M. Smeitink, W.J.H. Koopman, P.H.G.M. Willems, J.C.M. van Hest, R. Brock, Octaarginine boosts the penetration of elastin-like polypeptide nanoparticles in 3D cancer models, European Journal of Pharmaceutics and Biopharmaceutics 137 (2019) 175-184.

[74] C. Gonzalez-Obeso, A. Girotti, J.C. Rodriguez-Cabello, A transferrin receptorbinding mucoadhesive elastin-like recombinamer: In vitro and in vivo characterization, Acta Biomaterialia 88 (2019) 241-250.

[75] C. García-Arévalo, J.F. Bermejo-Martín, L. Rico, V. Iglesias, L. Martín, J.C. Rodríguez-Cabello, F.J. Arias, Immunomodulatory Nanoparticles from Elastin-Like Recombinamers: Single-Molecules for Tuberculosis Vaccine Development, Molecular Pharmaceutics 10(2) (2013) 586-597.

[76] R.S. Ingrole, W. Tao, J.N. Tripathy, H.S. Gill, Synthesis and Immunogenicity Assessment of Elastin-Like Polypeptide-M2e Construct as an Influenza Antigen, Nano LIFE 04(02) (2014) 1450004.

[77] P. Wang, S. Dong, P. Zhao, X. He, M. Chen, Direct loading of CTL epitopes onto MHC class I complexes on dendritic cell surface in vivo, Biomaterials 182 (2018) 92-103.

[78] M.J. Piña, S.M. Alex, F.J. Arias, M. Santos, J.C. Rodriguez-Cabello, R.M. Ramesan, C.P. Sharma, Elastin-like recombinamers with acquired functionalities for gene-delivery applications, Journal of Biomedical Materials Research Part A 103(10) (2015) 3166-3178.

33

[79] M.J. Piña, A. Girotti, M. Santos, J.C. Rodríguez-Cabello, F.J. Arias, Biocompatible ELR-Based Polyplexes Coated with MUC1 Specific Aptamers and Targeted for Breast Cancer Gene Therapy, Molecular Pharmaceutics 13(3) (2016) 795-808.

[80] C.H. Lee, R.S.J. Ingrole, H.S. Gill, Generation of induced pluripotent stem cells using elastin like polypeptides as a non-viral gene delivery system, Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease (2019).

[81] I. González de Torre, M. Santos, L. Quintanilla, A. Testera, M. Alonso, J.C. Rodríguez Cabello, Elastin-like recombinamer catalyst-free click gels: Characterization of poroelastic and intrinsic viscoelastic properties, Acta Biomaterialia 10(6) (2014) 2495-2505.

[82] A. Ibáñez-Fonseca, T.L. Ramos, I. González de Torre, L.I. Sánchez-Abarca, S. Muntión, F.J. Arias, M.C. del Cañizo, M. Alonso, F. Sánchez-Guijo, J.C. Rodríguez-Cabello, Biocompatibility of two model elastin-like recombinamer-based hydrogels formed through physical or chemical cross-linking for various applications in tissue engineering and regenerative medicine, Journal of Tissue Engineering and Regenerative Medicine 12(3) (2018) e1450-e1460.

[83] A. Fernández-Colino, F. Wolf, H. Keijdener, S. Rütten, T. Schmitz-Rode, S. Jockenhoevel, J.C. Rodríguez-Cabello, P. Mela, Macroporous click-elastin-like hydrogels for tissue engineering applications, Materials Science and Engineering: C 88 (2018) 140-147.

[84] C.M. Madl, S.C. Heilshorn, Tyrosine-Selective Functionalization for Bio-Orthogonal Cross-Linking of Engineered Protein Hydrogels, Bioconjugate Chemistry 28(3) (2017) 724-730. [85] Y.-N. Zhang, R.K. Avery, Q. Vallmajo-Martin, A. Assmann, A. Vegh, A. Memic, B.D. Olsen, N. Annabi, A. Khademhosseini, A Highly Elastic and Rapidly Crosslinkable Elastin-Like Polypeptide-Based Hydrogel for Biomedical Applications, Advanced Functional Materials 25(30) (2015) 4814-4826.

[86] L. Martín, F.J. Arias, M. Alonso, C. García-Arévalo, J.C. Rodríguez-Cabello, Rapid micropatterning by temperature-triggered reversible gelation of a recombinant smart elastin-like tetrablock-copolymer, Soft Matter 6(6) (2010) 1121-1124.

[87] L.W. Jennifer, J.M. James, Vascularization of Engineered Tissues: Approaches to Promote Angiogenesis in Biomaterials, Current Topics in Medicinal Chemistry 8(4) (2008) 300-310.

[88] J.J. Kim, L. Hou, N.F. Huang, Vascularization of three-dimensional engineered tissues for regenerative medicine applications, Acta Biomaterialia 41 (2016) 17-26.

[89] G.M. Mitchell, W.A. Morrison, In Vitro and In Vivo Approaches for Prevascularization of 3-Dimensional Engineered Tissues, in: W. Holnthoner, A. Banfi, J. Kirkpatrick, H. Redl (Eds.), Vascularization for Tissue Engineering and Regenerative Medicine, Springer International Publishing, Cham, 2017, pp. 1-27.

[90] E. Ruoslahti, RGD AND OTHER RECOGNITION SEQUENCES FOR INTEGRINS, Annual Review of Cell and Developmental Biology 12(1) (1996) 697-715.

[91] S.P. Massia, J.A. Hubbell, Vascular endothelial cell adhesion and spreading promoted by the peptide REDV of the IIICS region of plasma fibronectin is mediated by integrin alpha 4 beta 1, Journal of Biological Chemistry 267(20) (1992) 14019-14026.

[92] S. Akthar, D.F. Patel, R.C. Beale, T. Peiró, X. Xu, A. Gaggar, P.L. Jackson, J.E.Blalock, C.M. Lloyd, R.J. Snelgrove, Matrikines are key regulators in modulating the

amplitude of lung inflammation in acute pulmonary infection, Nature Communications 6 (2015) 8423.

[93] S.M. Staubli, G. Cerino, I. Gonzalez De Torre, M. Alonso, D. Oertli, F. Eckstein, K. Glatz, J.C. Rodríguez Cabello, A. Marsano, Control of angiogenesis and host response by modulating the cell adhesion properties of an Elastin-Like Recombinamer-based hydrogel, Biomaterials 135 (2017) 30-41.

[94] A.S. Alagoz, J.C. Rodriguez-Cabello, V. Hasirci, PHBV wet-spun scaffold coated with ELR-REDV improves vascularization for bone tissue engineering, Biomedical Materials 13(5) (2018) 055010.

[95] A. Hoeben, B. Landuyt, M.S. Highley, H. Wildiers, A.T. Van Oosterom, E.A. De Bruijn, Vascular Endothelial Growth Factor and Angiogenesis, Pharmacological Reviews 56(4) (2004) 549-580.

[96] L. Cai, C.B. Dinh, S.C. Heilshorn, One-pot synthesis of elastin-like polypeptide hydrogels with grafted VEGF-mimetic peptides, Biomaterials Science 2(5) (2014) 757-765.

[97] T. Flora, I.G. de Torre, M. Alonso, J.C. Rodríguez-Cabello, Tethering QK peptide to enhance angiogenesis in elastin-like recombinamer (ELR) hydrogels, Journal of Materials Science: Materials in Medicine 30(2) (2019) 30.

[98] C.M. Madl, L.M. Katz, S.C. Heilshorn, Tuning Bulk Hydrogel Degradation by Simultaneous Control of Proteolytic Cleavage Kinetics and Hydrogel Network Architecture, ACS Macro Letters 7(11) (2018) 1302-1307.

[99] K.S. Straley, S.C. Heilshorn, Dynamic, 3D-Pattern Formation Within Enzyme-Responsive Hydrogels, Advanced Materials 21(41) (2009) 4148-4152. [100] J.C. Rodríguez-Cabello, L. Martín, A. Girotti, C. García-Arévalo, F.J. Arias, M. Alonso, Emerging applications of multifunctional elastin-like recombinamers, Nanomedicine 6(1) (2011) 111-122.

[101] I.G. de Torre, F. Wolf, M. Santos, L. Rongen, M. Alonso, S. Jockenhoevel, J.C. Rodríguez-Cabello, P. Mela, Elastin-like recombinamer-covered stents: Towards a fully biocompatible and non-thrombogenic device for cardiovascular diseases, Acta Biomaterialia 12 (2015) 146-155.

[102] A. Mahara, K.L. Kiick, T. Yamaoka, In vivo guided vascular regeneration with a non-porous elastin-like polypeptide hydrogel tubular scaffold, Journal of Biomedical Materials Research Part A 105(6) (2017) 1746-1755.

[103] K.E. Inostroza-Brito, E. Collin, O. Siton-Mendelson, K.H. Smith, A. Monge-Marcet, D.S. Ferreira, R.P. Rodríguez, M. Alonso, J.C. Rodríguez-Cabello, R.L. Reis, F. Sagués, L. Botto, R. Bitton, H.S. Azevedo, A. Mata, Co-assembly, spatiotemporal control and morphogenesis of a hybrid protein–peptide system, Nature Chemistry 7 (2015) 897.

[104] F. Cipriani, M. Krüger, I.G. de Torre, L.Q. Sierra, M.A. Rodrigo, L. Kock, J.C. Rodriguez-Cabello, Cartilage Regeneration in Preannealed Silk Elastin-Like Co-Recombinamers Injectable Hydrogel Embedded with Mature Chondrocytes in an Ex Vivo Culture Platform, Biomacromolecules 19(11) (2018) 4333-4347.

[105] E. Tejeda-Montes, A. Klymov, M.R. Nejadnik, M. Alonso, J.C. Rodriguez-Cabello, X.F. Walboomers, A. Mata, Mineralization and bone regeneration using a bioactive elastin-like recombinamer membrane, Biomaterials 35(29) (2014) 8339-8347.

[106] M. Vila, A. García, A. Girotti, M. Alonso, J.C. Rodríguez-Cabello, A. González-Vázquez, J.A. Planell, E. Engel, J. Buján, N. García-Honduvilla, M. Vallet-Regí, 3D

37

silicon doped hydroxyapatite scaffolds decorated with Elastin-like Recombinamers for bone regenerative medicine, Acta Biomaterialia 45 (2016) 349-356.

[107] D. Pescador, A. Ibáñez-Fonseca, F. Sánchez-Guijo, J.G. Briñón, F.J. Arias, S. Muntión, C. Hernández, A. Girotti, M. Alonso, M.C. del Cañizo, J.C. Rodríguez-Cabello, J.F. Blanco, Regeneration of hyaline cartilage promoted by xenogeneic mesenchymal stromal cells embedded within elastin-like recombinamer-based bioactive hydrogels, Journal of Materials Science: Materials in Medicine 28(8) (2017) 115.

[108] D.J. Coletta, A. Ibáñez-Fonseca, L.R. Missana, M.V. Jammal, E.J. Vitelli, M. Aimone, F. Zabalza, J.P.M. Issa, M. Alonso, J.C. Rodríguez-Cabello, S. Feldman, Bone Regeneration Mediated by a Bioactive and Biodegradable Extracellular Matrix-Like Hydrogel Based on Elastin-Like Recombinamers, Tissue Engineering Part A 23(23-24) (2017) 1361-1371.

[109] T. Johnson, P. Koria, Expression and Purification of Neurotrophin-Elastin-Like Peptide Fusion Proteins for Neural Regeneration, BioDrugs 30(2) (2016) 117-127.

[110] C.M. Madl, B.L. LeSavage, R.E. Dewi, C.B. Dinh, R.S. Stowers, M. Khariton, K.J. Lampe, D. Nguyen, O. Chaudhuri, A. Enejder, S.C. Heilshorn, Maintenance of neural progenitor cell stemness in 3D hydrogels requires matrix remodelling, Nature Materials 16 (2017) 1233.

[111] I. González de Torre, A. Ibáñez-Fonseca, L. Quintanilla, M. Alonso, J.-C. Rodríguez-Cabello, Random and oriented electrospun fibers based on a multicomponent, in situ clickable elastin-like recombinamer system for dermal tissue engineering, Acta Biomaterialia 72 (2018) 137-149.

[112] A. Fernández-Colino, F. Wolf, S. Rütten, J.C. Rodríguez-Cabello, S. Jockenhoevel,P. Mela, Combining Catalyst-Free Click Chemistry with Coaxial Electrospinning to

Obtain Long-Term, Water-Stable, Bioactive Elastin-Like Fibers for Tissue Engineering Applications, Macromolecular Bioscience 18(11) (2018) 1800147.

[113] A.R. Swartz, Q. Sun, W. Chen, Ligand-Induced Cross-Linking of Z-Elastin-like Polypeptide-Functionalized E2 Protein Nanoparticles for Enhanced Affinity Precipitation of Antibodies, Biomacromolecules 18(5) (2017) 1654-1659.

[114] A.R. Swartz, W. Chen, SpyTag/SpyCatcher Functionalization of E2 Nanocages with Stimuli-Responsive Z-ELP Affinity Domains for Tunable Monoclonal Antibody Binding and Precipitation Properties, Bioconjugate Chemistry 29(9) (2018) 3113-3120.

[115] R. Wieduwild, M. Howarth, Assembling and decorating hyaluronan hydrogels with twin protein superglues to mimic cell-cell interactions, Biomaterials 180 (2018) 253-264.

[116] M.G. Haugh, T.J. Vaughan, C.M. Madl, R.M. Raftery, L.M. McNamara, F.J. O'Brien, S.C. Heilshorn, Investigating the interplay between substrate stiffness and ligand chemistry in directing mesenchymal stem cell differentiation within 3D macro-porous substrates, Biomaterials 171 (2018) 23-33.

[117] C.E. Mills, Z. Michaud, B.D. Olsen, Elastin-like Polypeptide (ELP) Charge Influences Self-Assembly of ELP–mCherry Fusion Proteins, Biomacromolecules 19(7) (2018) 2517-2525.

[118] A. Ibáñez-Fonseca, M. Alonso, F.J. Arias, J.C. Rodríguez-Cabello, Förster Resonance Energy Transfer-Paired Hydrogel Forming Silk-Elastin-Like Recombinamers by Recombinant Conjugation of Fluorescent Proteins, Bioconjugate Chemistry 28(3) (2017) 828-835.

[119] M.J. Brennan, B.F. Kilbride, J.J. Wilker, J.C. Liu, A bioinspired elastin-based protein for a cytocompatible underwater adhesive, Biomaterials 124 (2017) 116-125.

[120] S. Yang, S. Wei, Y. Mao, H. Zheng, J. Feng, J. Cui, X. Xie, F. Chen, H. Li, Novel hemostatic biomolecules based on elastin-like polypeptides and the self-assembling peptide RADA-16, BMC Biotechnology 18(1) (2018) 12.

[121] W. Huang, A. Tarakanova, N. Dinjaski, Q. Wang, X. Xia, Y. Chen, J.Y. Wong,
M.J. Buehler, D.L. Kaplan, Design of Multistimuli Responsive Hydrogels Using
Integrated Modeling and Genetically Engineered Silk–Elastin-Like Proteins, Advanced
Functional Materials 26(23) (2016) 4113-4123.

[122] E.G. Roberts, N.-G. Rim, W. Huang, A. Tarakanova, J. Yeo, M.J. Buehler, D.L. Kaplan, J.Y. Wong, Fabrication and Characterization of Recombinant Silk-Elastin-Like-Protein (SELP) Fiber, Macromolecular Bioscience 18(12) (2018) 1800265.

[123] K.J. Isaacson, M.M. Jensen, A.H. Watanabe, B.E. Green, M.A. Correa, J. Cappello,H. Ghandehari, Self-Assembly of Thermoresponsive Recombinant Silk-ElastinlikeNanogels, Macromolecular Bioscience 18(1) (2018) 1700192.

[124] T. Luo, K.L. Kiick, Noncovalent Modulation of the Inverse Temperature Transition and Self-Assembly of Elastin-b-Collagen-like Peptide Bioconjugates, Journal of the American Chemical Society 137(49) (2015) 15362-15365.

[125] J. Qin, T. Luo, K.L. Kiick, Self-Assembly of Stable Nanoscale Platelets from Designed Elastin-like Peptide–Collagen-like Peptide Bioconjugates, Biomacromolecules (2019).

[126] H. Wang, L. Cai, A. Paul, A. Enejder, S.C. Heilshorn, Hybrid Elastin-like Polypeptide–Polyethylene Glycol (ELP-PEG) Hydrogels with Improved Transparency and Independent Control of Matrix Mechanics and Cell Ligand Density, Biomacromolecules 15(9) (2014) 3421-3428. [127] S. Singh, D.E. Demco, K. Rahimi, R. Fechete, J.C. Rodriguez-Cabello, M. Möller, Aggregation behaviour of biohybrid microgels from elastin-like recombinamers, Soft Matter 12(29) (2016) 6240-6252.

[128] Q. Zeng, M.S. Desai, H.-E. Jin, J.H. Lee, J. Chang, S.-W. Lee, Self-Healing Elastin–Bioglass Hydrogels, Biomacromolecules 17(8) (2016) 2619-2625.

[129] D. Zhu, H. Wang, P. Trinh, S.C. Heilshorn, F. Yang, Elastin-like protein-hyaluronic acid (ELP-HA) hydrogels with decoupled mechanical and biochemical cues for cartilage regeneration, Biomaterials 127 (2017) 132-140.

[130] K.E. Inostroza-Brito, E.C. Collin, A. Majkowska, S. Elsharkawy, A. Rice, A.E. del Río Hernández, X. Xiao, J. Rodríguez-Cabello, A. Mata, Cross-linking of a biopolymerpeptide co-assembling system, Acta Biomaterialia 58 (2017) 80-89.

[131] D. Mozhdehi, K.M. Luginbuhl, J.R. Simon, M. Dzuricky, R. Berger, H.S. Varol, F.C. Huang, K.L. Buehne, N.R. Mayne, I. Weitzhandler, M. Bonn, S.H. Parekh, A. Chilkoti, Genetically encoded lipid–polypeptide hybrid biomaterials that exhibit temperature-triggered hierarchical self-assembly, Nature Chemistry 10(5) (2018) 496-505.