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Abstract: A flow-through reactor for the extraction of hemicelluloses with hot pressurized water was scaled up. Experiments were conducted with the two systems using catalpa wood as raw material at different temperatures, to determine the effectiveness of the scale-up.

The one pilot reactor system was subsequently upgraded by designing and building a manifold system capable of working in series. Technological innovations implemented, permitted a continuous operability, minimizing downtime when replacing the biomass during loading and unloading phases. A study was performed to determine the evolution of the composition and molecular weight of the extracted solution, by varying its residence time within the system. The plant worked homogeneously and there were no deviations in the characteristics of the liquid product flowing from one unit to the next.

The main objective of this work was to verify the possibility of using the plant set-up for an industrial process by comparing two scales.

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Well-respected Editor and Reviewers,

We offer our piece of research to publish in Bioresource Technologies (70.040: Other thermochemical processes).

*“Hydrothermal extraction of hemicellulose from lab to pilot scale”*

In this research, a pilot multistage flow-through reactor for the liquid hot water extraction of hemicellulose from lignocellulosic biomass was designed, built and tested.

A laboratory-scale plant was initially scaled with a factor of 72 and the effectiveness of the scale-up was verified through experiments with the two systems.

The pilot plant was implemented with technological innovations to become a multistage pilot plant with a continuous operability, minimum downtimes and easiness in the operation.

The evolution of the characteristics of the liquid extract was studied, when changing the liquid and solid residence time within the system, and the operating temperature.

The **main fields** considered in the paper and the **innovation level** are:

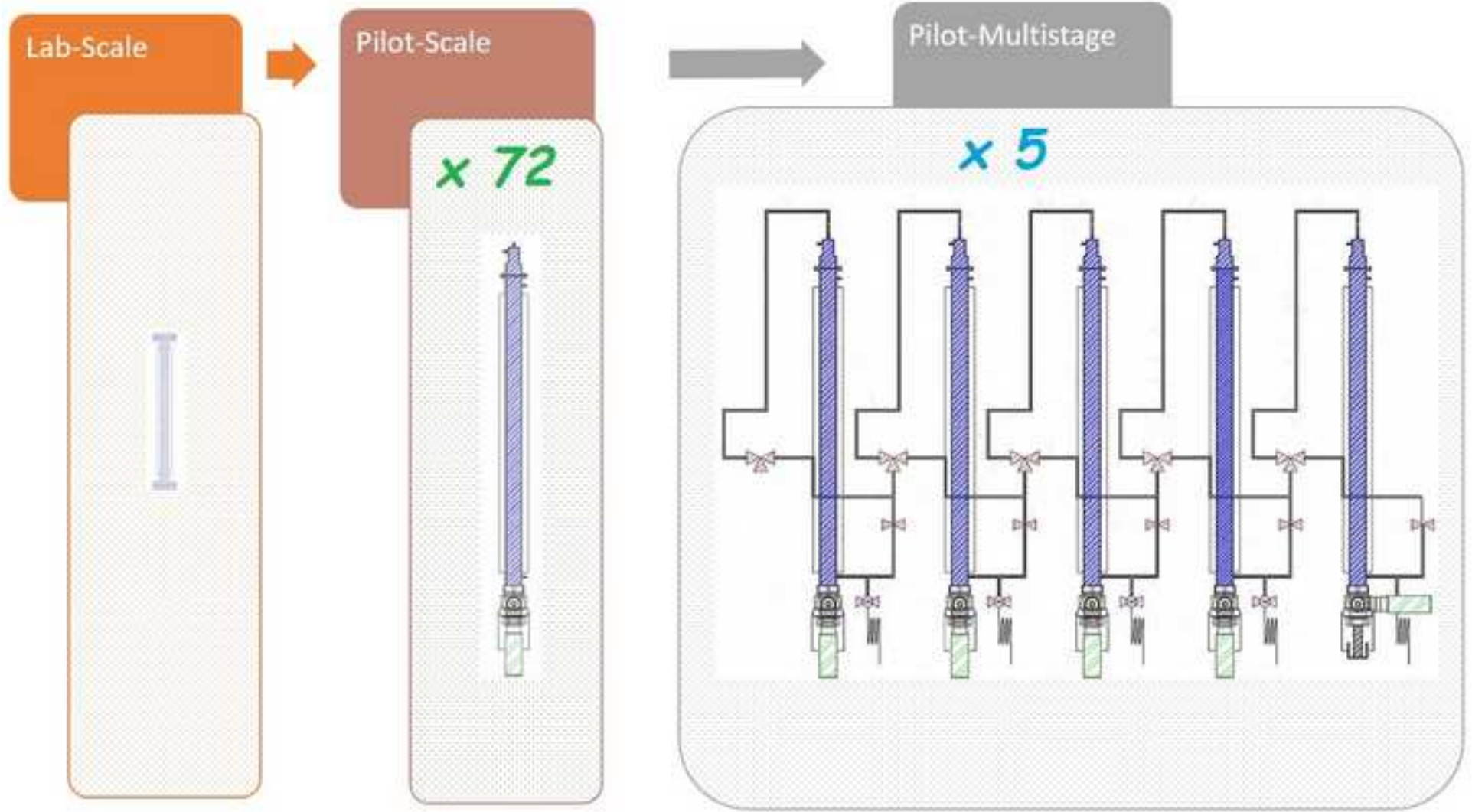
Process innovative scale-up	
Hydrothermal fractionation	
Process improvement	

We strongly believe that our findings would appeal to the readership of Bioresource Technologies as in this work we propose and test a totally new plant set-up for biomass pretreatment, demonstrating its manageability and speed of operation, key features in an industrial process.

It is the original work of the authors, not previously submitted to BITE. All the authors mutually agree that it should be submitted to BITE.

Sincerely yours,

The authors.



## **HIGHLIGHTS**

- Novel pilot plant for hydrothermal pretreatment of lignocellulosic biomass.
- Optimization of reactor set-up to minimize downtimes in hemicellulose extraction.
- Verification of scale-up efficacy through experiment at different conditions.
- Evolution of the liquid extract when changing its residence time in the system.

1 Hydrothermal extraction of hemicellulose from lab to  
2 pilot scale

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4

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18 **Abstract**

19 A flow-through reactor for the extraction of hemicelluloses with hot pressurized water was scaled up.  
20 Experiments were conducted with the two systems using catalpa wood as raw material at different  
21 temperatures, to determine the effectiveness of the scale-up.

22 The one pilot reactor system was subsequently upgraded by designing and building a manifold system  
23 capable of working in series. Technological innovations implemented, permitted a continuous  
24 operability, minimizing downtime when replacing the biomass during loading and unloading phases.

25 A study was performed to determine the evolution of the composition and molecular weight of the  
26 extracted solution, by varying its residence time within the system. The plant worked homogeneously  
27 and there were no deviations in the characteristics of the liquid product flowing from one unit to the  
28 next.

29 The main objective of this work was to verify the possibility of using the plant set-up for an industrial  
30 process by comparing two scales.

31

32 **Keywords**

33 Pilot plant; hemicelluloses; fractionation; biorefinery; hydrothermal.

34

## 35 **1. Introduction**

36 Hemicellulose is a biopolymer naturally synthesized by most of the existing trees and plants; its  
37 presence in all lignocellulosic biomasses with percentages between 15 and 35% makes hemicellulose  
38 the second polysaccharide with the highest abundance in nature after cellulose (Gatenholm et al., 2004).  
39 Unlike cellulose, which has technological interest in many fields such as plastic, pharmaceutical and  
40 paper industry, hemicellulose applications are being investigated and the interest for this polymer is  
41 growing in recent years.

42 It was demonstrated that hemicellulose polysaccharides have interesting properties in the manufacturing  
43 of food additives (Masutake et al., 1994), emulsifiers (Nakamura et al., 2000), plastic film for the  
44 protection of foods (Gatenholm et al., 2008) or superabsorbent hydrogels (Zhang et al., 2015). The  
45 monomers that constitute hemicellulose are also of great interest. Xylans, present mainly in hardwoods,  
46 grasses and straws, are mainly composed by xylose, precursor of high-value products such as xylitol or  
47 furfural. Xylitol is produced by hydrolysing xylans and subsequently hydrogenating the xylose  
48 monomers. The use of xylitol as a substitute to traditional sugar can help in preventing tooth decay and  
49 promote remineralisation of small lesions (Makinen et al., 1995). It can even be of help for osteoporosis  
50 prevention (Mattila et al., 2002). Furfural, a dehydration product of xylose, is used as a solvent in  
51 petrochemical industry to extract dienes (such as those used to produce synthetic rubber) from other  
52 hydrocarbons and it is also an intermediate product in the production of solvents such as furan  
53 (Hoydonckx et al., 2000).

54 Because of its non-crystalline morphology, in contrast to that of cellulose, hemicelluloses are generally  
55 broken and degraded during conventional pulping processes. However, thanks to biomass pre-  
56 treatments and novel separation processes it is possible to obtain and isolate hemicellulose oligomers  
57 with suitable characteristics to fabricate several specialised materials.

58

59

60



## 61 *Methods for extraction of hemicelluloses*

62 Traditional methods used for the extraction of hemicelluloses from biomass involve the use of mineral  
63 acid solvents. For instance, acid hydrolysis with concentrated H<sub>2</sub>SO<sub>4</sub> or HCl is an effective way to  
64 extract and convert approximately 100 % of cellulose and hemicellulose into monomers. Yet, special  
65 equipment resistant to corrosion and a recover of acid after hydrolysis are required under such  
66 conditions.

67 Dilute acid hydrolysis is a wide used method. H<sub>2</sub>SO<sub>4</sub> or HCl with concentration in the range of 2-5 %  
68 are generally used (pH close to 0), at 160 to 230°C. This treatment extracts hemicellulose from biomass  
69 effectively and hydrolyses it into monomeric sugars and often into degradation product such as furfural  
70 or 5-hydroxymethylfurfural (Ur-Rehman et al., 2015). These processes are suitable if the main purpose  
71 is to obtain a high yield of monosaccharides. Nonetheless, if the aim is to obtain oligomers of  
72 hemicellulose for fabrication of specialised materials such as gel or plastics, other pre-treatments are  
73 required.

74 Hydrothermal treatment is a technique with several advantages over the methods previously mentioned.  
75 Depolymerisation of oligomers and degradation of monomers is notably reduced. Furthermore, as the  
76 unique solvent employed is water, the environmental impact is reduced. Pressurized water at  
77 temperatures above 120 °C is subjected to an ionization process forming H<sub>3</sub>O<sup>+</sup> ions that induce the  
78 partial depolymerisation of hemicellulose. Hemicellulose has acetyl groups alternatively embed in its  
79 pentose-hexose structure. Cleavage of acetyl groups consequently leads to the further formation of  
80 H<sub>3</sub>O<sup>+</sup> ions that catalyses the depolymerisation of hemicellulose. This process, called autohydrolysis, fits  
81 very well when the main purpose is to obtain hemicellulose oligomers with higher selectivity over  
82 monomers formation and monosaccharide degradation (Garrote et al., 1999).

83

## 84 *Hydrothermal extraction plants*

85 There is a huge number of examples regarding hydrothermal hydrolysis of biomass at laboratory scale.  
86 In most cases, the process is carried out in batch reactors (dos Santos Rocha et al., 2017; Garrote et al.,

87 2001b; Lu et al., 2017; Sánchez-Bastardo et al., 2017), being a minor number of researches conducted  
88 with semi-continuous flow-through reactors (Gallina et al., 2016; Tanaka et al., 2012; Yedro et al.,  
89 2015a) or continuous reactors (Cantero et al., 2015; Kazachkin et al., 2014).

90 Flow-through pre-treatments were recognised to be the best in terms of extraction efficiency and energy  
91 economy: thanks to a more effective mass transfer, they allow highest extraction yields respect to batch  
92 reactors, and they do not require extreme milling of biomass and suspension pumping such in the case  
93 of continuous-flow reactors.

94

95 Pilot plants for hydrothermal extraction of hemicellulose are not many. One example is the Integrated  
96 Biomass Utilization System (IBUS) which converts biomass into sugars and lignin, and then to ethanol  
97 and energy, by using three reactors (Larsen et al., 2008; Petersen et al., 2009; Thomsen et al., 2008).

98 This system uses a particle pump to move the biomass into the high-pressure systems.

99 In a previous project developed by our research group, a laboratory-scale flow-through reactor was  
100 developed, operated and optimised (Gallina et al., 2016; Yedro et al., 2015a; Yedro et al., 2015b). That  
101 rig, similarly to most of the existing lab plants, including pilot plant reactors (Kilpeläinen et al., 2014;  
102 Reynolds et al., 2015), consisted of a stainless steel tube filled with biomass, axially crossed by a  
103 constant flow of pressurized hot water.

104 One of the main difficulties in this kind of system was to properly load the raw material and discharge  
105 the spent solid contained in the reactor once the extraction process was completed. In fact, the wet  
106 biomass swells and agglomerates inside this kind of reactors, forming a compact structure difficult to  
107 remove without stopping the operation and being necessary to open the reactor.

108 Sometimes brute force methods, such as drilling, are necessary to extract the compacted-swollen spent  
109 solid. In an industrial context, this implies long periods of inactivity and consequently an economic  
110 disadvantage. An example can be found in delayed coking, where the coke vessels operate in semi-batch  
111 and high pressure water cutting is needed to recover the final coke product (Predel, 2000).

112 A good technique to solve this problem is to introduce biomass into a cartridge, which can be inserted  
113 and removed quickly, as in the system designed by Smirnova et al. (Reynolds et al., 2016; Reynolds et  
114 al., 2015; Zetzl et al., 2012).

115 Results obtained with our laboratory-scale installation, drove our group to design a semi-continuous  
116 pilot plant. Initially, a scaled-up reactor was built, with a volume 73 times bigger than our laboratory  
117 unit, where biomass could be replaced thanks to the insertion and removal of a cartridge through a ball  
118 valve placed on the bottom of the reactor.

119 The system was then integrated with some characteristics of a batch-wise cascade reactor located in Åbo  
120 Akademi (Finland) (Grénman et al., 2011; Rissanen et al., 2014a). That system consisted of 5 Parr units  
121 containing biomass, in which a flow of water circulated in closed loop. Each unit could be excluded  
122 from the system thanks to a valve system that allowed to deviate the water flow.

123 After further improvements, we designed a plant consisting of five semi-continuous reactors, each one  
124 working in series with the others or with the option of being excluded from the system. A manifold of  
125 valves and cartridges, which will be explained later in this work, made possible to extract hemicellulose  
126 from biomass without needing to stop the plant. It was even possible to cool down and disassemble the  
127 reactors separately during the loading and unloading phases, thus minimizing downtime. The extraction  
128 process could be carried out in a pseudo-continuous way, replacing rapidly the raw material and entirely  
129 recollecting the spent solid.

130 In the first part of this paper, we will focus on the scale-up of a laboratory equipment, we will discuss  
131 the scale-up criteria and how we verified the efficiency of the scale up.

132 Subsequently, we will explain how the pilot reactor was implemented to become a multistage flow-  
133 through reactor.

134 The system was then comprehensively tested by a temperature study, analysing the yield of  
135 hemicellulose extraction and its molecular weight. This operation allowed to study the variations in the  
136 characteristics of extracted hemicelluloses varying the residence time of the liquid product within the  
137 system. It was also studied the depolymerisation and degradation of oligosaccharides varying their

138 residence time inside the plant. The objective of this work was to create a system able to operate in a  
139 continuous and fast way, allowing to obtain a product with constant characteristics and composition, and  
140 being considered for an industrial production of hemicellulose by hydrothermal extraction.

## 141 **2. Experimental**

### 142 **2.1 Raw material characterization**

143 *Catalpa bignonioides* wood used as the main raw material in all the experiments was originated from  
144 Valladolid (Spain). This species was chosen for its abundance in the Castilla y Leon area and the  
145 easiness to find pruning residues.

146 Dry wooden branches were grinded with a chipper, obtaining wood chips with variable particle size  
147 between 0.6 and 3.5 cm (showed in Supporting Information). They were kept in a dry room inside  
148 closed bags until the day of the tests. No bark removal was carried out, since the goal was to start off  
149 from low cost biomass, which had undergone a minimum number of pre-treatments.

150 The composition of the raw material in terms of structural carbohydrates, extractives, ashes, humidity  
151 and lignin were determined according to the standard methods published by National Renewable Energy  
152 Laboratory (NREL) (Hames et al., 2005; Sluiter et al., 2008a; Sluiter et al., 2008b; Sluiter et al., 2008c).

153 Dried biomass was treated with water in a Soxhlet equipment, in order to remove the water soluble  
154 extractives, and lately with ethanol to remove remaining extractives. 300 mg of dried and free-  
155 extractives solid were hydrolysed in 3 mL of 72% wt sulphuric acid solution for 60 min at 30 °C. The  
156 mixture was diluted using 84 mL of Milli-Q water and heated in autoclave at 120 °C for 60 min. Solid  
157 was separated from the liquid solution by vacuum filtration and placed in a muffle at 550 °C for 24 h.

158 The remaining residue was weighted before and after this step to calculate the insoluble lignin and the  
159 ash content of the sample. A liquid aliquot was analysed with UV-Vis spectrophotometer at 320 nm  
160 with extinction coefficient of  $34 \text{ Lg}^{-1}\text{cm}^{-1}$  (Sun, Cao, Li, Xu, & Sun, 2014) to calculate the amount of  
161 soluble lignin. Another liquid aliquot was neutralized to pH range 6 to 7, filtered using a 0.2  $\mu\text{m}$   
162 membrane and analysed by HPLC to determine the carbohydrates composition.

163

164 **2.2 Analytical methods**

165 **2.2.1 Analysis of liquid samples composition**

166 The total number of compounds contained in the liquid samples was determined by hydrolysing  
167 oligomers extracted during the process. 0.8 mL of sulphuric acid (72%) and 15 mL of Milli-Q water  
168 were added to 5 mL of liquid samples. This solution was autoclaved at 121 °C for 1h. Prior to the HPLC  
169 analysis, liquid samples were filtered (Pore size 0.22 µm, Diameter 25 mm, Nylon; FILTER-LAB)  
170 (Sluiter et al.).

171 Original liquid samples (before acid hydrolysis) obtained by the extraction process were also filtered  
172 and analysed with HPLC.

173 The column used for the separation of the compounds was SUGAR SH-1011 Shodex at 50.0 °C with a  
174 flow of 0.8 mL/min, using a solution of 0.01N of sulphuric acid and water Milli-Q as mobile phase. A  
175 Waters IR detector 2414 and Waters dual λ absorbance detector 2487 (210 nm and 254 nm) was used to  
176 identify the sugars and their derivatives.

177 The calibration reagents used for HPLC analysis were: cellobiose (+98%), glucose (+99%), fructose  
178 (+99%), glyceraldehyde (95%), pyruvaldehyde (40%), arabinose (+99%), glycolaldehyde (+98%), 5-  
179 hydroxymethylfurfural (99%), lactic acid (85%), formic acid (98%), glucuronic acid (99%), mannose  
180 (+99%), xylose (+99%), galactose (+99%), rhamnose (+99%), galacturonic acid (+99%), furfural  
181 (+99%), acetic acid (+99%), all of them purchased from Sigma-Aldrich and used without further  
182 modification.

183 For the analysis of sugars, sulphuric acid (96%) and calcium carbonate (+ 99 %), purchased from  
184 Panreac were used.

185

186 **2.2.2 Analysis of molecular weights**

187 Molecular weight of the hemicelluloses in the liquid extract was determined by Size Exclusion  
188 Chromatography (HPLC-SEC). The column used was a GPC column (SB-804 HQ; Shodex) protected  
189 by a guard column (SB-G; Shodex) at 35 °C with a flow rate of the mobile phase (NaNO<sub>3</sub> 0.1 M +

190 NaN<sub>3</sub> 0.02% in Milli-Q water) set at 0.5 mL min<sup>-1</sup>. A Waters IR detector 2414 was used for the  
191 determination of the molecular weight of the extracted hemicelluloses. Calibration curve was obtained  
192 with a set of eight pullulan standards (STANDARD P-82; Shodex) ranged between 6.1 and 642 kDa of  
193 average molecular weight, dissolved in milli-Q water.

194

### 195 **2.2.3 Determination of pH**

196 The pH of the extracted solution (hydrolysate) was measured online, with intervals of 1 minute in the  
197 liquid outlet. An electronic pH-meter (Crison CRI10123.99) was used.

198

### 199 **2.3 Raw Material Composition**

200 Humidity, determined after drying a sample of wood in a convection oven at 105 °C, was calculated to  
201 be 14% wt. Dry raw material was composed by: 20.8% of extractives, 16.2 % of lignin, 0.3% of ashes,  
202 32.3% of cellulose (glucose), 23.7% of hemicelluloses (xylose, arabinose and acetic acid) and 6.8% of  
203 pectins (galacturonic acid). Amounts of single compounds are represented in Table S1 in the Supporting  
204 Information.

205 Along the manuscript, xylose will be chosen as the monosaccharide representing the hemicelluloses, as  
206 xylans constitutes the main hemicelluloses proceeding from hardwoods like *Catalpa bignonioides*  
207 (Yoon et al., 2016).

208 Wood proceeding from branches and bark is known to have a great amount of extractives respect to  
209 stem wood (Krutul et al., 2014; Nurmi, 1992) and these results were confirmed by our composition  
210 analysis.

211 46.3% of extractives was water soluble, while the remaining 53.7% was soluble in ethanol. This  
212 proportion is similar to that found from other authors in other species like *Eucalyptus globulus* (Morais  
213 & Pereira, 2012).

214 12.9% of the water soluble extractives were monosaccharides and sugar acids (32.0% glucuronic acid,

215 28.0% glucose, 14.0% xylose and 26.0% arabinose).

## 216 **2.4. Experimental set-up and operation**

217 This section is explained later within section 3 (lab and pilot scale for 1 reactor) and section 4 (dedicated  
218 to the pilot 5-reactor system manifold), as the scale-up was one of the objectives of the work.

219

220

## 221 **3. Reactors scale-up: from lab scale to pilot scale**

222 The set-up and the operational tips of both the laboratory scale and pilot scale reactors are described in  
223 this section; the criteria for the scale up is explained in detail. A discussion on the characterization of the  
224 hydrolysate effluents produced with these systems is included at the end of the section, comparing the  
225 results and assessing the effectiveness of the scale-up.

226

### 227 **3.1 Laboratory scale flow-through reactor**

228 Laboratory-scale reactor is schematized in Figure 1. A PU-2080 HPLC pump (P-01) took water from a  
229 deposit (D-01) and propelled it through a concentric tube heat exchanger working in counter-current  
230 mode (E-01, 2m length, 1/8"-1/4"). Then water passed through a pre-heater (H-01, 200 cm of 1/8" SS  
231 316 pipe, electrically heated by two electric resistors of 300 W) which ensured a uniform temperature at  
232 the reactor inlet. Water entered from the top of the reactor (R-01), consisting of a SS316 pipe (R-01, 38  
233 cm length, 1/2" O.D., 0.38" I.D.), charged with wood chips. Reactor was covered on the top and on the  
234 bottom by two metallic filters, in order to avoid the loss of solid particles during the experiments. The  
235 reactor was heated by three electric clamp resistors of 300 W/each, placed axially along a machined  
236 aluminum bar with 5.08 cm O.D. Outlet flow passed through a concentric tube heat exchanger E-01  
237 (preheating the inner flow).

238 Pressure was controlled by a Go-back pressure valve (BPV-01) installed at the liquid outlet. The outer  
239 flow pH was measured online using an electronic pH-meter (Crison PH 29).

240

### 241 **3.1.1 Operation of laboratory reactor**

242 Wood from *Catalpa bignonioides* was used as raw material; 6 g of chips with a medium particle size of  
243 about 0.6 cm were loaded in the reactor R-01. A cold liquid pressure test was made before each  
244 experiment: cold water was circulated through the system for 5 min in order to check the presence of  
245 leaks and to ensure the wetting of the wood. After that, the pump was switched off, while electric heater  
246 and clamp resistances placed along the reactor were set at a temperature of 20 °C above the operating  
247 temperature. When the temperature was reached, water was pumped through the system, starting the  
248 experiment (time 0). Experiments carried out with other raw materials in the laboratory scale reactor  
249 (Gallina et al., 2016), indicated that the optimal flow to get a good mass transfer with the minimum  
250 amount of water, corresponded to 3.5 mL/min; the same flow rate was used in this work.

251 The first sample (time 0) was collected as soon as the first drop of liquid came out from the system.  
252 Water reached the operating temperature in about 7 min from the taking of the first sample. Samples  
253 were collected after 5, 10, 20, 30, 40, 60 and 90 minutes from the beginning of the operation. At the end  
254 of the process, the heating was turned off and fresh water was passed through the system to cool down  
255 the reactor. When a temperature of 50 °C was reached, the pump was switched off, the system was  
256 depressurized and water was let to flow out. The reactor was finally dismounted, placed in a mechanical  
257 grip and, using a steel punch and a hammer, the wet solid was removed.

258

### 259 **3.2 Scale-up to single stage pilot reactor**

260 The scale-up of the laboratory reactor was made by following the criteria indicated in Table 1. The  
261 volume of the pilot reactor was 2000 mL, 73 times bigger than the laboratory-scale reactor. Geometrical  
262 similarity between the two systems was maintained, by keeping constant the ratio between length and  
263 internal diameter (L/ID). An optimal flowrate of 250 mL/min was set in the pilot system to have the  
264 same superficial velocity of water as in the lab-system. The porosity of the bed was also preserved in  
265 both reactors, to have the same residence time of the liquid (aprox. 6.0 min).

266



267 One of the objectives of this work was to minimize the difficulties of replacing the biomass in the  
268 extractor in order to ease the operation and maintenance of the pilot reactor. At laboratory scale, there  
269 are many methods that can be used to remove the spent solid (some of them are ‘brute-force’ methods,  
270 e.g. drilling, pushing out with compressed air, etc.) that are difficult to use at pilot or industrial systems.  
271 To facilitate the replacement of biomass in the flow-through reactor, a cartridge mode has been  
272 implemented.

273 The reactor unit consisted of 3 main parts (Figure 2a):

- 274 • An open cylinder, constructed with a wire mesh (7) that could be opened longitudinally. Internal  
275 diameter of the cylinder was 4 cm and length was 159 cm.
- 276 • Two stainless steel cylinders with the same diameter (4 and 5), one of which (5) had several  
277 orifices (with a diameter of 1 mm) at the bottom (6), working as a filter. A glass wool layer  
278 could be placed over the holes to decrease the dimension of the voids.

279 The wire mesh was inserted between the two cylinders, forming a cartridge, which was filled  
280 with biomass. The inner diameter of the two cylinders was 4.3 cm; so that the mesh adhered  
281 perfectly to the walls. The cylinders thickness was 2 mm.

- 282 • An outer stainless-steel cylinder (2) with two opening (A and B) closed at the upper end with a  
283 mechanized flange (1) with an opening (C), and with a ball valve (3) screwed on the lower end.  
284 Internal diameter of the valve and of the cylinder was 5.1 cm, so that the cartridge could be  
285 introduced from the bottom and inserted completely into the system.

286 This system greatly facilitated the replacement of biomass, which could be removed from the system by  
287 simply opening the valve and pulling out the cartridge. The longitudinal opening of the wire mesh,  
288 moreover, reduced the effort required to remove the wet biomass from the cartridge.

289 A flow diagram of the single-stage pilot system is represented in Figure 2b. A constant flow of water  
290 was drawn from a vessel (D-01) and propelled with a Tuthill DGS.68 pump (P-01) through the external  
291 tubes of a system composed by three concentric tube heat exchangers E-01 to E-03 (18 m total length,

292 1/4" internal tube-3/8" external tube) and then through an electric heater H-01 with a maximum power  
293 of 5 kW.

294 Given the high power of the heater, a special procedure was designed to avoid the overheating of this  
295 unit. Unlike in the laboratory scale system, where the heater was left over for a time heating with no  
296 contact with water, in the case of the pilot plant, water flowing through a coil was in constant contact  
297 with the heater wall.

298 A three way valve (3V-01) was placed between the heater and the reactor, which could direct the liquid  
299 flow to the inlet of the reactor (placed on the top) or out of the system. A ball valve (V-01) was placed  
300 just after the outlet of the reactor.

301 During the preheating phase, the 3-way valve was turned so that water did not enter the reactor, while  
302 valve V-01 was closed to prevent water from returning in the reactor through its output.

303 Water stream entered subsequently in the inner part of the concentric tube heat exchangers (E-01 to E-  
304 03), where it was cooled down by the feeding water flowing countercurrent in the external part, before  
305 leaving the system through a Go-back pressure valve (BPV-01). In this way it was possible to pre-heat  
306 the water to the desired temperature before introducing it into the reactor.

307 Reactor R-01 was homogeneously coated with four clamp resistors, which power was 250 W/each (total  
308 nominal power was 1 kW).

309 The entire system was thermally insulated with a layer of 2 cm glass wool, protected with aluminum  
310 foil.

311 Temperature of water entering and leaving the reactor was measured with two thermocouples placed at  
312 the inlet and at the outlet of R-01.

313 A flow-meter was placed after the pump to measure the liquid flow rate. An online pH-meter was placed  
314 after the Go-back pressure valve to measure the pH of the solution produced by the process.

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### 321 **3.2.1 Operation of single stage pilot reactor**

322 The cartridge was assembled by introducing the wire mesh between the two half cylinders then. 250 g of  
323 chips with a medium particle size of about 2.0 cm were placed inside. The quantity and size of the  
324 particles was selected to maintain the same porosity as in the laboratory system.

325 At the beginning of the operation, the cartridge was introduced into the reactor through the bottom, and  
326 the ball valve (3) was closed.

327 A flow of cold water was pumped inside the reactor from the top, until it was completely filled. During  
328 this procedure, ball valve V-01 was closed to keep the water inside the reactor.

329 Another ball valve (V-02), was connected to an outlet (B) of the reactor placed on the top. This valve  
330 was opened during the filling procedure, and was connected to a plastic container (C-01). When water  
331 wet the container the reactor could be considered completely filled. At this moment, the ball valve V-02  
332 was closed and the valve (V-01) was opened, letting water exiting from the reactor.

333 A cold liquid pressure test was made, by increasing the pressure of the system to 17 bar. Subsequently,  
334 the three-way valve was switched and the valve (V-01), so that the flow of water by-passed the reactor.

335 Heater was turned on to heat-up the liquid flow 20 °C above the operating temperature; water contained  
336 in the reactor was preheated to 95 °C through the clamp resistances which wrapped it around.

337 Temperature of water inside the reactor was set to a value minor than 100 °C to avoid the extraction of  
338 structural carbohydrates.

339 When the water reached the desired temperature, the three-way valve was switched, and the flow was  
340 directed into the reactor; at the same time, the ball valve (V-01) was opened, letting water exiting the  
341 reactor.

342 Time 0 (zero) was set at this moment and sample 0 (zero) was collected from the system outlet. Other  
343 samples were collected after 5, 10, 20, 30, 40, 60 and 90 minutes from the starting of the operation.

344 At the end of the process, the heating system was turned off and fresh water was passed through the  
345 system to cool it down. When a temperature of 50 °C was reached, the pump was switched off, the  
346 system was depressurized and water was let to flow out. The ball valve (3) on the bottom of the reactor  
347 was opened and the cartridge containing biomass was extracted and dismantled to remove the solid.

348

### 349 **3.3 Comparison between results obtained with laboratory and pilot reactor**

350 Hydrothermal extraction was carried out with the laboratory scale and with the pilot reactor at 160 and  
351 170 °C. Figure 3a represents the cumulative yields of total xylose, produced with the two systems after  
352 the extraction process and the acid hydrolysis of the solution obtained. Yield was calculated as the ratio  
353 between the total mass of the xylose extracted and the total mass of xylose in the wood contained in the  
354 reactor at the beginning of the experiments.

355 A cumulated yield of 33.9% was obtained after 90 minutes of operation, using the laboratory scale  
356 reactor at 160 °C; a yield of 35.7% was reached with the pilot reactor at the same temperature and  
357 extraction time.

358 In experiments conducted at 170 °C, the yields increased in both cases, reaching final values of 38.8%  
359 and 41.7% when using the lab-scale and the pilot reactors respectively. Yields of all the compounds  
360 extracted are represented in Table S2 in the Supporting Information.

361 The positive effect of temperature in increasing the extraction yield is well known and widely  
362 documented (Nitsos et al., 2016; Qian et al., 2015; Yedro et al., 2017). However, in this work, the main  
363 concern was to identify the differences that occurred when the process was scaled-up.

364 Figure 3b shows the percent deviation between the yields obtained with laboratory scale and pilot scale  
365 reactors, at 160 and 170 °C.

366 At both temperatures, the deviations followed a decreasing trend: it was 100% at time 0, after 5 min it  
367 decreased to 64.0 % in the experiment at 170 °C and 44.0 % at 160 °C. After 20 min the deviations  
368 reached negative values and then tended to 0.

369 The “100% deviation”, found at time 0 was associated to the fact that while with the laboratory system  
370 no xylose was produced, small amounts were detected in the solutions proceeding from the pilot plant at  
371 the very beginning of the extraction. The presence of this monosaccharide, rather than to hemicelluloses  
372 breakdown, was almost certainly due to the free sugars contained in the biomass, which can be extracted  
373 at temperatures below 100 °C (Alañón et al., 2009). Considering the composition of water soluble  
374 extractives, determined as explained in the section 2.3, it was calculated that xylose extracted at time 0  
375 with the pilot reactor, corresponded to the 25.8% of the monomeric xylose in the raw material. This  
376 initial yield of monomeric xylose is the same in the experiments carried out at 160 and 170 °C, as  
377 temperature of water inside the reaction chambers was always set at 95 °C (as explained in section 3.2.1)  
378 at the beginning of the operations.

379 The deviations between the yields obtained with pilot scale and laboratory scale plants were higher  
380 during the first 10 minutes of extraction. This behaviour could be due to: (1) the different temperature  
381 profile followed during the process, as represented in the Supporting Information and (2) the differences  
382 in the time scale due to the piping, which length change have more influence at the beginning of the  
383 process, when the slope in the extraction curve is high.

384 The warming up to the operating temperature occurred faster in the pilot plant (3 minutes less) in  
385 comparison with the laboratory scale system. This delay, due to the different method employed in the  
386 heating, is responsible for the initial deviation between the yields obtained in the two systems. In the  
387 pilot plant, the wood particles were completely submerged in water before starting the process. Then,  
388 they had undergone to a preheating to 95 °C, during which their structure weakened and released part of  
389 the water-soluble compounds.

390 On the other hand, in the laboratory scale plant, part of the water that was injected during the pressure  
391 control step left the system during the preheating phase, when the pump was switched off. Thus, chips  
392 impregnation was less effective than in the pilot reactor, leading to a less breakdown of the wood  
393 particles (Bäckström et al., 2016; Malkov et al., 2001). The lower amount of water inside the reactor  
394 also resulted in a greater inertia to reach the operating temperature, as it took about 5 min to completely  
395 fill the system and then warm it up.

396 Figure 3c represents the cumulative yield of monomeric xylose obtained at different times of extraction  
397 in the pilot and laboratory scale plant. As it is known, temperature favours the extraction of  
398 hemicellulose oligomers and increases hydrolysis and formation of monosaccharides (Rogalinski et al.,  
399 2008), which is in accordance with the results shown in Figure 3c. An increasing in temperature from 160  
400 to 170 °C meant an increasing in the yield of around 2%.

401 Comparing subplots in Figure 3 it is possible to deduce that the yield was slightly higher in the pilot  
402 reactor than in the lab scale in general. The higher differences appeared during the first 20 minutes of  
403 extraction, attributable mainly to the different preheating procedure. No degradation product, such as  
404 furfural (from xylose cyclodehydration) or 5-hydroxymethylfurfural (5-HMF from glucose  
405 dehydration), were detected in the experiments, suggesting that the temperatures and residence times  
406 tested were not strong enough to decompose the monosaccharides.

407

#### 408 **4. Multistage pilot flow-through reactor**

409 After establishing the effectiveness of the scale-up, several modifications were made in the pilot plant,  
410 aimed at conducting the hydrothermal extraction in a continuous and rapid way: the single stage flow-  
411 through pilot plant was implemented to become a multistage flow-through pilot plant.

412 This section will explain the setup of the manifold-system and the standard operating mode for which it  
413 was designed. Finally, a study was conducted to verify the possibility of a continuous production of  
414 hemicellulose.

415

#### 416 **4.1 Set-up of multistage reactor**

417 The complete layout of the plant is shown in Figure 4. Five reactors with the same geometry and  
418 operating principle as the one shown in Figure 2a, were connected in series (R-01 to R-05). A three-way  
419 valve (3V-01 to 3V-05) was placed before each reactor (manifold). Each valve could divert the flow to  
420 the reactor inlet or to the next 3-way valve, by-passing the reactor. Ball valves (V-01, V-03, V-05, V-07  
421 and V-09) placed after the reactors could be closed to prevent water return.

422 Right after the outlet of each reactor, needle valves (NV-01 to NV-05) connected to concentric tube heat  
423 exchangers (ES-01 to ES-05) allowed for the withdrawn of liquid samples from the reactors. Liquid  
424 solutions flowed through the internal pipe of heat exchangers (1 m length, 1/8" internal tube- 1/4"  
425 external tube) when needle valves were open, and were cool-down by tap water flowing through the  
426 external pipe.

427 A centrifugal pump Marathon Electric 5KH36 (P-02) took fresh water from a vessel (V-02) and  
428 transferred it through a pipe, ball valves (V-11 to V-15) could be opened to let water enter into the  
429 reactors. Valves V-02, V-04, V-06, V-08 and V-10, connected to plastic containers (C-01 to C-05)  
430 worked as level control system: reactors were filled when first drops wetted the containers.

431 Pump P-01 (Tuthill DGS.68) transferred water from a vessel (D-01), to the external section of three  
432 concentric tube heat exchangers E-01 to E-03 (18 m total length, 1/4" internal tube-3/8" external tube)  
433 and then through an electric heater H-01 with a maximum power of 5 kW.

434 Each reactor was coated with four clamp resistors with a total power of 1 kW/reactor. The water left the  
435 system after being cooled down in heat exchangers E-01 to E-03, and depressurized through a Go back-  
436 pressure valve (BPV-01). A pH meter placed after the valve measured online the pH of the final solution  
437 every minute. The whole system was thermally insulated with a layer of glass wool (about 2 cm)  
438 covered with aluminum foil.

439

#### 440 **4.2 Standard operation of multistage reactor**

441 In a standard operation, three reactors (R-01, R-02 and R-03) were charged with biomass by inserting  
442 the cartridge, as explained in section 3.2.

443 *System preparation*

444 Valves (V-01, V-03, and V-05) placed after the outlet of the reactors were closed.

445 Centrifugal pump P-02 was turned on, valves V-11, V-12, and V-13 were opened to let water enter into  
446 the reactors (dotted arrows), and valves V-02, V-04 and V-06 were also opened to let water flow-out to  
447 plastic containers (C-01 to C-03) when reactors were completely filled.

448 When reactors were filled, the pump P-02 was switched off, while valves V-11, V-12, V-13 and V-02,  
449 V-04, V-06 were closed.

450 At this time, pump P-01 was turned on and set to the desired flow rate, feeding the system with fresh  
451 water.

452 3-way valves 3V-01, 3V-02 and 3V-03 were set in order to let water enter into the reactors, valves V-  
453 01, V-03, and V-05 were opened to let water flow out.

454 The direction of the flow is represented by the continuous arrows in Figure 4: water flowed through the  
455 external pipes of heat exchangers (E-01 to E-03), through the spirally wound pipes around the heater H-  
456 01, entered reactor R-01 from the top and left it from the bottom, passing through the biomass  
457 contained.

458 3-way valve 3V-02 and 3V-03 connected reactors R-02 and R-03 in series with R-01.

459 Water by-passed reactors R-04 and R-05, entered into the internal pipes of heat exchangers (E-01 to E-  
460 03) and left the system.

461 *Pressurization and warming up*

462 Pressure of the system was increased to 17 bar and a cold liquid pressure test was made, stabilizing at  
463 the same time the liquid hold-up of the system.

464 When the liquid flow-rate (measured through a flow-meter placed after pump P-01) was constant, 3-way  
465 valves (3V-01 to 3V-03) were switched to make water by-pass the reactors R-01 to R-03, while valves  
466 V-01, V-03 and V-05 were closed to avoid water return.



467 Circulating water flow was pre-heated to 20 °C above the operating temperature (by turning on the  
468 heater H-01), while water contained in reactors R-01 to R-03 was heated to 95 °C by turning on the  
469 clamp resistors which covered the reactors.

#### 470 *Extraction*

471 After reaching the desired temperature, 3-way valves 3V-01 to 3V-03 were switched and hot water  
472 flowed through the three reactors, extracting soluble compounds from biomass.

473 Liquid samples could be withdrawn from each reactor at regular times, by opening the needle valves  
474 (NV-01 to NV-03).

475 When it was desired to interrupt the extraction in one of the reactors, i.e. R-01, the unit was isolated  
476 from the system, by switching the 3-way valve that preceded it (3V-01) and closing the ball valve V-01  
477 simultaneously. Needle valve (NV-01) was opened to remove the water contained in the reactor and to  
478 depressurize it. Ball valve on the bottom of the reactor was opened and the cartridge containing the  
479 biomass was discharged.

480 Meanwhile, a new cartridge could be loaded in another reactor (i.e. R-04), which was filled with fresh  
481 water through pump P-02 and warmed up to 95 °C. When extraction process ended in reactor R-01,  
482 circulating solution was let to enter in reactor R-04, by switching the 3-way valve 3V-04 and extracting  
483 compounds from the new biomass.

484 This operation could be repeated for each unit in the system, in this way each reactor could be integrated  
485 into the extraction process or could be by-passed. The system allowed continuous operation: each time  
486 that a reactor was stopped to replace the feedstock, another reactor could operate. Removal of the raw  
487 material was easy and fast, and it could be done without the necessity of disassembling the extraction  
488 unit. Moreover, all the solid could be recollected at the end of the extraction.

489

#### 490 **4.3 Extraction in series in the multistage pilot reactor**

491 To assess the ability of the plant to work in a continuous way, with the possibility to quickly replace the  
492 biomass, as described in the paragraph 4.2, it was necessary to verify that the reactors worked in a

493 homogeneous manner and that there were no alterations in the composition of the effluent produced by  
494 the individual reactors.

495 Another important matter to study was the evolution in the composition of the extracted products,  
496 varying their residence time within the system, to understand if there was degradation or  
497 depolymerization of the hemicelluloses extracted when flowing from one unit to the other.

498 Experiments were carried out at four different temperatures (140, 150, 160 and 170 °C), using three  
499 reactors connected in series, with a constant water flow of 15 L/h. Temperature profiles inside the  
500 reactors are depicted in the Supporting Information. The temperature of the water flow over time is  
501 plotted (see SI): before the heater (after preheating in the heat exchangers), after the heater (before  
502 reactor R1), after the reactors (before cooling through the heat exchangers) and at the system outlet  
503 (after cooling). In the system, 78% of the heat was recovered in the pre-heating, while 100% of the heat  
504 was dissipated during cooling.

505 The reactors were loaded with their respective cartridges, each one filled with 250 g of catalpa wood-  
506 chips with an average particle size of 2 cm. Reactors R1 to R3 were then pre-filled with distilled water  
507 and pre-heated to 95 °C.

508 A water stream was also preheated to about 20 °C above the operating temperature (thus 160, 170, 180  
509 and 190°C) and, at time 0, it was injected inside the first reactor; which outlet flow was entering into the  
510 second unit, connected in turn with the third unit in series.

511 In about 3 minutes, the three reactors reached the same operating temperature with a constant water  
512 stream flowing through them and preheating the feeding water before exiting the system.

513 The whole operation lasted 90 minutes. Liquid samples were collected at regular intervals of time (0, 5,  
514 10, 20, 30, 40, 60 and 90 minutes) from each of the three reactors. After 90 min, the three units were  
515 isolated from the system. Then, they were emptied and the cartridges containing the biomass were  
516 removed. Pretreated wood proceeding from each unit was entirely collected, dried in an oven at 105 °C  
517 and finally weighted. No replacement of biomass was made (although the system is ready for a pseudo-  
518 continuous operation).

519 The whole liquid extract proceeding from each experiment was collected in a tank; six vials were filled  
520 with 2 mL of every solution and lyophilized to determine the average concentration of all the  
521 compounds extracted; the difference between this value and the concentration of hemicellulose in the  
522 final solution allowed to calculate the amount of lignin and extractives also resulting from the process.

523 Liquid samples withdrawn from each unit at various residence times were analyzed to determine the  
524 composition, the molecular weight and the ratio between monomers and oligomers contained in the  
525 products.

526 The mass balance was verified by adding the weight of the processed wood remaining at the end of the  
527 experiments and the mass of the total solid extracted during the experiments. The mass of the total solid  
528 extracted was calculated as the concentration of solid in the final solution multiplied by the total volume  
529 of liquid leaving the system during the experiments.

530

#### 531 **4.3.1 Temperature study in multistage pilot reactor**

532 Figure 5a shows the cumulative yields of total xylose obtained in the multistage pilot plant at four  
533 different operating temperatures. Table S3 in the Supporting Information shows the results for all the  
534 compounds.

535 The residence time of liquid in the system was 6 min after crossing the unit R1, 12 min after R2, and 18  
536 min after crossing R3.

537

#### 538 **4.3.1.1 Yields of extracted hemicelluloses**

539 Yields were calculated as the ratios between the mass of total xylose detected after acid hydrolysis of  
540 the liquid samples collected from each reactor and the total xylose contained in the biomass processed  
541 respectively in R1, R2 and R3. Figure 5a shows that, as expected, the highest temperatures led to the  
542 highest yields. Furthermore, the reaction rates increased, since temperature influenced the mass transfer

543 by increasing the diffusion coefficient inside the wood particles and opening the pore structure  
544 (Rissanen et al., 2014a).

545 In addition, at constant temperature, the yields were very similar and the variation was minimal at  
546 increasing the liquid residence time in the system, as the values were very similar in the samples  
547 obtained from the three reactors.

548 Average values of cumulative yield after 90 min of operation were:  $9.3\pm 0.7\%$ ,  $22.0\pm 0.5\%$ ,  $35.1\pm 1.6\%$   
549 and  $40.6\pm 1.4\%$ , respectively, at 140, 150, 160 and 170 °C. The values indicate the average yields and  
550 the standard deviations between R1, R2 and R3. Thus, the experimental error was very low (between  
551 2.2 and 7.9%).

552 During the process, the extracted products accumulated in the effluent flowing from one reactor to the  
553 next. The system operated as a long reactor divided into three equivalent sections where the extraction  
554 took place homogeneously. Moreover, there was no perturbation in the composition of the liquid  
555 effluent flowing from one unit to another.

#### 556 **4.3.1.2 Molecular weight of extracted hemicelluloses**

557 Since the aim of this work was to verify the possibility of using the plant set-up for an industrial  
558 process, it was necessary to check if the effluent was homogeneous not only in terms of product yield,  
559 but also in terms of composition and length of the oligomers.

560 The molecular weight of the outputs from the three units during the process at different extractions  
561 times was then analysed. Values obtained at different temperatures from units R1, R2 and R3, are  
562 depicted in figure 5b. Results are represented in Table S4 in the Supporting Information. In all cases, the  
563 molecular weight exhibited an upward trend during the first 10 minutes of extraction, and then it  
564 remained constant or decreased with the extraction time. An exception occurred in the experiment

565 carried out at 140 °C, wherein the higher molecular weights were obtained after 30 minutes of  
566 extraction.

567 In general, the molecular weight decreased with an increasing in the operating temperature. After 90  
568 minutes of extraction, the molecular weight of hemicellulose oligomers extracted presented values of  
569 4078±23, 2963±142, 1922±95 and 1417±41 Da at 140, 150, 160 and 170 °C, respectively. Experimental  
570 errors were between 0.6 and 5.0%. This behavior was consistent with the results of other authors who  
571 worked with other tree species (Rissanen et al., 2014b; Yedro et al.). An increase in temperature is  
572 responsible for a higher cleavage of the hemicelluloses, thus reducing the size of the oligomers that  
573 detach from the matrix.

574 A similar consideration can be made also by observing the polydispersity index (Mw/Mn) shown in the  
575 Supporting Information, which decreased when increasing the temperature.

576 Hemicelluloses break more intensely and faster at high temperatures, for this reason, smaller and more  
577 homogeneous molecules were solubilized in experiments at 160 and 170 °C.

578 In the experiment conducted at 140 °C, polydispersity index increased from 1.2-1.7 during the first part  
579 of the reaction, reaching a maximum value of 2.5-3.2 after around 30-40 minutes, and then decreasing  
580 down to 2.2-2.4 till the end of the experiment. It seems that the extraction started with removing small  
581 molecules; as reaction time increased, larger molecules were broken, until their molecular weight was  
582 small enough to make them soluble.

#### 583 **4.3.1.2 Acidity of extracted hemicelluloses**

584 As explained so far, the rupture and hydrolysis of the hemicelluloses occurred initially inside the wood  
585 chips, because of the kinetics enhanced temperature and catalyzed by the acetyl groups integrated the  
586 matrix. Subsequently, the oligomers solubilized in the liquid phase, experience a further hydrolysis  
587 catalyzed by the dissolved acetyl groups in the bulk liquid. Higher temperatures favored a greater

588 release and solubilization of acetyl groups, and hence, a stronger and faster hydrolysis of hemicellulose  
589 oligomers. Figure 5c shows the instantaneous yield of acetic acid produced in the system during the  
590 experiments at different temperatures. The amount of acetic acid increased along with temperature,  
591 enhancing the hydrolysis of the oligomers in the liquid phase. Moreover, while the formation of acetic  
592 acid at 140 and 150 ° C was almost linear over time, at 160 and 170 ° C there was a maximum  
593 production within 5 to 20 minutes from the beginning of the process. Therefore, the molecular weight of  
594 the extracted oligomers decreased from the first minutes of reaction at highest temperatures.

595 The variation of acetic acid extracted is reflected also in the pH of the solution leaving the reactor,  
596 measured online every minute. Values of pH are depicted in the Supporting Information; more acidic  
597 solutions were obtained when increasing the operating temperature. At 140 and 150 °C, pH decreased  
598 during the whole reaction, while at 160 and 170 °C it reached a minimum value at around 20 minutes  
599 and then increased slowly.

600 At constant temperature, the molecular weights of the hydrolysate solutions extracted from the three  
601 units had similar values over time. This behaviour can suggest two hypotheses: 1) oligosaccharides  
602 extracted from biomass contained in a unit are further hydrolysed in the next unit, in which extraction of  
603 new oligosaccharides restored the average molecular weight value, 2) there was a simple accumulation  
604 of oligosaccharides and monosaccharides between one unit and the next one, and the oligosaccharides  
605 were not subsequently hydrolysed (when their residence time in the system increased passing through  
606 the next unit).

607 To solve this question, the ratio between the monomeric xylose extracted during the process and the  
608 total extracted xylose in each unit was analysed. In this way, it was possible to determine whether the  
609 extracted oligomers were hydrolysed when flowing from one unit to the next one, as shown next.

610

#### 611 **4.3.1.3 Monomers content in the liquid product**

612 Figure 5d shows that the ratio between monomeric xylose and total xylose increased slightly with the  
613 residence time of the liquid within the system, with an increasing slope when temperature increased.  
614 Values are referred to the cumulative amount of sugars extracted from each unit at the end of the  
615 experiments (after 90 min of operation). The highest difference between the maximum and the  
616 minimum value resulted at 170 °C, where the ratio of monomeric xylose to total xylose was 0.18 after 6  
617 min and 0.21 after 18 min of liquid residence time. Considering the average values of the ratios at the  
618 various temperatures, it can be seen that between 140 and 160 °C, the values were very similar.  
619 However, at 170 °C, there was a general increasing in monomeric xylose compared to the total xylose  
620 extracted. Other authors have found only a slight formation of monomeric xylose from xylans  
621 hydrolysis at temperatures lower than 190 °C and residence time below 20 minutes (Garrote et al.,  
622 2001a; Liu & Wyman, 2004; Lloyd & Wyman, 2003). The phenomena may be due to the temperature  
623 itself, the main responsible of the cleavage of hemicelluloses, and to the presence of acetic acid, which  
624 at temperatures above 160 °C increased its concentration in the aqueous medium catalysing the  
625 breakdown of the solubilized oligomers.

626 Degradation products as furfural or 5-HMF were under the detection limit at the conditions used in these  
627 experiments. Therefore, it can be stated that there was an accumulation of the compounds extracted  
628 from one unit to the next, while a weak hydrolysis of the oligosaccharides occurred with increasing the  
629 liquid residence time, more accentuated at high temperatures.

630 The final mass balance of the four experiments is shown in Figure 6. The errors (the highest error was  
631 below 10.5%) which is in the order of other authors in the field. As the temperature rose, an increasing  
632 in the solubilized material was noticed, both of the hemicelluloses and of the other compounds, mostly  
633 made up of lignin and extractives.

634 The raw material had no ideal characteristics for extracting hemicellulose in a laboratory-level study, as  
635 the extractives and lignin content was high and the particle size was large; however, the material could  
636 be well used in an industrial context where only a minimal number of treatments is necessary, in order

637 to reduce the costs. The disadvantages represented by this choice are reflected in the low purity of the  
638 extract of hemicellulose due to the considerable amount of extractives in the raw material, and in a  
639 lower yield respect to other experiments in which the wood particles were smaller and diffusion  
640 limitations played a minor role (Kilpeläinen et al., 2014; Rissanen et al., 2014a). A possible solution to  
641 improve the process without excessively increasing the operational expenditure (OPEX) could be to  
642 perform a first hydrothermal pre-treatment at a temperature between 120 and 140 °C, during which part  
643 of the undesirable soluble compounds would be eliminated without extracting the hemicellulose. Post-  
644 treatments to concentrate and fractionate the hydrolysate would be also necessary. Although some  
645 improvements are needed in the operations, to obtain products more pure in hemicellulose, the plant  
646 described in this document has proved to be versatile and suitable for its intended purpose and can be  
647 considered in industrial technology.

648

#### 649 **4. Conclusions**

650 In this work, we have established the basic criteria for the scale-up of a process from a 27.5 mL lab  
651 extractor to a 2000 mL pilot extractor for extraction of hemicellulose from biomass with hot pressurized  
652 water.

653 A shape parameter of  $L/D=40$ , with a bed porosity of 0.71 and a liquid residence time of 6.0 min are  
654 efficient for such purpose.

655 From the results obtained, we constructed a manifold of 5-reactors that can operate in series. It was  
656 verified that the reactors can work continuously, without changings in the composition of the product.

657

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- 663 Alañón, M.E., Ruiz-Matute, A.I., Martínez-Castro, I., Díaz-Maroto, M.C., Pérez-Coello, M.S. 2009.  
664 Optimisation of pressurised liquid extraction for the determination of monosaccharides and  
665 polyalcohols in woods used in wine aging. *Journal of the Science of Food and Agriculture*,  
666 **89**(15), 2558-2564.
- 667 Bäckström, M., Jensen, A., Brännvall, E. 2016. Influence of chip presteaming conditions on kraft pulp  
668 composition and properties. *Holzforschung*, **70**(5), 393-399.
- 669 Cantero, D.A., Martínez, C., Bermejo, M.D., Cocero, M.J. 2015. Simultaneous and selective recovery of  
670 cellulose and hemicellulose fractions from wheat bran by supercritical water hydrolysis. *Green*  
671 *Chemistry*, **17**(1), 610-618.
- 672 dos Santos Rocha, M.S.R., Pratto, B., de Sousa, R.J., Almeida, R.M.R.G., Cruz, A.J.G.D. 2017. A  
673 kinetic model for hydrothermal pretreatment of sugarcane straw. *Bioresource Technology*, **228**,  
674 176-185.
- 675 Gallina, G., Cabeza, Á., Biasi, P., García-Serna, J. 2016. Optimal conditions for hemicelluloses  
676 extraction from Eucalyptus globulus wood: hydrothermal treatment in a semi-continuous reactor.  
677 *Fuel Processing Technology*, **148**, 350-360.
- 678 Garrote, G., Domínguez, H., Parajó, J.C. 2001a. Kinetic modelling of corncob autohydrolysis. *Process*  
679 *Biochemistry*, **36**(6), 571-578.
- 680 Garrote, G., Domínguez, H., Parajó, J.C. 1999. Mild autohydrolysis: An environmentally friendly  
681 technology for xylooligosaccharide production from wood. *Journal of Chemical Technology and*  
682 *Biotechnology*, **74**(11), 1101-1109.
- 683 Garrote, G., Domínguez, H., Parajó, J.C. 2001b. Study on the deacetylation of hemicelluloses during the  
684 hydrothermal processing of Eucalyptus wood. *Holz als Roh- und Werkstoff*, **59**(1-2), 53-59.
- 685 Gatenholm, P., Bodin, A., Gröndahl, M., Dammstrom, S., Eriksson, L. 2008. Polymeric film or coating  
686 comprising hemicellulose, Google Patents.
- 687 Gatenholm, P., Tenkanen, M., Cellulose, A.C.S., Division, R.M., Meeting, A.C.S. 2004.  
688 *Hemicelluloses: Science and Technology*. American Chemical Society.
- 689 Grénman, H., Eränen, K., Krogell, J., Willför, S., Salmi, T., Murzin, D.Y. 2011. Kinetics of Aqueous  
690 Extraction of Hemicelluloses from Spruce in an Intensified Reactor System. *Industrial &*  
691 *Engineering Chemistry Research*, **50**(7), 3818-3828.
- 692 Hames, B., Ruiz, R., Scarlata, C., Sluiter, A., Sluiter, J., Templeton, D. 2005. Preparation of Samples for  
693 Compositional Analysis. in: *Laboratory Analytical Procedure (LAP)*.
- 694 Hoydonckx, H.E., Van Rhijn, W.M., Van Rhijn, W., De Vos, D.E., Jacobs, P.A. 2000. Furfural and  
695 Derivatives. in: *Ullmann's Encyclopedia of Industrial Chemistry*, Wiley-VCH Verlag GmbH &  
696 Co. KGaA.
- 697 Kazachkin, D.V., Colakyan, M., Moesler, F.J. 2014. Supercritical hydrolysis of biomass, Google  
698 Patents.
- 699 Kilpeläinen, P.O., Hautala, S.S., Byman, O.O., Tanner, L.J., Korpinen, R.I., Lillandt, M.K.J., Pranovich,  
700 A.V., Kitunen, V.H., Willför, S.M., Ilvesniemi, H.S. 2014. Pressurized hot water flow-through  
701 extraction system scale up from the laboratory to the pilot scale. *Green Chemistry*, **16**(6), 3186-  
702 3194.
- 703 Krutul, D., Zielenkiewicz, T., Zawadzki, J., Radomski, A., Antczak, A., Drozddek, M. 2014. Influence  
704 of urban environment originated heavy metal pollution on the extractives and mineral substances  
705 content in bark and wood of oak(*Quercus robur* L.). *Wood Research*, **59**(1), 177-190.
- 706 Larsen, J., Østergaard Petersen, M., Thirup, L., Wen Li, H., Krogh Iversen, F. 2008. The IBUS Process  
707 – Lignocellulosic Bioethanol Close to a Commercial Reality. *Chemical Engineering &*  
708 *Technology*, **31**(5), 765-772.

709 Liu, C., Wyman, C.E. 2004. The effect of flow rate of very dilute sulfuric acid on xylan, lignin, and total  
710 mass removal from corn stover. *Industrial and Engineering Chemistry Research*, **43**(11), 2781-  
711 2788.

712 Lu, H., Lv, C., Zhang, M., Liu, S., Liu, J., Lian, F. 2017. Optimization of hydrothermal pretreatment for  
713 co-utilization C-5 and C-6 sugars of cassava alcohol residue. *Energy Conversion and*  
714 *Management*, **132**, 251-260.

715 Lloyd, T., Wyman, C.E. 2003. Application of a depolymerization model for predicting thermochemical  
716 hydrolysis of hemicellulose. *Applied Biochemistry and Biotechnology - Part A Enzyme*  
717 *Engineering and Biotechnology*, **108**(1-3), 53-68.

718 Makinen, K.K., Pape, H.R., Makinen, P.L., Bennett, C.A., Hujoel, P.P., Isokangas, P.J. 1995. Xylitol  
719 Chewing Gums and Caries Rates: A 40-month Cohort Study. *Journal of Dental Research*,  
720 **74**(12), 1904-1913.

721 Malkov, S., Tikka, P., Gullichsen, J. 2001. Towards complete impregnation of wood chips with aqueous  
722 solutions. *Paperi ja Puu/Paper and Timber*, **83**(6), 468-473.

723 Masutake, K., Maeda, H., Kawamata, T., Miyamae, M., Taguchi, K., Yonemitsu, Y., Yoshizaki, M.,  
724 Toyama, T. 1994. Food additive comprising water-soluble hemicellulose, Google Patents.

725 Mattila, P.T., Svanberg, M.J., Jämsä, T., Knuutila, M.L.E. 2002. Improved bone biomechanical  
726 properties in xylitol-fed aged rats. *Metabolism: Clinical and Experimental*, **51**(1), 92-96.

727 Morais, M.C., Pereira, H. 2012. Variation of extractives content in heartwood and sapwood of  
728 Eucalyptus globulus trees. *Wood Science and Technology*, **46**(4), 709-719.

729 Nakamura, A., Kato, M., Takahashi, T., Maeda, H. 2000. Process for producing emulsifiers, and  
730 emulsified compositions, Google Patents.

731 Nitsos, C.K., Choli-Papadopoulou, T., Matis, K.A., Triantafyllidis, K.S. 2016. Optimization of  
732 hydrothermal pretreatment of hardwood and softwood lignocellulosic residues for selective  
733 hemicellulose recovery and improved cellulose enzymatic hydrolysis. *ACS Sustainable*  
734 *Chemistry and Engineering*, **4**(9), 4529-4544.

735 Nurmi, J. 1992. Measurement and evaluation of wood fuel. *Biomass and Bioenergy*, **2**(1), 157-171.

736 Petersen, M.Ø., Larsen, J., Thomsen, M.H. 2009. Optimization of hydrothermal pretreatment of wheat  
737 straw for production of bioethanol at low water consumption without addition of chemicals.  
738 *Biomass and Bioenergy*, **33**(5), 834-840.

739 Predel, H. 2000. Petroleum Coke. in: *Ullmann's Encyclopedia of Industrial Chemistry*, pp. 1–21.

740 Qian, S., Wang, H., Zarei, E., Sheng, K. 2015. Effect of hydrothermal pretreatment on the properties of  
741 moso bamboo particles reinforced polyvinyl chloride composites. *Composites Part B:*  
742 *Engineering*, **82**, 23-29.

743 Reynolds, W., Baudron, V., Kirsch, C., Schmidt, L.M., Singer, H., Zenker, L., Zetzl, C., Smirnova, I.  
744 2016. Odor-Free Lignin from Lignocellulose by Means of High Pressure Unit Operations:  
745 Process Design, Assessment and Validation. *Chemie-Ingenieur-Technik*, **88**(10), 1513-1517.

746 Reynolds, W., Singer, H., Schug, S., Smirnova, I. 2015. Hydrothermal flow-through treatment of wheat-  
747 straw: Detailed characterization of fixed-bed properties and axial dispersion. *Chemical*  
748 *Engineering Journal*, **281**, 696-703.

749 Rissanen, J.V., Grénman, H., Willför, S., Murzin, D.Y., Salmi, T. 2014a. Spruce Hemicellulose for  
750 Chemicals Using Aqueous Extraction: Kinetics, Mass Transfer, and Modeling. *Industrial &*  
751 *Engineering Chemistry Research*, **53**(15), 6341-6350.

752 Rissanen, J.V., Grénman, H., Xu, C., Willför, S., Murzin, D.Y., Salmi, T. 2014b. Obtaining spruce  
753 hemicelluloses of desired molar mass by using pressurized hot water extraction. *ChemSusChem*,  
754 **7**(10), 2947-2953.

755 Rogalinski, T., Ingram, T., Brunner, G. 2008. Hydrolysis of lignocellulosic biomass in water under  
756 elevated temperatures and pressures. *The Journal of Supercritical Fluids*, **47**(1), 54-63.

757 Sánchez-Bastardo, N., Romero, A., Alonso, E. 2017. Extraction of arabinoxylans from wheat bran using  
758 hydrothermal processes assisted by heterogeneous catalysts. *Carbohydrate Polymers*, **160**, 143-  
759 152.

- 760 Sluiter, A., Hames, B., Hyman, D., Payne, C., Ruiz, R., Scarlata, C., Sluiter, J., Templeton, D., Wolfe, J.  
761 2008a. Determination of Total Solids in Biomass and Total Dissolved Solids in Liquid Process  
762 Samples. in: *Laboratory Analytical Procedure (LAP)*.
- 763 Sluiter, A., Hames, B., Ruiz, R., Scarlata, C., Sluiter, J., Templeton, D. Determination of Sugars,  
764 Byproducts, and Degradation Products in Liquid Fraction Process Samples. Laboratory  
765 Analytical Procedure (LAP). *Laboratory Analytical Procedure (LAP)*.
- 766 Sluiter, A., Hames, B., Ruiz, R., Scarlata, C., Sluiter, J., Templeton, D., Crocker, D. 2008b.  
767 Determination of structural carbohydrates and lignin in biomass. in: *Laboratory Analytical  
768 Procedure (LAP)*.
- 769 Sluiter, A., Ruiz, R., Scarlata, C., Sluiter, J., Templeton, D. 2008c. Determination of Extractives in  
770 Biomass in: *Laboratory Analytical Procedure (LAP)*.
- 771 Tanaka, M., Takamizu, A., Hoshino, M., Sasaki, M., Goto, M. 2012. Extraction of dietary fiber from  
772 Citrus junos peel with subcritical water. *Food and Bioproducts Processing*, **90**(2), 180-186.
- 773 Thomsen, M.H., Thygesen, A., Thomsen, A.B. 2008. Hydrothermal treatment of wheat straw at pilot  
774 plant scale using a three-step reactor system aiming at high hemicellulose recovery, high  
775 cellulose digestibility and low lignin hydrolysis. *Bioresource Technology*, **99**(10), 4221-4228.
- 776 Ur-Rehman, S., Mushtaq, Z., Zahoor, T., Jamil, A., Murtaza, M.A. 2015. Xylitol: A Review on  
777 Bioproduction, Application, Health Benefits, and Related Safety Issues. *Critical Reviews in  
778 Food Science and Nutrition*, **55**(11), 1514-1528.
- 779 Yedro, F.M., Cantero, D.A., Pascual, M., García-Serna, J., Cocero, M.J. 2015a. Hydrothermal  
780 fractionation of woody biomass: Lignin effect on sugars recovery. *Bioresource Technology*, **191**,  
781 124-132.
- 782 Yedro, F.M., García-Serna, J., Cantero, D.A., Sobrón, F., Cocero, M.J. 2015b. Hydrothermal  
783 fractionation of grape seeds in subcritical water to produce oil extract, sugars and lignin.  
784 *Catalysis Today*, **257, Part 2**, 160-168.
- 785 Yedro, F.M., Grénman, H., Rissanen, J.V., Salmi, T., García-Serna, J., Cocero, M.J. 2017. Chemical  
786 composition and extraction kinetics of Holm oak (*Quercus ilex*) hemicelluloses using subcritical  
787 water. *Journal of Supercritical Fluids*.
- 788 Yedro, F.M., Grénman, H., Rissanen, J.V., Salmi, T., García-Serna, J., Cocero, M.J. Chemical  
789 composition and extraction kinetics of Holm oak (*Quercus ilex*) hemicelluloses using subcritical  
790 water. *The Journal of Supercritical Fluids*.
- 791 Yoon, J., Lee, H.W., Sim, S., Myint, A.A., Park, H.J., Lee, Y.-W. 2016. Hydrolysis kinetics of tulip tree  
792 xylan in hot compressed water. *Bioresource Technology*, **214**, 679-685.
- 793 Zetzl, C., Gairola, K., Kirsch, C., Perez-Cantu, L., Smirnova, I. 2012. One-reactor design for the  
794 fractionation of lignocellulosic biomass under high pressure. *Chemie-Ingenieur-Technik*, **84**(1-  
795 2), 27-35.
- 796 Zhang, J., Xiao, H., Li, N., Ping, Q., Zhang, Y. 2015. Synthesis and characterization of super-absorbent  
797 hydrogels based on hemicellulose. *Journal of Applied Polymer Science*, **132**(34), n/a-n/a.

798

799 **List of Tables**

800 **Table 1.** Scale up criteria from lab-scale to pilot-scale reactor.

801

802 **List of Figures**

803 **Figure 1.** Schematic flow diagram of the lab-scale experimental system. Equipment: D-01 water  
804 deposit, P-01 pump, E-01 concentric tube heat exchanger, H-01 electric heater, R-01 flow-through  
805 reactor, BPV-01 Go-back pressure valve.

806 **Figure 2.** a) Section of reactor composition: 1. Cap of the outer SS cylinder; A,B,C. Openings; 2.  
807 External SS cylinder; 3. Ball valve; 4. Superior internal cylinder; 5. Inferior internal cylinder; 6. Bottom  
808 with orifices of the inferior cylinder, 7. Metallic mesh. b) Schematic flow diagram of the pilot-scale  
809 experimental system. Equipment: D-01 water deposit, P-01 centrifugal pump, E-01, E-02, E-03  
810 concentric tube heat exchangers, H-01 electric heater, R-01 flow-through reactor, 3V-01 three way  
811 valve, V-01, V-02 ball valves; BPV-01 Go-backpressure valve, C-01 plastic container.

812 **Figure 3.** a) Yield of xylose extracted at 160 and 170 °C with a lab-scale and a pilot-scale reactor; b)  
813 Percentage error between yields of xylose obtained with lab-scale and pilot-scale reactor at 160 °C and  
814 170 °C. c) Cumulative yields of xylose monomers extracted with a lab-scale and a pilot-scale reactor at  
815 160 and 170 °C.

816 **Figure 4.** Schematic flow diagram of the multistage pilot flow-through reactor where three units are  
817 operating in series. Equipment: D-X water deposits, P-X pumps, E-X concentric tube heat exchanger,  
818 ES-X concentric tube heat exchangers for sample withdrawn, H-X electric heaters, R-X flow-through  
819 reactors, 3V-X three way valves, BPV-X Go-back pressure valves, V-X ball valves, NV-X needle  
820 valves, C-X plastic containers. Liquid flow entering in reactors R-01, R-02 and R-03 connected in series  
821 and by-passing reactors R-04 and R-05.

822 **Figure 5.** a) Cumulative yield of total xylose extracted, obtained from units 1, 2 and 3 during 90 min of  
823 operation. b) Molecular weight of hemicelluloses extracted obtained from units 1, 2 and 3 of the system  
824 during 90 min of operation. c) Instantaneous values of acetic acid yield obtained from unit 3 of the  
825 system during 90 min of operation. d) Ratio between monomeric xylose obtained after the hydrothermal

826 treatment and total xylose extracted after 90 min of operation, as a function of the residence time of the  
827 extracted solutions within the system.

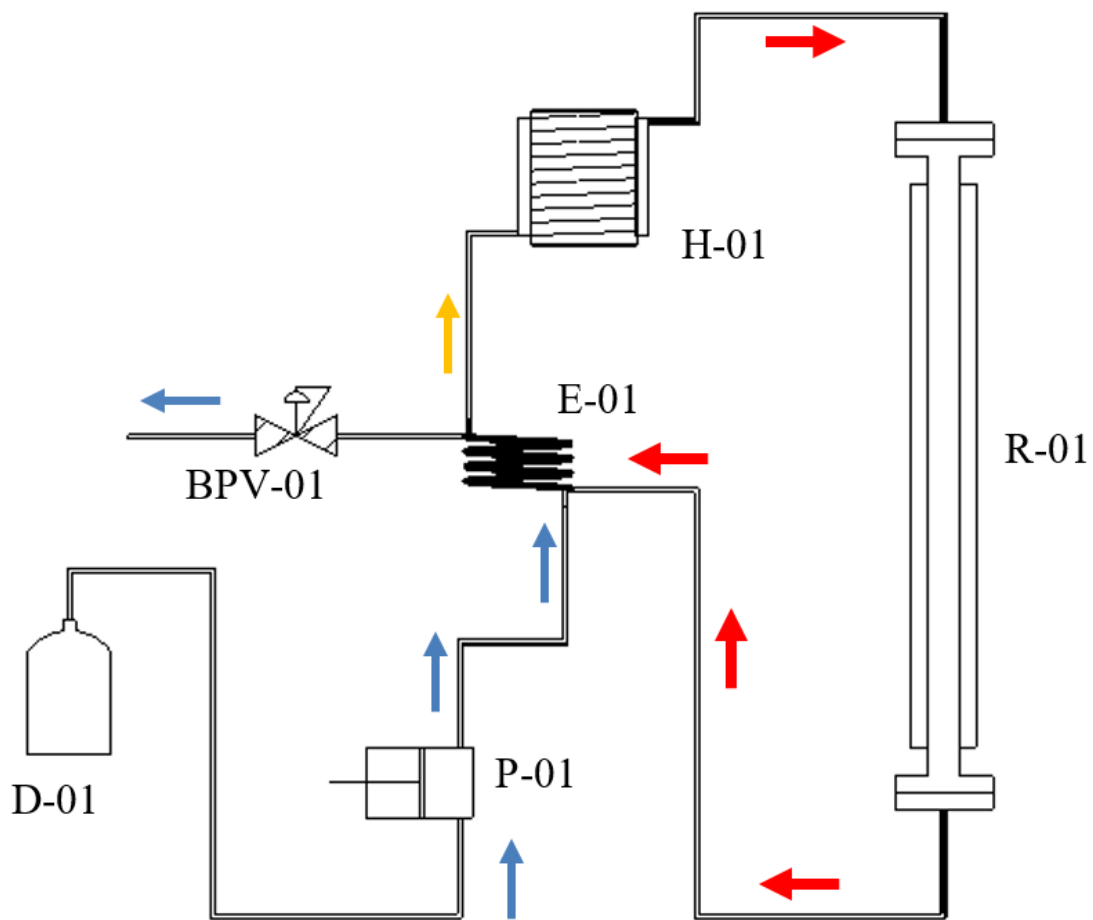
828 **Figure 6.** Mass balance calculated at the end of extractions at 140, 150, 150, 160 and 170 °C.

829 **Table 1.**

830

<b>Parameter</b>	<b>Unit</b>	<b>LAB SCALE</b>	<b>PILOT SCALE</b>
Flow rate	mL/min	3.5	250
Internal diameter	cm	0.96	4
Length	cm	38	159
Volume	cm <sup>3</sup>	27.5	2000
L/ID		40	40
Liquid Residence time	min	6.0	6.0
Porosity		0.71	0.71

831 **Figure 1.**



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833 **Figure 2.**

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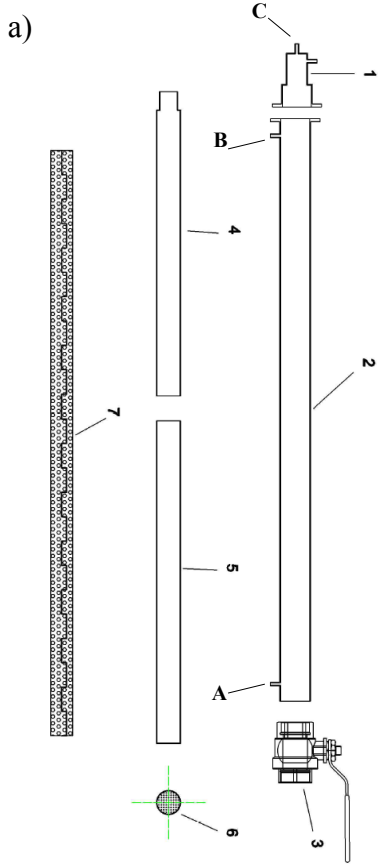
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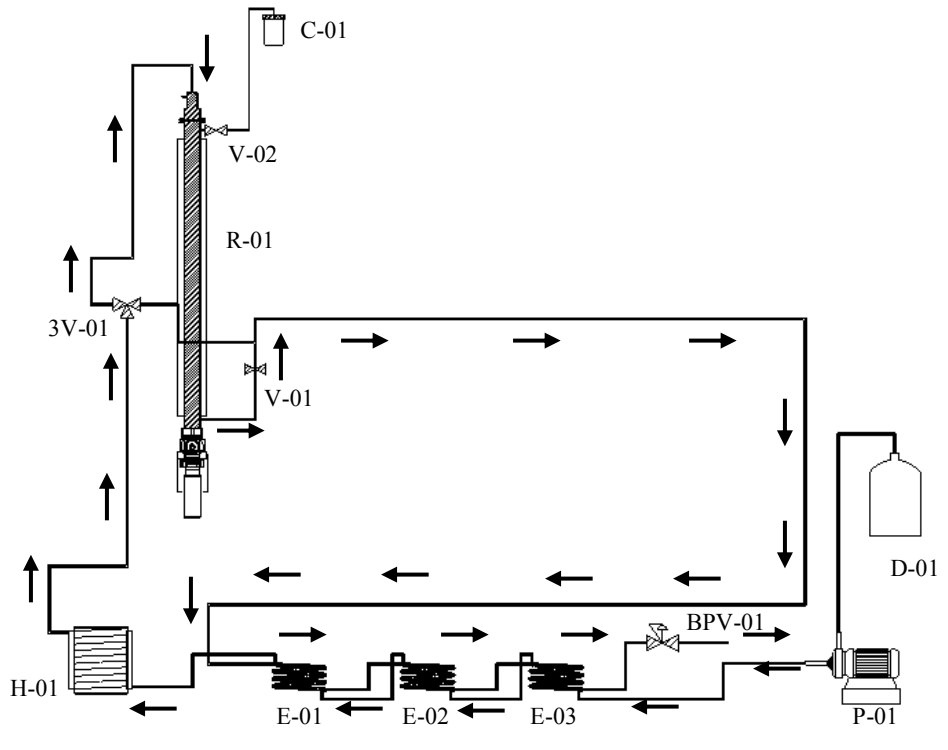
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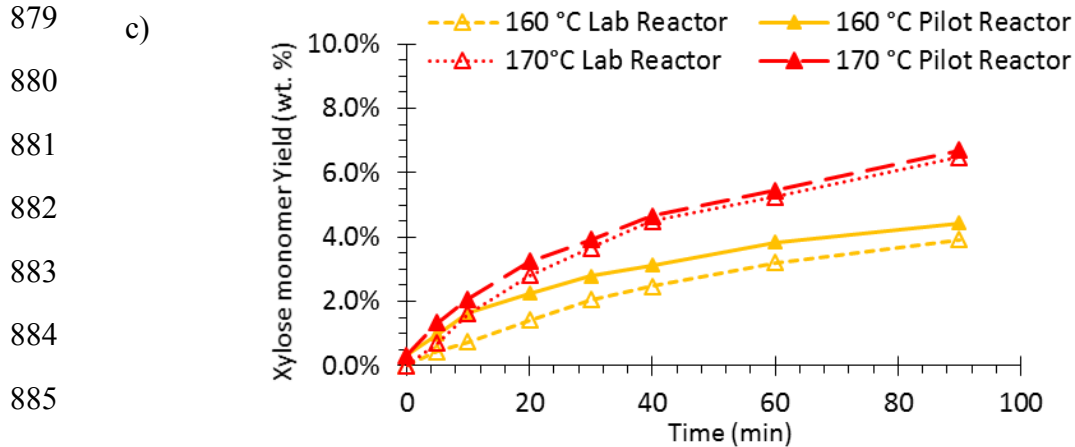
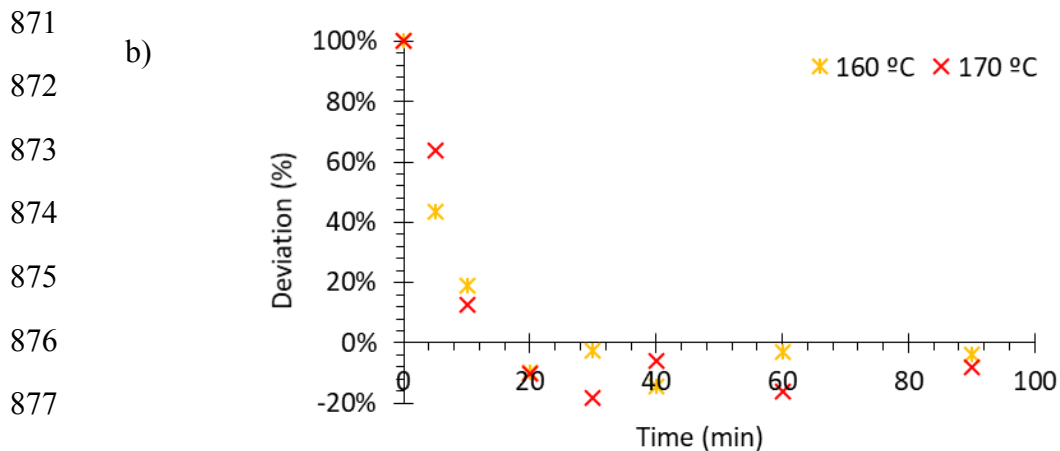
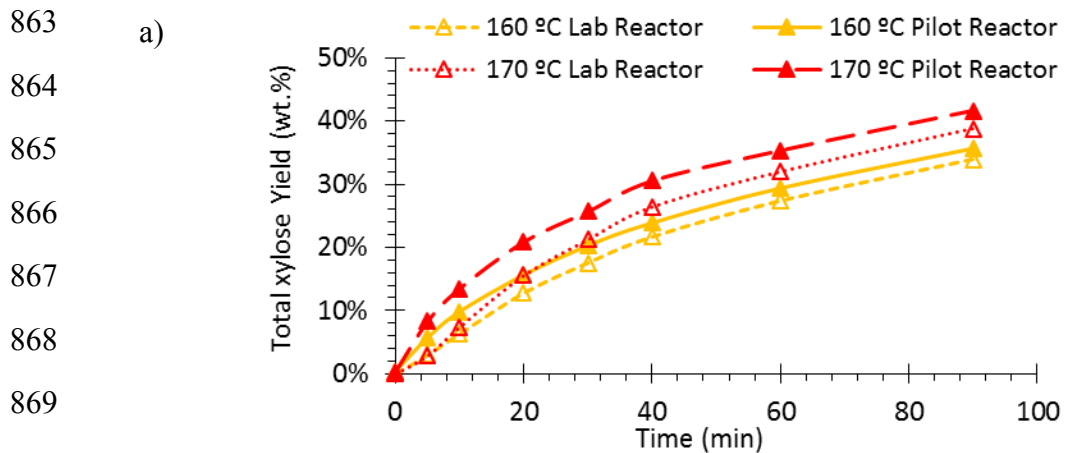
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862 **Figure 3**



890 **Figure 4**

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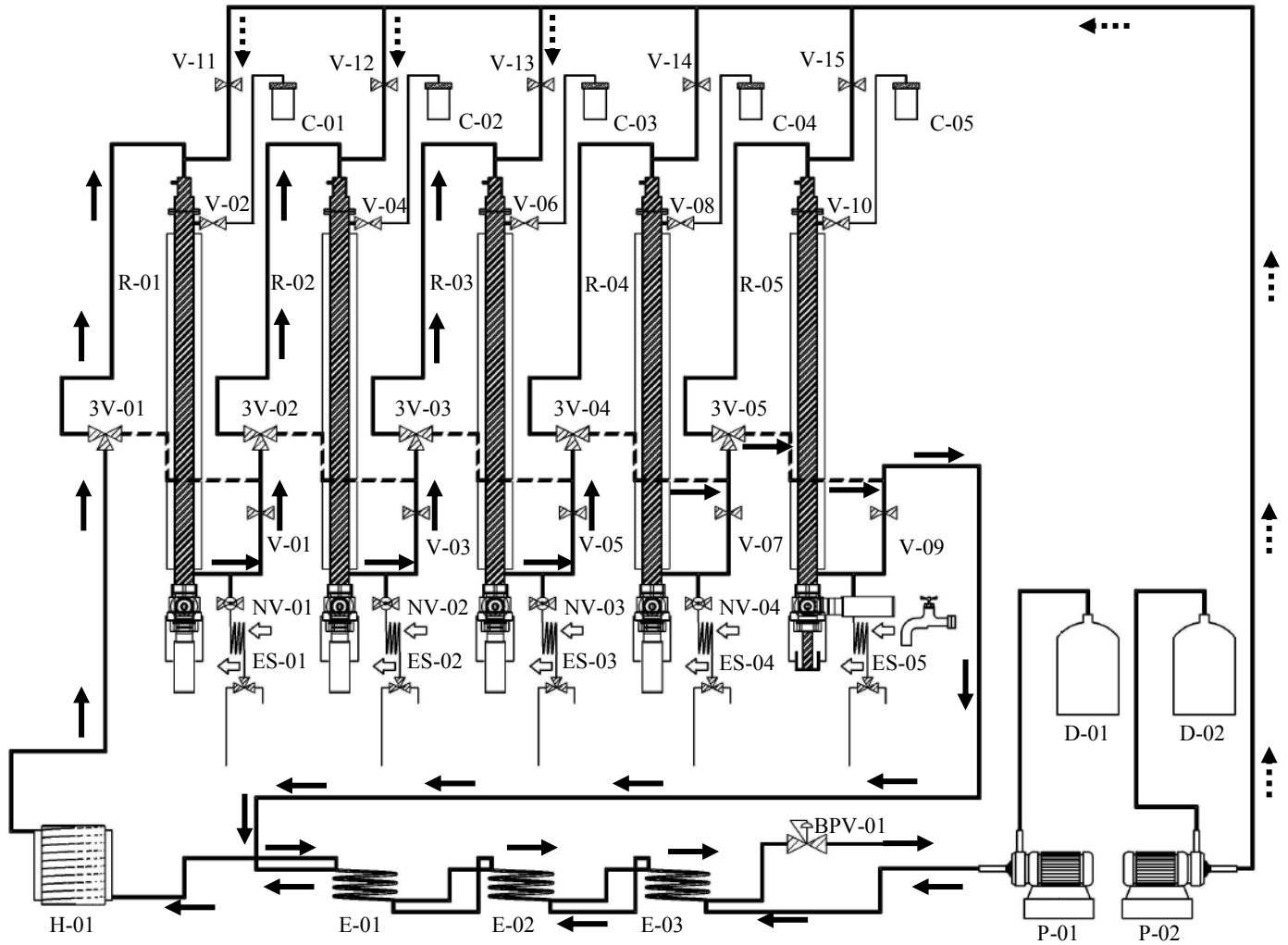
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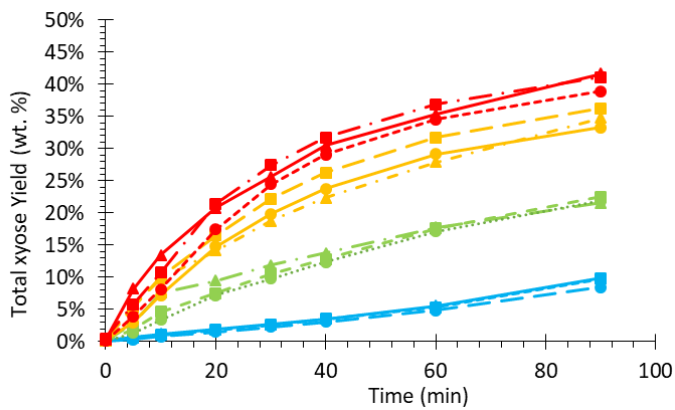
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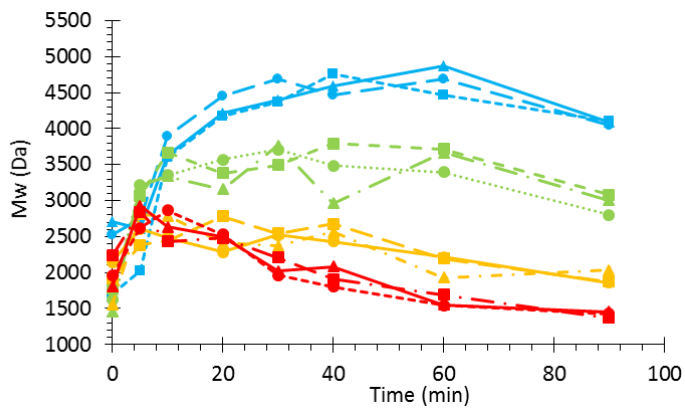


914 **Figure 5**

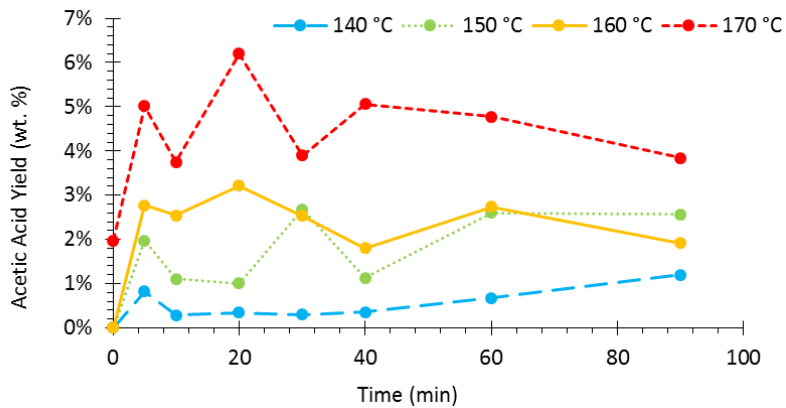
915 a)



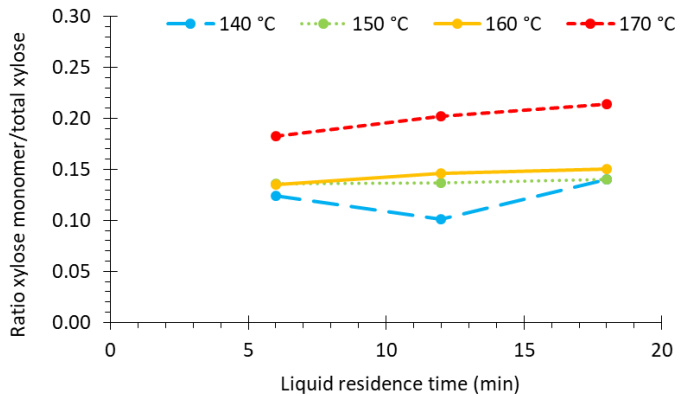
922 b)



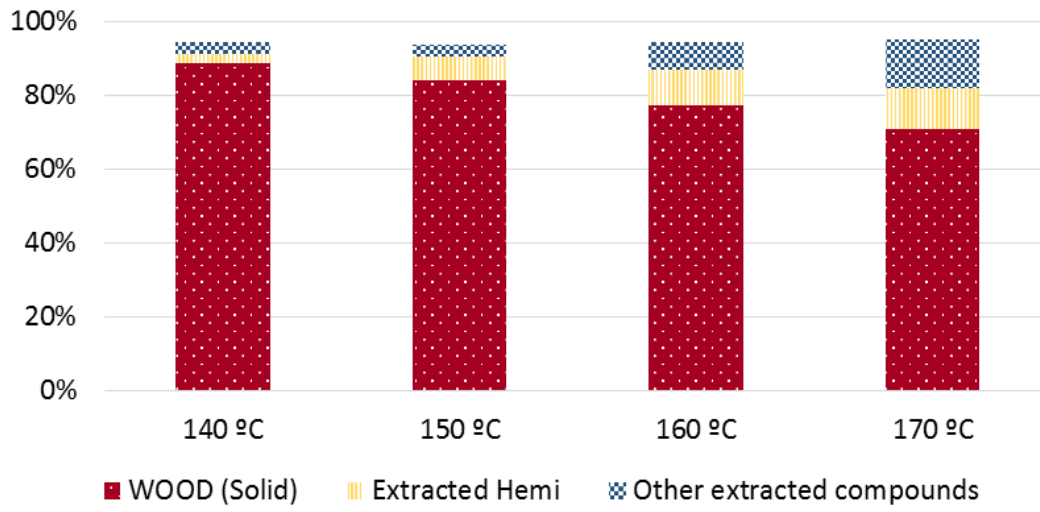
928 c)



935 d)



942 **Figure 6.**



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**Supplementary info**

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