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2 **extracts by Co-precipitation in Supercritical**
3 **Antisolvent (SAS) technology.**

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25 **Formulation of açai (E. oleracea Mart.) pulp and seeds**
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35
36 **Abstract.** *Açai (Euterpe oleracea Mart.) is a black-purple berry, typically found in*
37 *Amazon Rainforest, and a natural phytochemical source, which shows a high content of*
38 *polyphenols and flavonoids, with remarkable properties as an antioxidant, anti-*
39 *inflammatory, antimicrobial, and natural dye. The precipitation and encapsulation of*
40 *Açai extracts with biopolymers by Supercritical Anti-Solvent (SAS) process were*
41 *investigated and proposed as primary formulation to protect the active compounds from*
42 *early degradation and its application in pharmaceutical, cosmetic, and alimentary*
43 *products. The extractives were obtained from pulp and seeds by Pressurized Microwave*
44 *assisted Energy (PMAE)(300W and 1.5bar) using acidified ethanol/water (1:1 v/v) as a*
45 *solvent. The extracts were characterized in terms of total polyphenols content (TPC). The*
46 *SAS process was carried out by semi-continuous batch at constant conditions (40°C,*
47 *100bar). Particle morphology was studied by SEM, TGA and FTIR. The best results were*
48 *obtained when ethanol and PVP were applied. The process also allow the particle*
49 *micronization and TPC value increment.*

50 **Keywords:** *natural product formulation, Antioxidant, Microencapsulation, Supercritical*
51 *CO₂.*

52 1. Introduction

53 Açaí (*Euterpe Oleracea Mart.*) is a black-purple berry typically found in
54 Amazon Rainforest. The fruit and seeds have remarkable properties as an antioxidant due
55 to its high content of polyphenols, flavonoids, and, anthocyanin. Products such as
56 flavonoids and anthocyanin are studied because of their pharmacological and natural dye
57 potential. The seeds are the main by-product obtained during the industrial processes to
58 extract the fruit pulp, and pulp that is considered inappropriate for human consumption
59 also becomes an industrial residue.

60 The benefits provided by polyphenols, flavonoids, and anthocyanin convert this
61 class of compounds in an interesting study area. These studies usually focus on the
62 relation of antioxidant effect and its bioavailability [1]. As occurs with many other natural
63 compounds, the application of these biochemical compounds may be limited by their low
64 solubility in aqueous systems. Encapsulating the antioxidant extract in a polymeric matrix
65 for its protection is an excellent method to protect it for an earlier degradation, as well to
66 improve its solubility and, as a consequence, its bio-disponibility [2].

67 The SAS (Supercritical Anti Solvent) encapsulation is a very versatile process
68 for processing various natural and pharmaceutical compounds, with reasonable control of
69 particle properties, and without product degradation or contamination [3]. In this process,
70 the particle is solubilized in an organic solvent by which it has an affinity. Then, with the
71 aid of a pump, this solution is introduced into a precipitator containing high-pressure CO₂.
72 By introducing the solution into the precipitator as a spray, the dissolving droplets mix
73 with CO₂, which acts as an anti-solvent, as it decreases the solubility of the solute relative
74 to the solvent. In this way, the precipitation of the compound occurs due to the high
75 supersaturation achieved [4]. Compared to precipitation techniques using liquid

76 antisolvents, the SAS technique has the advantage that, due to the favorable properties in
77 the supercritical medium, very high and homogeneous supersaturation can be achieved
78 very quickly, which favours control over fluid properties. In addition, moderate
79 temperatures below 80°C can be applied, so the process becomes suitable for working
80 with sensitive substances such as products of natural origin [3]. The precipitation and
81 encapsulation technical study apply to sensitive compounds. It has been shown, in a
82 previous study, that polyvinylpyrrolidone (PVP) and Pluronic® F127 are viable
83 encapsulating surfactant option when natural products were formulated [5].

84 Spray-drying and vacuum-drying, among other techniques, are described in the
85 literature as advantageous formulation process for natural compounds. Many of those
86 techniques require to elevate temperature, as in the spray-drying process where it can
87 reach 125°C, corroborating to the degradation of active molecules, even utilizing low
88 pressure [5]. Vacuum-drying (considered beneficial to natural product formulation by
89 allows water as a solvent) promotes non-spherical particle formation and requires
90 temperature which may also result in degradation of interest compound [6]. The
91 temperature required in vacuum-drying is lower than in spray-drying, however, it is still
92 higher than in other types of formulation, such as in supercritical anti-solvent (SAS).

93 Concerning the encapsulation material, different cyclodextrins have been
94 purposed by many authors. The cyclodextrin inclusion complex [7], provides the specific
95 water solubility and antioxidant activity improvement [8,9]. The amorphous structured
96 material, resulting from the encapsulation of antioxidants compounds in mixture of
97 Eudragit® E and polyvinyl alcohol, supported by hydrogen bonds has shown a favourable
98 improvement of bioavailability [10], similar to lipid-particles charged with antioxidant
99 compounds by an emulsion freezing process [11].

100 The present study analyses the use of Supercritical Anti-solvent (SAS) as
101 formulation process for the co-precipitation of açai (*E. oleracea* Mart.) pulp and seeds
102 extracts in biopolymers (Pluronic F-127 and PVP). The process aims to promote an
103 formulation of antioxidant obtained from both açai extracts, improving the water
104 solubility and reducing the early degradation. The effect of the main process parameters
105 (polymer:extract ratio, initial concentration) was studied and products were characterized
106 considering its morphology (size and shape) and composition (total polyphenol content,
107 total anthocyanin content and antioxidant activity).

108

109 **2. Material and methods**

110

111 *2.1 Extract preparation*

112 Two fractions of açai by-products (seeds and pulp), obtained from Obidos-PA-
113 Brazil, has been prepared to the extraction process following different protocols. The
114 seeds fraction was milled and dried, and the pulp fraction was lyophilized. The subsequent
115 preparation was to remove the oil content present in both materials [12].

116 Extracts were obtained by Pressurized Microwave-Assisted Extraction (PMAE)
117 (300W and 1.5bar). Ethanol/water (1:1 v/v) was used as a solvent for the extraction, and
118 citric acid as pH regulator (pH 3). Individually, each raw-material was placed in a
119 pressurizable flask, mixed and homogenized with the prepared solvent. The flask
120 extractor was then introduced into a Circular Energy-Microwave (CEM). CEM was set
121 to constant power (300W), varying the temperature and pressure till 1.5 bar and sample
122 was suddenly cooled introducing the flask into a cold bath. The flask content was
123 transferred into centrifuge-tube. The tube was centrifugated at 5000 rpm for 10 min, and

124 the supernatant recovered and dried to be used in the preparation of the feed solution, as
125 described in section 2.2.

126 *2.2 Preparation of the feed solution*

127 The encapsulation process starts with the preparation of the solution to be
128 encapsulated. Selected extract, polymer, and solvent are mixed at constant stirring. The
129 effect of polymer:extract ratio was studied varying the proportions among both materials
130 ((1:1 m/m), (2:1 m/m), (1:2 m/m), (1:4 m/m)). The effect of the concentration on the
131 formation of the particles was also studied, changing the ratio of total solid (mass) per
132 volume of solvent (0.50 g/mL, 0.33 g/mL, 0.50 g/mL). Furthermore, SAS experiments
133 with the pure polymer and the extract were also performed for comparison purposes.

134

135 *2.3 SAS Preparation*

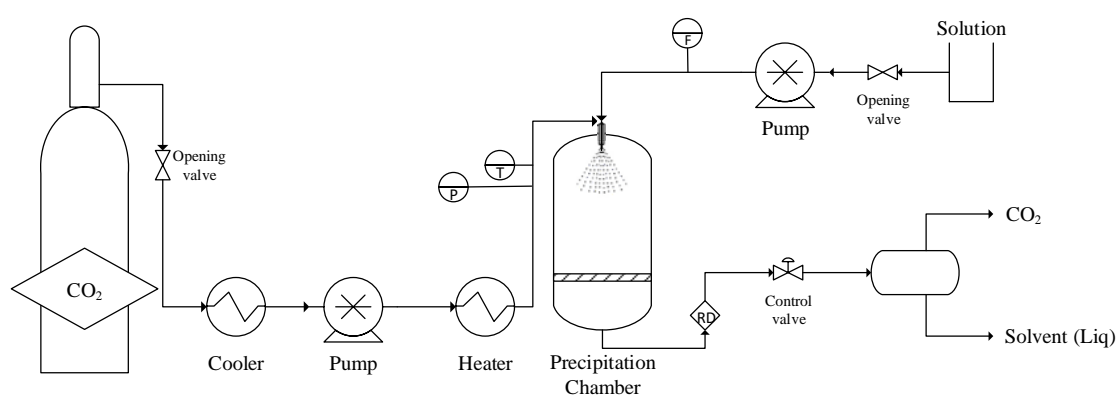
136 The general scheme of the equipment used for SAS experiments can be observed
137 in Figure 1. The precipitator consists of a metallic jacketed cylinder that is continuously
138 heated by the refluxing water of a thermostatic bath at 40 °C. It has two flanges, the upper
139 one with three main holes through which the barometer and the safety valve are
140 connected, in the second one, the entrance of the CO₂ and the solution, and in the third
141 one, the flow, temperature, and pressure meters. In the flow flange, the outlet of CO₂ and
142 solvent is placed. In the CO₂ line, after the opening valve, CO₂ is liquefied and pumped
143 by the piston pump with a flow rate 2kg/h. Before the chamber CO₂ is again heated to
144 40°C and introduced into the precipitator while measuring temperature and flow. The
145 second inlet line drives the solution for encapsulation, from the baker to the precipitator,
146 helped by a chromatography pump (model 305 Gilson) and the flow is controlled at 2
147 mL/min. In the precipitator, the inlet-lines have a concentric entrance, which produces a
148 spray effect when the solution and CO₂ go inside simultaneously. This leads to the

149 solubilisation of the solvent and the precipitation of particles from solution onto the filter
150 placed at the bottom of the recipient. The process is carried out by semi-continuous batch,
151 constant temperature (40°C), and pressure (100 bar). A rupture disc set at 210 bars, an
152 outlet GO-type valve and a flask that separates the CO₂ from the residual solvent are
153 placed at the exit of the precipitator [13].

154

155

(Figure1)



156

157 Figure 1- Complete SAS flow diagram. (P- pressure controlling; T- temperature
158 controlling; F-flow controlling; and RD-Rupture disc)

159

160 2.4 Product characterization

161 2.4.1 Total polyphenol content (TPC)

162 For TPC analysis a capped test tube was used, adding 40µL of the extract, 3 mL
163 of ultrapure water and 200 µL of ciocalteau reagent. It is also necessary to prepare a
164 control sample using 40µL of extraction solvent, 3 mL of ultrapure water and 200 µL of
165 folin-ciocalteau reagent. Tubes were closed and homogenized at 40°C for 5 minutes.
166 After this period 600µL of Na₂CO₃ (20% v/v) solution was added, tubes were vigorously
167 stirred, and kept in hot-water-bath at 40°C for 30 minutes. Finally, the samples were

168 analysed by spectrophotometer ($\lambda = 765$ nm). Tests were performed in duplicate and the
169 average of TPC concentration is given in Gallic acid equivalent per 100 g of dry material.

170 [14]

171

172 2.4.2 Total anthocyanin content (TAC)

173 The TAC analyses were performed with the aid of a spectrophotometry. Samples
174 were diluted (1:4) in a potassium hydroxide buffer (0.025M KCl) at pH 1.0 and buffered
175 with acetate trihydrate buffer ($\text{CH}_3\text{CO}_2\text{Na} \cdot 3\text{H}_2\text{O}$ 0.4 M) at pH 4.5. Samples were diluted
176 in both buffers solution, at pH 1.0 and pH 4.5, and each dilution was measured at 520 nm
177 and 700 nm. Tests were performed in duplicate and the average of concentration of
178 anthocyanins in each sample is given in g of cyanidin equivalent per 100 g of dry material.

179 [15]

180

181 2.4.3 Sample morphology

182 The study of particle morphology was carried by Scanning Electron Microscopy
183 (SEM) analysis (FLEX SEM 1000 Hitachi), and Fourier transform infrared spectroscopy
184 (FTIR) analysis (Bruker ALPHA FT-IR spectrometry) with a single sampling module of
185 platinum ATR diffraction.

186

187

188 3. Results and Discussions

189 Table 1 reports the experimental conditions applied in all the experiments
190 performed, as well as the total polyphenol content of the particles produced. The higher
191 TPCs were observed when ethanol was applied as solvent and PVP as a carrier. The assays
192 performed using acetone as solvent produced a sample with a plasticized aspect; in

193 addition, precipitation was observed in the initial solution. It was also determined that
194 applying moderate heat (30°C) helped in the homogenization of encapsulation solution.

195 Different seeds-extract to PVP ratios of 2:1, 1:1, 1:2, and 1:4 were processed.
196 Pulp extract and PVP were successfully processed on ratios of 2:1, and 1:1. The
197 encapsulated material resulting from the processes utilizing the ratio of 2:1, 1:1, 1:2, had
198 shown powder aspect. However, when a ratio of 1:4 was applied the material obtained
199 from the processing had irregular and pelletized aspect. Exploratory experiments
200 performed with ratios above 1:4 resulted in plasticized material (results not shown). All
201 experiments performed with Pluronic F127 were unsuccessful.

202

203 *3.1 Composition analysis*

204 Composition analysis revealed that the material obtained from the processing of
205 seeds-extracts had the expected total polyphenol content (TPC) corresponding to the
206 encapsulated ratio; however, those materials wherein pulp-extract was applied did not
207 present significant phenolic content besides the presence of typical color produced by
208 anthocyanin. In Table 1, the results obtained from the analyses of the total phenolic
209 content (TPC) is expressed in Gallic acid equivalent (GAE) per gram of precipitate. The
210 original extract was processed with no polymer addition and taken as a reference to
211 compare particle ratio formation. The original extract processed by SAS was enriched in
212 terms of polyphenol content compared to the original sample that was not processed by
213 SAS, due to the selectivity of CO₂. During the precipitation, not all the components in the
214 initial extract were precipitated, as there were still remaining components in the extract
215 that showed affinity to CO₂ (such as residual oil). However, CO₂ has low affinity to
216 polyphenols that are precipitated. So, the obtained products are particles enriched in

217 polyphenols in relation to the initial extracts. The influence of the initial concentration of
 218 the solution was also studied using a constant extract/polymer ratio (1:1) and varying the
 219 extract/solvent proportions (2.50 g/L, 3.33 g/L and 5.00g/L). The results were based on
 220 TPCs values. The highest TPC value was observed in those particles obtained by the
 221 proportion of 3.33 g/L, 211 mg GAE/g particle, while for the other proportions studied
 222 (2.50 and 5.00 g/L) the TPC values were 80 and 78 mg GAE/g particle, respectively. In
 223 addition, the SAS processing increased the TPC value of processed extract (500 mg
 224 GAE/g particle) concerning the original extract (261 mg GAE/g particle), the TPC value
 225 found was almost twice higher than initial TPC showing the capacity of SAS acts as
 226 purification process for natural extracts.

227 Similar to that observed with the açai seeds extract, the pulp product also
 228 presented better results when intermediate concentrations were applied. Although it was
 229 not possible to recover or perform the relevant probes at the single processed pulp extract,
 230 it was possible to compare the results with the original pulp and to observe that the
 231 processed material has lower TPC than expected. Tests were also performed to evaluate
 232 the total content of total anthocyanins by differential buffer pH in spectrophotometry, and
 233 none of the tested materials showed detectable anthocyanin content although all have
 234 light-pink coloration, typically induced by the presence of this substance in trace amounts.

235

236 Table 1 - Experimental conditions and total polyphenol content of SAS-
 237 processed particles.

238

(Table 1)

Extract (Material)	Polymer	Solvent	Polymer: extract Ratio	Initial solution concentration (g/L)	TPC content mg GAE/g particle	SD
Seeds (unprocessed)	-	-	-	-	261	3
Seeds	-	Ethanol	-	3.33	500	5
Seeds	PVP	Ethanol	1:1	5.00	78	3

Seeds	PVP	Ethanol	1:1	3.33	211	8
Seeds	PVP	Ethanol	1:1	2.50	80	4
Seeds	Pluronic	Acetone	1:1	3.33	132	8
Seeds	PVP	Ethanol	2:1	5.00	142	6
Seeds	PVP	Ethanol	2:1	3.33	187	8
Seeds	PVP	Ethanol	1:2	5.00	54	3
Seeds	PVP	Ethanol	1:2	3.33	72	3
Pulp (unprocessed)	-	-	-	-	108	3
Pulp	-	Ethanol	-	-	-	-
Pulp	PVP	Ethanol	2:1	3.33	7	>1
Pulp	PVP	Ethanol	1:1	5.00	27	1
Pulp	PVP	Ethanol	1:1	3.33	31	2
Pulp	PVP	Ethanol	1:1	2.50	35	>1

239

240

241

242 3.2 Morphological study

243 The morphology was studied by Scanning electron microscope (SEM) images.

244 The SAS process has promoted the particle seed-extract micronization. SEM images were

245 used to analyse the differences in the morphology of the particles obtained from the

246 single-processed seed-extract with respect to the original material. Moreover, when PVP

247 was applied as carrier in the encapsulation process, it was possible to produce even

248 smaller particles (Table 2). Pluronic F-127 did not show good properties as a carrier for

249 seed-extract, because all experiments resulted in a plasticized material.

250

251

252 Table 2- Comparative of SEM images of encapsulated material, PVP, seed-extract, and,

253 seed-extracts SAS processed.

254

(Table 2)

255

256 As observed from TPC analysis, the solvent to solute proportion was considered
257 an important variable once the mass/solvent ratio affected the co-precipitation. The
258 particles presented in Figure 2 have appropriate morphologies, with particles well covered
259 with the polymer. These particles were also the ones with the highest TPC value, 211 mg
260 GAE/g particle.

261

262 (Figure2)

263 Figure 2 – Seed-extract co-precipitated with PVP at (1:1) ratio, 3.33g/L.

264

265 However, the excessive reduction of the mass/solvent ratio caused the pump to
266 malfunction, causing blockages, which impairs the constant flow pumping of suspended
267 material. The resulting malformed content produced by these problems can be observed
268 in figure 3.

269

270

271 (Figure 3)

272 Figure 3 – Seed-extract co-precipitated with PVP at (1:1) ratio, 5.00 g /L.

273

274 The best concentration found was 2.5g/L, at this concentration, a more efficient
275 co-precipitation, particles with better distribution, less exposed areas and small particles
276 sizes were obtained. The solvation ratio changed the final particle size.

277

278 (Figure 4)

279 Figure 4 – Seed-extract co-precipitated with PVP at (1:1) ratio, 2.50 g /L.

280

281 When pulp-extract was single processed, without polymer, no particles were
282 obtained, which was attributed to a strong micronization, where the particle could not be
283 retained by the filter (0,45 μm). SEM images (Figure 5) of the product, showed small
284 extract fragments of extract inserted in the polymer. Açai pulp is a very oily material; 43
285 % of its dry-weight is oil content. The pulp was treated to remove the oil before the
286 extraction, but small remaining amounts of oil in the Açai pulp extract can justify the
287 differences observed between both extracts.

288

289 (Figure 5)

290 Figure 5 - Pulp-extracted and PVP processed at (1:1) ratio, (a) 5.00g/L, and (b)3.33g/L.

291

292

293 *3.3 Thermostability*

294 The thermostability study was made by the thermogravimetric analysis (TGA).
295 The analysis measured the loss of mass in percentage through the temperature increment.
296 The first thermo-decay, called onset-temperature (T_{onset}), can be taken as a qualitative
297 measurement of thermostability. In this way, to increase the T_{onset} is a relevant
298 advantage of the formulation process. Seed-extract single processed (Figure 6), with no
299 carrier addition, showed a very small loss in T_{onset} concerning the original seed extract,
300 the difference observed was around 2°C , the process achieved to increase the extract
301 water-solubility. In the ratio of 1:1, the TGA obtained was similar to pure PVP, presenting
302 a small variation in T_{onset} . The material thermostability increased when more polymer
303 was added to the process, as the T_{onset} was higher the following gap formed between
304 baseline and tangent was less evident, being almost imperceptible for the ratio of (1: 2).

305

306

(Figure 6)

307

Figure 6- Thermogravimetric (TG) curve obtained for seed-extract co-

308

precipitated indifferent proportions with PVP.

309

310

The Derivative Thermogravimetry (DTG) is the first derivative curve obtained

311

from TGA analysis and simulation of Differential Scan Calorimetry (DSC). This

312

analytical technique is able to detect the precise inflection point of the TG curve, marking

313

the point of mass changes due to vaporization or degradation. The results provided in

314

Figure 7 showed the existence of a first peak relative to the loss of humidity, followed by

315

two degradation peaks. It is possible to see (figure 7) that all peaks are narrow and simple,

316

denoting that each loss process happened in a unique step. In addition, second peaks are

317

displaced among them denoting the soft variation in the thermostability of obtained

318

material, as previously commented.

319

320

(Figure 7)

321

Figure 7 – Derivative Thermogravimetry (DTG) curve of co-precipitated seed-extract

322

and PVP material.

323

324

Meanwhile, Pulp-extract showed a different behaviour when it was processed by

325

the SAS technique. The materials obtained did not present modifications on TGA or peaks

326

displacement in DTG denoting the encapsulation was not performed successfully.

327

328

3.4 FTIR

329 The FTIR analysis of the formulated material can be compared to the source
330 material. In both cases, the formulated material showed characteristic peaks of the source
331 material seeds-extract (Figure 8), and pulp-extract (Figure 9).

332

333 (Figure 8)

334 Figure 8 – FTIR of PVP, original seeds-extract, and encapsulated material:

335 characteristic peaks of (I)seeds-extract and (II)PVP.

336

337 (Figure 9)

338 Figure 9 – FTIR of PVP, original pulp-extract, and encapsulated material: characteristic

339 peaks of (I)pulp-extract and (II)PVP.

340

341 **4. Conclusion**

342 In this work the encapsulation of active extract obtained by pressure-assisted
343 microwave extraction using as a source of extract the seeds and pulp residues (not suitable
344 for direct human consumption) of the açai fruit (*Euterpe oleracea* Mart) was studied. The
345 encapsulations were tested with different polymers (PVP and Pluronic F127) and solvents
346 (ethanol and acetone). Tests were also performed with variations of the extract/polymer
347 and solid/liquid ratio present in the starting solution. The best results were obtained when
348 ethanol was applied as the solvent and PVP as the encapsulating polymer, as these
349 conditions yielded higher TPC contents and more regular particle morphologies.

350 Among the proven extracts, better results were observed for the material
351 extracted from the açai seeds, which is attributed to the higher oil content of pulp extracts.
352 The experiments in which the extract obtained from the seeds and PVP were carried out

353 yielded viable results for the proportions (1:1), (2:1), and (1:2). No complete coating was
354 achieved in any of the experiments carried out in the SAS study. When the extract to PVP
355 ratio was increased to (1: 4), the resulting material showed particle plasticization.

356 The pulp extract was processed by SAS with PVP in different proportions, the
357 resulting materials showed no significant differences between each other by mean of TG
358 analysis. Although the precipitated content has shown the typical coloration conferred by
359 the presence of anthocyanin, and the FTIR analysis indicated the presence of both
360 substances (extract and PVP) on it, the materials obtained did not have a substantial total
361 polyphenols content. In addition to the characteristic colour, the FTIR analysis showed
362 co-precipitation of the material. It was not possible to carry out a study to analyse the pulp
363 extract processed without polymer addition, once the material could not be recovered
364 after SAS processing.

365 The importance of the solid to liquid ratio on the particle formation was
366 demonstrated within the study of seeds extract co-precipitation. This parameter not only
367 affects the distribution of the co-precipitate but also implies the particle size.

368

369

370

371

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373

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378

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449 **Figure Caption:**

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451 Figure 1 – Complete SAS flow diagram

452 Figure 2 – Seeds-extract co-precipitated with PVP at (1:1) ratio, 3.33 g/L.

453 Figure 3 – Seeds-extract co-precipitated with PVP at (1:1) ratio, 5.00 g/L.

454 Figure 4 – Seeds-extract co-precipitated with PVP at (1:1) ratio, 2.50 g/L.

455 Figure 5 – Pulp-extracted and PVP processed at (1:1) ratio, (a) 5.00 g/L, and (b) 3.33 g/L.

456 Figure 6 – Thermogravimetric (TG) curve obtained for seeds-extract co-precipitated in
457 different proportions with PVP.

458 Figure 7 – Derivative Thermogravimetry (DTG) curve of co-precipitated seed-extract and
459 PVP material.

460 Figure 8 – FTIR of PVP, original seeds-extract, and encapsulated material: characteristic
461 peaks of (I)seeds-extract and (II)PVP.

462 Figure 9 – FTIR of PVP, original pulp-extract, and encapsulated material: characteristic
463 peaks of (I)pulp-extract and (II)PVP.

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474 **Tables:**

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476 **(Table 1)**

477 Table 1 – Experimental conditions and total polyphenol content of SAS-
 478 processed particles.

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Extract (Material)	Polymer	Solvent	Polymer: extract Ratio	Initial solution concentration (g/L)	TPC content mg GAE/g particle	SD
Seeds (unprocessed)	-	-	-	-	261	3
Seeds	-	Ethanol	-	3.33	500	5
Seeds	PVP	Ethanol	1:1	5.00	78	3
Seeds	PVP	Ethanol	1:1	3.33	211	8
Seeds	PVP	Ethanol	1:1	2.50	80	4
Seeds	Pluronic	Acetone	1:1	3.33	132	8
Seeds	PVP	Ethanol	2:1	5.00	142	6
Seeds	PVP	Ethanol	2:1	3.33	187	8
Seeds	PVP	Ethanol	1:2	5.00	54	3
Seeds	PVP	Ethanol	1:2	3.33	72	3
Pulp (unprocessed)	-	-	-	-	108	3
Pulp	-	Ethanol	-	-	-	-
Pulp	PVP	Ethanol	2:1	3.33	7	>1
Pulp	PVP	Ethanol	1:1	5.00	27	1
Pulp	PVP	Ethanol	1:1	3.33	31	2
Pulp	PVP	Ethanol	1:1	2.50	35	>1

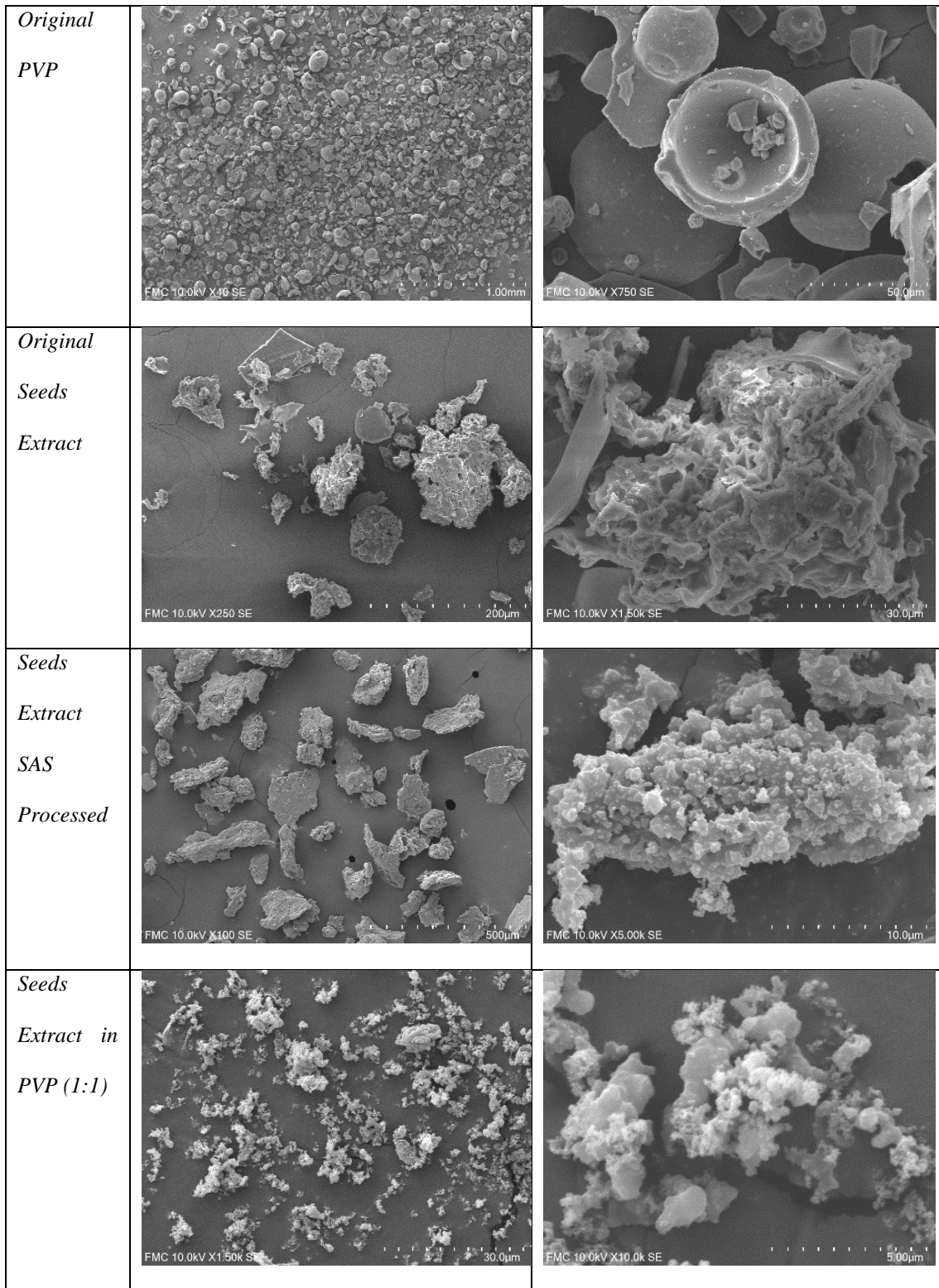
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482 **(Table 2)**

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484 Table 2- Comparative of SEM images of encapsulated material, PVP, seeds-extract, and
 485 seeds-extract processed by SAS



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