

**“Biogas valorization via continuous polyhydroxybutyrate production  
by *Methylocystis hirsuta* in a bubble column bioreactor”**

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## 1 **Abstract**

2 Creating additional value out of biogas during waste treatment has become a priority in  
3 past years. Biogas bioconversion into valuable bioproducts such as biopolymers has  
4 emerged as a promising strategy. This work assessed the operational feasibility of a  
5 bubble column bioreactor (BCB) implemented with gas recirculation and inoculated  
6 with a polyhydroxybutyrate (PHB)-producing strain using biogas as substrate. The BCB  
7 was initially operated at empty bed residence times (EBRTs) ranging from 30 to 120  
8 min and gas recirculation ratios (R) from 0 to 30 to assess the gas-to-liquid mass  
9 transfer and bioconversion of CH<sub>4</sub>. Subsequently, the BCB was continuously operated  
10 at a R of 30 and an EBRT of 60 min under excess of nitrogen and nitrogen feast-famine  
11 cycles of 24h:24h to trigger PHB synthesis. Gas recirculation played a major role in  
12 CH<sub>4</sub> gas-liquid transfer, providing almost fourfold higher CH<sub>4</sub> elimination capacities  
13 (~41 g CH<sub>4</sub> m<sup>-3</sup> h<sup>-1</sup>) at the highest R and EBRT of 60 min. The long-term operation  
14 under N excess conditions entailed nitrite accumulation (induced by O<sub>2</sub> limiting  
15 conditions) and concurrent methanotrophic activity inhibition above ~60 mg N-NO<sub>2</sub><sup>-</sup> L<sup>-1</sup>.  
16 <sup>1</sup>. Adjusting the N-NO<sub>3</sub><sup>-</sup> supply to match microbial N demand successfully prevented  
17 nitrite accumulation. Finally, the N feast-famine 24h:24h strategy supported a stable  
18 CH<sub>4</sub> conversion with a removal efficiency of 70% along with a continuous PHB  
19 production, which yielded PHB accumulations of 14.5 ± 2.9% (mg PHB mg<sup>-1</sup> total  
20 suspended solids × 100). These outcomes represent the first step towards the integration  
21 of biogas biorefineries into conventional anaerobic digestion plants.

22

23 **Keywords:** bioplastics; biorefinery; gas-liquid mass transfer; methane conversion;  
24 methanotrophic bacteria; polyhydroxyalkanoate production

25 **Abbreviation list:**

26 BCB: bubble column bioreactor

27 CH<sub>4</sub>-EC: methane elimination capacity

28 CH<sub>4</sub>-RE: methane removal efficiency

29 EBRT: empty bed residence time

30 MOB: methane oxidizing bacteria

31 MSW: municipal solid waste

32 NMS: nitrate mineral salt medium

33 PCO<sub>2</sub>: volumetric carbon dioxide production rate

34 PHAs: polyhydroxyalkanoates

35 PHB: poly-3-hydroxybutyrate

36 TN: total nitrogen

37 TSS: total suspended solids

38 R: gas recirculation ratio

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## 46        **1. Introduction**

47        Polyhydroxyalkanoates (PHAs), which exhibit similarity in their mechanical  
48        properties to polypropylene and polyethylene, are regarded as an attractive alternative  
49        for replacing oil-based plastics due to the striving necessity of meeting the increasing  
50        societal demand for more environmentally friendly materials (Lee & Na, 2013). These  
51        renewable biopolyesters, with the added advantages of biocompatibility and  
52        biodegradability, are synthesized intracellularly as carbon and energy storage by a broad  
53        collection of microorganisms under nutrient deprivation and carbon surplus conditions  
54        (Castilho et al., 2009; Myung et al., 2017; Narancic et al., 2018). Although the market  
55        size of PHA is foreseen to quadruple by 2023, it is still relatively small. Indeed, PHA  
56        represented only 1.4% of the global biopolymer market by 2018, which accounted for  
57        2.1 million tonnes (European Bioplastics, 2018). Its expansion is currently hampered by  
58        the significant contribution of the carbon feedstock (usually sugars, vegetable oils and  
59        fatty acids) acquisition to the total production costs (up to 50%) (Koller et al., 2017).  
60        Thus, PHAs prices (4 to 20 € kg<sup>-1</sup>) are nowadays up to fifteen-fold higher than those of  
61        their fossil counterparts (Blunt et al., 2018; Cantera et al., 2018).

62        On this scenario, the use of industrial by-products or wastes such as biogas as a  
63        feedstock represents an opportunity to decrease the cost of PHAs production (Cal et al.,  
64        2016; Strong et al. 2016). Biogas resulting from the anaerobic decomposition of the  
65        biodegradable fraction of organic waste in anaerobic digestion plants or landfills is  
66        primarily made up of methane (30-70%), carbon dioxide (20-50%) and hydrogen  
67        sulfide (< 2%) (Nikiema et al., 2007; Muñoz et al., 2015). Gasification and methanation  
68        of wood or recalcitrant organic waste can also generate a biomethane with CH<sub>4</sub>  
69        concentration higher than 90% (IEA, 2018). Methane, aside from being a potent  
70        greenhouse gas, has been traditionally regarded as a green energy vector and is

71 increasingly used as a C source in industrial biotechnology. Indeed, the integration of  
72 biogas into biorefineries for manufacturing high added value bioproducts such as PHA,  
73 protein or ectoine is increasingly drawing attention due to the recent stabilization of the  
74 biogas industry expansion (Mühlemeier et al., 2018).

75 PHA biosynthesis from CH<sub>4</sub> relies on the ability of type II-methane oxidizing  
76 bacteria (MOB), also referred as methanotrophs, to synthesize PHA granules under  
77 growth limiting conditions (Pieja et al., 2011a; Rostkowski et al., 2013; Zhang et al.,  
78 2017). Type II-MOB metabolize methane and one-carbon compounds via the serine  
79 cycle (Pieja et al., 2011a). When methane is used as the sole C and energy feedstock,  
80 cells naturally synthesize the short-chain-length PHA poly-3-hydroxybutyrate (PHB).  
81 Among type II-methanotrophs, the strain *Methyloscystis hirsuta* CSC1 has drawn  
82 interest due to its high PHB-accumulating ability and metabolic plasticity (Bordel et al.,  
83 2019b). In a recent work, López et al. (2018) obtained comparable cell growth and PHB  
84 accumulation (43%) when synthetic biogas rather than CH<sub>4</sub> was used as carbon source  
85 in *M. hirsuta* regardless of the presence of hydrogen sulfide. Nevertheless, to the best of  
86 the authors' knowledge, the potential of biogas as a feedstock for the continuous  
87 production of PHB and the constraints associated to a long-term continuous operation  
88 have not been yet reported.

89 The major operational constraint associated to CH<sub>4</sub>/biogas bioconversion  
90 technologies is the poor mass transfer of O<sub>2</sub> and CH<sub>4</sub> (Henry's law constants ( $k_H$ ) of  
91  $1.3 \cdot 10^{-3}$  and  $1.4 \cdot 10^{-3}$  M atm<sup>-1</sup> at standard conditions, respectively) (Sander, 2015). In  
92 this regard, turbulent contactors such as bubble column bioreactors (BCBs) engineered  
93 with innovative gas-liquid mass transfer strategies (i.e. the utilization of a non-aqueous  
94 phase or the implementation of internal gas recirculation) can support an enhanced  
95 methane biodegradation (Cantera et al., 2016; Rocha-Rios et al., 2011). Moreover, this

96 type of suspended-growth bioreactors allows an easy biomass harvesting and bioproduct  
97 downstream processing (López et al., 2019).

98 This study aims at optimizing the biogas residence time and the internal  
99 recirculation rate to maximize CH<sub>4</sub> mass transfer and at assessing the long-term (> 4  
100 weeks of stable operation) production of PHB from biogas by *M. hirsuta* in a  
101 continuous BCB equipped with gas recirculation.

## 102 **2 Materials and methods**

103 To fulfill the above-mentioned objectives, the experimental research was structured  
104 into two main assays that were carried out in a bubble column bioreactor whose  
105 configuration was described in section 2.2.1. A first approach pursued the optimization  
106 of operating parameters through a mass transfer test in which 18 different conditions for  
107 EBRT and R (2.2.2) were assayed. A second approach aimed at investigating the  
108 process stability (2.2.3) under non-nutrient limited conditions prior the implementation  
109 of sequential nitrogen feast-famine cycles (2.2.4) to induce the PHB synthesis. Sections  
110 2.2.3 and 2.2.4 were performed at the same EBRT and R conditions.

### 111 **2.1 Chemicals, culture media and inoculum**

112 **2.1.1 Chemicals.** The chemicals used for PHB extraction (trichloromethane (≥ 99%), 1-  
113 propanol (99.7%), benzoic acid (≥ 99.5%), and hydrochloric acid (37% w/v)) and for  
114 the culture medium preparation were acquired from PanReac AppliChem (Spain),  
115 except KNO<sub>3</sub>, which was purchased from COFARCAS (Spain). Poly(3-  
116 hydroxybutyrate-co-3-hydroxyvalerate) (PHBV) with a mole fraction 3HB/3HV of  
117 88/12 (≥ 99.99%) was acquired from Sigma-Aldrich (USA). O<sub>2</sub> (≥ 99.5%), CH<sub>4</sub> (≥

118 99.995%), He ( $\geq 99.5\%$ ), and a synthetic biogas mixture containing 70% of CH<sub>4</sub> and  
119 30% of CO<sub>2</sub> were provided by Abelló Linde S.A. (Spain).

120 **2.1.2 Culture medium.** Unless otherwise specified, a nitrate mineral salt (NMS)  
121 medium containing the following macronutrients (g L<sup>-1</sup>) was employed: 0.2  
122 CaCl<sub>2</sub>·2H<sub>2</sub>O, 1.0 KNO<sub>3</sub>, 1.1 MgSO<sub>4</sub>·7H<sub>2</sub>O and the following trace elements (mg L<sup>-1</sup>):  
123 0.01 NiCl<sub>2</sub>·6H<sub>2</sub>O, 0.02 MnCl<sub>2</sub>·4H<sub>2</sub>O, 0.03 CoCl<sub>2</sub>, 0.015 H<sub>3</sub>BO<sub>3</sub>, 0.38 Fe-EDTA, 0.3  
124 Na<sub>2</sub>EDTA·2H<sub>2</sub>O, 0.4 Na<sub>2</sub>MoO<sub>4</sub>·2H<sub>2</sub>O, 0.4 ZnSO<sub>4</sub>·7H<sub>2</sub>O, 0.5 FeSO<sub>4</sub>·7H<sub>2</sub>O and 1.0  
125 CuSO<sub>4</sub>·5H<sub>2</sub>O. The NMS was stored in borosilicate glass bottles and autoclaved (121 °C,  
126 30 min). After cooling the sterile NMS down to room temperature, 10 mL of a sterile  
127 buffer solution (72 g L<sup>-1</sup> Na<sub>2</sub>HPO<sub>4</sub>·12H<sub>2</sub>O and 26 g L<sup>-1</sup> KH<sub>2</sub>PO<sub>4</sub>) per liter of NMS  
128 medium were added to adjust its pH to 6.8.

129 **2.1.3 Strain and inoculum preparation.** A stock culture of *Methylocystis hirsuta*  
130 CSC1 (DSM 18500) was purchased from Leibniz-Institut DSMZ (Germany) and stored  
131 at 4°C until use. The cultivation of *M. hirsuta*, which was grown up to a concentration  
132 of  $0.54 \pm 0.03$  g L<sup>-1</sup> prior to inoculation of the bioreactors, was conducted in two stages  
133 under strictly sterile conditions based on López et al. (2018) procedure. Initially,  
134 gastight serum vials of 125 mL containing 50 mL of NMS medium were inoculated  
135 from the DSMZ vial at 10% (v v<sup>-1</sup>) under an O<sub>2</sub>:CH<sub>4</sub> atmosphere (66.7:33.3% (v v<sup>-1</sup>)).  
136 The cultures were incubated in an orbital shaker at 200 rpm and 30°C for 8 days. Once  
137 the cultures were metabolically active, the headspace atmosphere was restored  
138 systematically up to a maximum of 5 times every 24 h. To that end, filtered oxygen was  
139 gassed for 5 min and replaced (25 mL) with methane afterwards with a 50 mL gastight  
140 syringe (Hamilton 1050 TLL, USA). Finally, aliquots of 10 mL of this active culture  
141 were transferred to sterile serum bottles (2.2 L) containing 0.4 L of NMS medium  
142 sealed with aluminum caps and chlorobutyl rubber stoppers under an O<sub>2</sub>:CH<sub>4</sub>:CO<sub>2</sub>

143 atmosphere of 58.3:29.2:12.5% (v v<sup>-1</sup>). The headspace atmosphere was obtained by  
144 flushing for 3 min a gas mixture composed of biogas and oxygen with the above  
145 mentioned composition from a 100 L-Tedlar gas sampling bag (Sigma-Aldrich, USA).  
146 The cultures were grown under continuous stirring at 300 rpm (Thermo Scientific  
147 Variomag Multipoint 6, USA) and 25°C in a thermostated room for 10-12 days until  
148 complete methane depletion.

149

## 150 **2.2 Experimental procedure**

### 151 **2.2.1 Experimental set-up**

152 The study herein presented was carried out in a bench-scale bubble column  
153 bioreactor (BCB) implemented with gas recirculation to ensure a high CH<sub>4</sub> and O<sub>2</sub> mass  
154 transfer to the cultivation broth (**Fig. 1**). The bioreactor, with a working volume of 2.5  
155 L, was equipped with a set of three micropore stainless steel diffusers (2 µm, Supelco,  
156 USA) and a magnetic stirrer (Agimatic S, JP Selecta, Spain, 500 rpm) located at the  
157 bottom of the column to ensure an adequate mixing throughout the column. A gas  
158 mixture composed of atmospheric air and synthetic biogas was continuously sparged  
159 into the bioreactor through the diffusers. The biogas stream was regulated by a mass  
160 flow controller (GFC17, Aalborg<sup>TM</sup>, USA), whereas the atmospheric air was supplied  
161 by an air compressor and controlled by a rotameter to deliver a gas mixture with O<sub>2</sub>:CH<sub>4</sub>  
162 ratios ranging from 1.5:1 to 2:1. A condenser (maintained at 10°C) was installed at the  
163 internal gas recirculation line in order to prevent operational problems derived from  
164 water condensation. The BCB was operated at 25°C in a temperature-controlled room.

165

<Figure 1>

166

167 **2.2.2 CH<sub>4</sub>/O<sub>2</sub> mass transfer optimization test.**

168 The bioreactor was first inoculated with a fresh *Methylocystis hirsuta* CSC1 culture  
169 resulting in an initial total suspended solids (TSS) concentration of 0.5 g L<sup>-1</sup>. The BCB  
170 was initially operated at an EBRT of 30 min with a CH<sub>4</sub> content of 90 ± 8 g CH<sub>4</sub> m<sup>-3</sup> (14  
171 ± 1 % v v<sup>-1</sup>) and without gas recirculation for four weeks. In this period, biomass  
172 concentration reached 3 g L<sup>-1</sup>, a concentration that was maintained under all operational  
173 conditions tested to guarantee that the process was not biologically limited. For this  
174 purpose, aliquots of 500 mL of culture broth from the BCB were daily centrifuged  
175 (10000 rpm, 8 min) and replaced by fresh NMS. In order to maintain a constant biomass  
176 concentration of ~3.5 g TSS L<sup>-1</sup> in the culture broth, the harvested biomass was either  
177 partially returned to the system or discarded. The diffusers were replaced when the  
178 pressure drop exceeded 0.5 bar. The nitrogen concentration of the NMS supplied was  
179 adjusted from 1 to 7 g L<sup>-1</sup> KNO<sub>3</sub> at each operational condition tested to avoid nitrogen  
180 limitation in the culture broth. Subsequently, the influence of the EBRT and the gas  
181 recirculation ratio (hereinafter referred to as R) on CH<sub>4</sub> biodegradation under balanced  
182 growth conditions was assessed (**Table 1**). Each operational condition was maintained  
183 for a period of 5-8 days, which ensured steady state operation regarding the liquid and  
184 gas phases.

185

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187

< **Table 1** >

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### 191 **2.2.3 Methane biodegradation under nitrogen surplus conditions in the BCB**

192 The bioreactor was inoculated with the above mentioned strain at  $90 \pm 10$  mg TSS  
193  $L^{-1}$  and operated under the optimum operational conditions selected in the previous test  
194 (EBRT of 60 min,  $R = 30$ , methane inlet load of  $68 \pm 8$  g  $CH_4$   $m^{-3} h^{-1}$  and an  $O_2:CH_4$   
195 molar ratio of  $1.8 \pm 0.3$ ) in order to identify long-term microbial and mechanical  
196 limitations during continuous  $CH_4$  abatement. Thus,  $CH_4$  biodegradation was  
197 investigated under nitrogen excess for 50 days (from day 0 to day 35, and from day 51  
198 to day 66) and nitrogen limiting conditions for 15 days (from day 36 to day 50) using a  
199 modified NMS medium with  $N-NO_3^-$  concentrations of 552 and 276 mg  $L^{-1}$ ,  
200 respectively. During the first week of operation, the biomass contained in 500 mL of  
201 culture broth was daily collected and returned to the BCB re-suspended into 500 mL of  
202 fresh NMS. From day 8 onwards, no biomass was returned to the BCB so that the  
203 biomass concentration was maintained at  $\sim 3$  g  $L^{-1}$ .

### 204 **2.2.4 Biogas utilization coupled to continuous PHB production in the BCB**

205 This test aimed at assessing the continuous PHB biosynthesis from biogas by *M.*  
206 *hirsuta*. The system was initially inoculated at  $190 \pm 0$  mg TSS  $L^{-1}$  with *M. hirsuta* and  
207 operated at an EBRT of 60 min, a  $R$  of 30 and a methane inlet load of  $60 \pm 3$  g  $CH_4$   $m^{-3}$   
208  $h^{-1}$  (corresponding to an  $O_2:CH_4$  ratio of  $2.1 \pm 0.1$ ).  $CH_4$  biodegradation was initially  
209 investigated under nitrogen-balanced conditions for 32 days of operation (using a NMS  
210 with a  $N-NO_3^-$  content of 345 mg  $L^{-1}$ ). By day 33, a nitrogen-free mineral salt (NFMS)  
211 medium was supplied to deplete the nitrogen in the BCB, and subsequently 15  
212 sequential nitrogen feast-famine cycles of 48 h (i.e. 24 h under nitrogen balanced  
213 growth conditions followed by 24 h under nitrogen starvation) were applied to trigger  
214 PHB biosynthesis. To this end, 500 mL  $d^{-1}$  of modified NMS medium (345 mg  $N-NO_3^-$

215 L<sup>-1</sup>) or NFMS medium were alternatively provided corresponding to a dilution rate of  
216 0.2 d<sup>-1</sup>.

217 Gas flow rate, pressure drop in the gas diffusers and gas composition in the inlet and  
218 outlet streams (CH<sub>4</sub>, CO<sub>2</sub> and O<sub>2</sub>) were monitored daily. Total dissolved nitrogen (TN),  
219 NO<sub>2</sub><sup>-</sup> and NO<sub>3</sub><sup>-</sup> in the culture broth were also recorded. Culture samples (40 mL) were  
220 collected for the determination of TSS, pH and PHAs. The pellets from the  
221 centrifugation of 3 mL of culture broth samples (10000 rpm, 10 min) were stored at -  
222 20°C prior PHAs analysis.

### 223 **2.3 Analytical procedures**

224 CH<sub>4</sub>, CO<sub>2</sub> and O<sub>2</sub> gas concentrations were measured in duplicate in a gas  
225 chromatograph coupled with a thermal conductivity detector (Bruker 430, Bruker  
226 Corporation, USA) and equipped with CP-Molsieve 5A and CP-PoraBOND Q columns.  
227 The pressure drop in the BCB was monitored with a pressure sensor (PI1696, Ifm  
228 Electronic, Germany). Outlet gas flow rate was estimated by using the water  
229 displacement method.

230 Biomass concentration, expressed as TSS, was determined according to the 2540  
231 method (APHA, 2017) using 0.45 µm pore size filters (Merck, Germany). Biomass  
232 density was also estimated using optical density measurements at 600 nm with a UV-  
233 2550 spectrophotometer (Shimadzu, Japan). A Basic 20 pH meter (Crison, Spain) was  
234 used for the measurement of pH.

235 NO<sub>2</sub><sup>-</sup> and NO<sub>3</sub><sup>-</sup> concentrations were analyzed by ion chromatography using a Waters  
236 432 HPLC conductivity detector (Waters Corporation, USA). TN and total organic  
237 carbon (TOC) concentrations were quantified simultaneously in a TOC-V analyzer  
238 equipped with a Shimadzu TNM-1 unit.

239 The biopolymer extraction procedure for PHB quantification via gas  
240 chromatography-mass spectrometry (GC-MS) was conducted according to Rodríguez et  
241 al. (2020).

## 242 **2.4 Calculation**

### 243 **2.4.1 Performance indicators of the BCB**

244 The elimination capacity ( $CH_4 - EC$ ), removal efficiency ( $CH_4 - RE$ ) and the  
245 volumetric carbon dioxide production rate ( $PCO_2$ ) are defined as:

$$246 \quad CH_4 - EC = \frac{Q \cdot (C_{CH_4, in} - C_{CH_4, out})}{V_R} \quad (\text{Eq.1})$$

$$247 \quad CH_4 - RE (\%) = \frac{C_{CH_4, in} - C_{CH_4, out}}{C_{CH_4, in}} \times 100 \quad (\text{Eq.2})$$

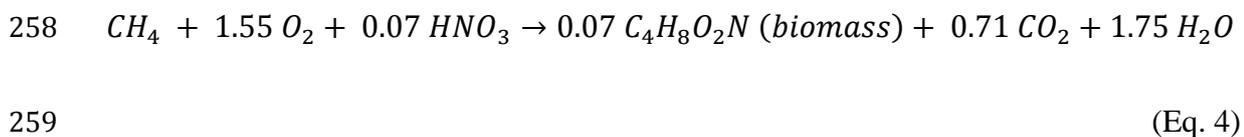
$$248 \quad PCO_2 = \frac{Q \cdot (C_{CO_2, out} - C_{CO_2, in})}{V_R} \quad (\text{Eq.3})$$

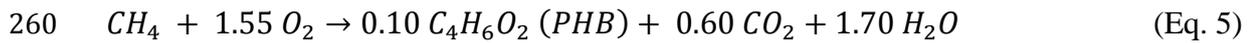
249 where  $C_{in}$  and  $C_{out}$  are the inlet and outlet concentration ( $\text{g m}^{-3}$ ), respectively,  $Q$  is the  
250 inlet gas flow ( $\text{m}^3 \text{h}^{-1}$ ) and  $V_R$  ( $\text{m}^3$ ) is the working volume of the bioreactor.

### 251 **2.4.2 Stoichiometry and theoretical PHB productivity in the BCB**

252 According to literature, the theoretical biomass and PHB yields in type-II  
253 methanotrophs are  $0.46 \text{ g biomass g}^{-1} \text{ CH}_4$  (using nitrate as nitrogen source) (Asenjo  
254 and Suk, 1986) and  $0.54 \text{ g PHB g}^{-1} \text{ CH}_4$ , respectively (Yamane, 1993; Khosravi-Darani  
255 et al., 2013).

256 The overall equations for biomass growth and PHB accumulation supporting these  
257 yields are given below.





261 The sum of Eq. 1 and Eq.2 taking into consideration the duration of both growth and  
 262 accumulation phases within the cycles (1/2) gives the theoretical PHB yield ( $Y_{PHB}^{th}$ ) of  
 263 the global process.

264 Thus, the theoretical PHB productivity ( $P_{th}$ ) for a 24h:24h growth:accumulation cycle  
 265 was estimated using the following formula:

266  $P_{PHB}^{th} (kg m^{-3} d^{-1}) = \frac{(1/2) \cdot 0.54 \cdot CH_4EC \cdot 24 (h/d)}{1000 (g/kg)}$  (Eq. 6)

267 where 0.54 is the  $Y_{PHB}^{th}$  (g PHB produced  $g^{-1}$   $CH_4$  removed) and  $CH_4EC$  is the methane  
 268 elimination capacity ( $g CH_4 m^{-3} h^{-1}$ ).

269

### 270 **3 Results and discussion**

#### 271 **3.1 $CH_4$ mass transfer optimization in the BCB: Influence of the empty bed** 272 **residence time and the gas recirculation on $CH_4$ degradation**

273 BCB operation at EBRTs of 30, 60 and 120 min with no internal gas recirculation  
 274 showed  $CH_4$ -elimination capacities ( $CH_4$ -ECs) of  $29.8 \pm 2.0$ ,  $11 \pm 1.7$  and  $6.9 \pm 1.8$  g  
 275  $CH_4 m^{-3} h^{-1}$ , with associated  $CH_4$ -removal efficiencies ( $CH_4$ -REs) of  $13.3 \pm 0.6$ ,  $12.3 \pm$   
 276  $1.7$  and  $12.7 \pm 1.2\%$ , respectively (**Fig. 2a and 2b**). This suggests that the enhancement  
 277 in the turbulence of the cultivation broth mediated by the decrease in the EBRT caused  
 278 a positive effect on the volumetric mass transfer coefficient ( $K_{LaCH_4}$ ) and consequently,  
 279 on the  $CH_4$ -EC.

280 Internal gas recirculation has been reported as a promising strategy for enhancing  
 281  $CH_4$ -biodegradation in biotrickling filters (> 2.5 times) and BCBs (> 2.1 times) during

282 the treatment of CH<sub>4</sub>-diluted air emissions (Estrada et al., 2014; García-Pérez et al.,  
283 2018). This approach allows decoupling the actual gas residence time in the reactor  
284 from the liquid turbulence in the bioreactor. Thus, at an EBRT of 30 min, the CH<sub>4</sub>-EC  
285 increased to 42.2 ± 1.0, 55.1 ± 1.7, 54.3 ± 1.6, 57.6 ± 1.4 and 73.8 ± 2.1 g CH<sub>4</sub> m<sup>-3</sup> h<sup>-1</sup> at  
286 R of 2.5, 5, 10, 15 and 30, respectively (**Fig. 2a**). Consequently, CH<sub>4</sub>-REs increased to  
287 20.5 ± 0.8, 25.0 ± 1.4, 27.1 ± 3.5, 32.7 ± 2.0 and 39.0 ± 3.6% at these R values (**Fig.**  
288 **2b**). Similarly, Rocha-Rios et al. (2011) reported an enhancement of 51% in the CH<sub>4</sub>  
289 biodegradation performance of an airlift loop bioreactor by increasing the gas  
290 recirculation rate from 0 to 1 volumes per minute (vvm).

291 At EBRTs of 60 and 120 min, the CH<sub>4</sub>-EC increased by a factor of 3.7 (from 11.0 ±  
292 1.7 to 41.1 ± 0.4 g CH<sub>4</sub> m<sup>-3</sup> h<sup>-1</sup>) and 3.2 (from 6.9 ± 1.8 to 22.2 ± 0.7 g CH<sub>4</sub> m<sup>-3</sup> h<sup>-1</sup>),  
293 respectively (**Fig. 2b**), at the highest R applied. A similar behavior was observed during  
294 the biodegradation of CH<sub>4</sub>-laden emissions (4% v v<sup>-1</sup>) in a BCB with internal gas  
295 recirculation. However, the lower CH<sub>4</sub> gas-liquid gradients during the biodegradation of  
296 diluted CH<sub>4</sub> emissions resulted in lower CH<sub>4</sub>-ECs (i.e.: 35.2 ± 0.4 g CH<sub>4</sub> m<sup>-3</sup> h<sup>-1</sup> at an  
297 EBRT of 30 and R of 15) (García-Pérez et al., 2018).

298 < **Figure 2** >

299 It can be inferred that internal gas recirculation induces opposite effects by  
300 increasing system turbulence at the expense of reducing CH<sub>4</sub> gas-liquid gradient in the  
301 column. Results in **Fig. 3** indicates that shorter gas contact times in the system mediated  
302 greater elimination capacities despite the negative effects which may be associated to a  
303 high turbulence such as bubble coalescence or *eddies* (Stone et al., 2017). The  
304 correlation observed is explained by the fact that the input of energy into the system  
305 reduces the liquid film and enhance the superficial contact area by breaking the bubbles  
306 (Kraakman et al., 2011). For instance, the operation of the BCB in the absence of gas

307 recirculation at an EBRT of 30 min and at an EBRT of 120 min with R of 2.5 showed  
308 similar virtual EBRTs (30 and 34.3 min, respectively), but a different CH<sub>4</sub>-degradation  
309 performance with ECs of  $29.8 \pm 2.0$  and  $11.7 \pm 0.5$  g CH<sub>4</sub> m<sup>-3</sup> h<sup>-1</sup> and CH<sub>4</sub>-REs of  $13.3$   
310  $\pm 0.6$  and  $23.1 \pm 1.5\%$ , respectively.

311 **<Figure 3>**

312 Overall, the elimination capacities herein recorded were superior to the  $\sim 20$  g m<sup>-3</sup>  
313 h<sup>-1</sup> achieved in an internal loop airlift reactor (Rocha et al., 2011). Furthermore, there  
314 was a satisfactory match with the  $\sim 75$  and  $22$  g m<sup>-3</sup> h<sup>-1</sup> (REs of 34 and 15%) achieved in  
315 a stirred tank reactor (at 800 rpm) and a trickling bed reactor both operated with a  
316 similar methane load. Conversely, the supplementation of the stirred tank reactor with  
317 10% silicon oil resulted in a higher EC ( $106$  g m<sup>-3</sup> h<sup>-1</sup>) than the maximum EC reached  
318 in this work ( $74$  g m<sup>-3</sup> h<sup>-1</sup>) (Rocha et al., 2009). Despite having proved to enhance the  
319 gas-liquid transfer at high stirring rates in turbulent contactors (Kraakman et al., 2011),  
320 the addition of a non-aqueous phase is up to date not suitable for biomass valorization  
321 (Stone et al., 2017).

322 The highest CH<sub>4</sub>-REs were targeted as selection criteria in order to maximize the  
323 utilization of CH<sub>4</sub> from biogas as a substrate for PHB biosynthesis (Pérez et al., 2019).  
324 Therefore, EBRTs of 60 and 120 min with R of 30 exhibited the highest potential for  
325 CH<sub>4</sub> bioconversion with CH<sub>4</sub>-REs of  $45.1 \pm 1.2$  and  $47.8 \pm 3.5\%$  and CH<sub>4</sub>-ECs of  $41.1 \pm$   
326  $0.4$  and  $22.2 \pm 0.7$  g CH<sub>4</sub> m<sup>-3</sup> h<sup>-1</sup>, respectively. In the light of the similar CH<sub>4</sub>-REs at  
327 both EBRTs, the operation at an EBRT of 60 min with a R of 30 was selected as the  
328 most appropriate condition for PHB production given the higher CH<sub>4</sub>-EC, which would  
329 ultimately result in a significant reduction of equipment costs and enhanced bioreactor  
330 productivities. This selection was also supported by a carbon footprint analysis (**Fig.**

331 **S2**), in which the selected operating conditions mediated the largest CO<sub>2</sub> equivalents  
332 reduction (30%) compared to a scenario where the biogas was vented.

333 The mass transfer experiments here presented were performed with a sufficiently  
334 high biomass concentration (>3 g TSS L<sup>-1</sup>) to guarantee that CH<sub>4</sub> gas-liquid mass  
335 transfer was the limiting step of the process. Likewise, the high dilution rate applied  
336 prevented nutrient-limiting conditions and secondary metabolites accumulation, which  
337 could negatively affect CH<sub>4</sub> biodegradation performance. In this regard, the minimum  
338 TN (41.56 ppm) and maximum TOC (74.6 ppm) concentrations corresponded to the  
339 maximum CH<sub>4</sub>-ECs achieved at an EBRT of 30 min and R of 30. Accordingly, the CH<sub>4</sub>  
340 mineralization ratio, expressed as the volumetric CO<sub>2</sub> production rate to methane  
341 elimination capacity ratio (PCO<sub>2</sub>/CH<sub>4</sub>-EC), remained constant at 2.4 ± 0.2, 2.2 ± 0.2 and  
342 2.4 ± 0.3 at EBRTs of 30, 60 and 120 min, respectively, which suggested a balanced  
343 methanotrophic metabolism along the entire experiment.

### 344 **3.2 Effect of the N supply on the continuous CH<sub>4</sub> abatement**

345 The CH<sub>4</sub>-EC rapidly increased up to 53.0 ± 2.3 g CH<sub>4</sub> m<sup>-3</sup> h<sup>-1</sup> concomitantly with an  
346 increase in PCO<sub>2</sub> up to 119.9 ± 0.1 g CO<sub>2</sub> m<sup>-3</sup> h<sup>-1</sup> following BCB inoculation (**Fig. 4a**).  
347 Both CH<sub>4</sub>-EC and PCO<sub>2</sub> remained stable at 57.1 ± 3.6 g CH<sub>4</sub> m<sup>-3</sup> h<sup>-1</sup> and 128.7 ± 11.4 g  
348 CO<sub>2</sub> m<sup>-3</sup> h<sup>-1</sup>, respectively, during the first 20 days. Biomass concentration increased up  
349 to 4.8 ± 0.1 g L<sup>-1</sup> by day 8 and steadily decreased to 2.4 ± 0.0 g L<sup>-1</sup> by day 20 (**Fig 4c**).  
350 Unexpectedly, CH<sub>4</sub>-EC and PCO<sub>2</sub> experienced a slight decrease by day 20 (**Fig. 4a**).

351 Thus, all biomass was collected by centrifugation and resuspended into fresh NMS  
352 medium on day 23 to prevent the culture from a potential accumulation of non-desired  
353 metabolites. Despite the system showed an almost complete recovery within the two  
354 next days, a sharp drop in CH<sub>4</sub> biodegradation performance occurred again. A new

355 steady state with CH<sub>4</sub>-EC of  $7.4 \pm 1.5 \text{ g CH}_4 \text{ m}^{-3} \text{ h}^{-1}$  and PCO<sub>2</sub> of  $21.5 \pm 4.2 \text{ g CO}_2 \text{ m}^{-3}$   
356 h<sup>-1</sup> was recorded from day 26 to 36 concomitantly with a gradual biomass wash-out  
357 (**Fig. 4c**).

358 The BCB was reinoculated by day 36 with *M. hirsuta* resuspended into NMS  
359 medium prepared with half of the nitrogen concentration ( $276 \text{ mg N-NO}_3^- \text{ L}^{-1}$ ). As  
360 shown in **Fig. 4a**, a consistent steady state was rapidly achieved and maintained for the  
361 next 14 days under no excess of nitrogen. When the N-NO<sub>3</sub><sup>-</sup> concentration of the NMS  
362 medium was restored to  $552 \text{ mg N-NO}_3^- \text{ L}^{-1}$ , and the system was no longer nitrogen  
363 limited, the CH<sub>4</sub>-EC remained constant for four days and eventually dropped from  $41.1$   
364  $\pm 2.7$  to  $5.2 \pm 0.7 \text{ g CH}_4 \text{ m}^{-3} \text{ h}^{-1}$ . PCO<sub>2</sub> was correlated to CH<sub>4</sub>-EC with a mineralization  
365 of  $79.2 \pm 7.6\%$  during both steady states achieved.

366 < **Figure 4** >

367 The analysis of the N species prevailing in the cultivation broth revealed that nitrate  
368 reduction to nitrite occurred from day 11 onwards, resulting in nitrite accumulation in  
369 the liquid broth, with a maximum concentration of  $83.3 \text{ mg N-NO}_2^- \text{ L}^{-1}$  by day 22. It can  
370 be inferred that nitrite accumulation (mediated by the O<sub>2</sub> limiting conditions in the  
371 cultivation broth at the low O<sub>2</sub>:CH<sub>4</sub> ratios used; 1.3:1-1.7:1) induced the inhibition of  
372 the methanotrophic metabolism, leading to a deterioration of the system performance  
373 despite the high NMS medium dilution rate applied ( $0.2 \text{ d}^{-1}$ ). Nitrite formation rapidly  
374 occurred again along with a decrease in CH<sub>4</sub>-EC after biomass resuspension into fresh  
375 NMS medium. Interestingly, when N supply was limited from day 36 to day 50, this  
376 accumulation did not occur, which allowed maintaining a stable process operation  
377 throughout this period. By day 51, the increase in N loading triggered again the  
378 accumulation of nitrite. This confirmed that nitrite accumulation and process inhibition  
379 was inherent to N surplus conditions.

380 In this context, the batchwise cultivation of *M. hirsuta* CSC1 in mineral salt medium  
381 prepared with nitrite as a nitrogen source ( $138 \text{ mg N-NO}_2^- \text{ L}^{-1}$ ) resulted in a complete  
382 growth inhibition (Rodríguez et al., 2020). The inhibitory effect of nitrite in type II-  
383 MOB was already reported in a previous study, where *Methylocystis* sp. growing in both  
384 nitrate- and ammonium-containing growth medium increased its doubling time by 65  
385 and 51%, respectively, when supplemented with  $2.5 \text{ mM NaNO}_2$  ( $35 \text{ mg N-NO}_2^- \text{ L}^{-1}$ )  
386 (Nyerges et al., 2010). Conversely, the type I strain *Methylochromobium album*  
387 maintained similar doubling times and  $\text{CH}_4$  removal rates to those of the control tests  
388 under identical conditions, with additional nitrous oxide production. Interestingly, the  
389 genome sequence of *Methylocystis hirsuta* CSC1 recently elucidated revealed that this  
390 strain possesses the mechanisms to conduct partially the denitrification pathway (Bordel  
391 et al., 2019a). To the best of the author's knowledge, this phenomenon has not been  
392 previously reported in continuous bioreactors devoted to methane abatement. In this  
393 regard, N supply limitation strategies can overcome this concurrent nitrite accumulation  
394 while inducing PHB synthesis in *M. hirsuta* (Rodríguez et al., 2020).

395 Finally, the pH of the cultivation broth during the stationary states achieved ( $7.2 \pm$   
396  $0.1$ ) suggests that the oxidation of methane releases basic metabolites that maintained  
397 the pH above the pH of the mineral salt medium ( $6.8 \pm 0.1$ ) despite the presence of  
398 buffer and the solubilization of the  $\text{CO}_2$  inherently present in biogas.

399

### 400 **3.3 Biogas utilization coupled to continuous PHB production in the BCB**

401 The BCB was continuously operated under the optimum operational parameters  
402 determined in the previous tests (EBRT of 60 min, R of 30 and a N supply of  $69 \text{ mg N-}$   
403  $\text{NO}_3^- \text{ d}^{-1} \text{ L}^{-1}$ ) to achieve a steady  $\text{CH}_4$  abatement along with a simultaneous biopolymer

404 production. Within the first two days of operation, the system achieved a stable CH<sub>4</sub>-EC  
405 of  $40.2 \pm 2.3 \text{ g CH}_4 \text{ m}^{-3} \text{ h}^{-1}$  corresponding to a CH<sub>4</sub>-RE of  $70.1 \pm 2.7\%$  (**Fig. 5a**). These  
406 CH<sub>4</sub>-EC values, which are in accordance with those achieved during the CH<sub>4</sub>/O<sub>2</sub> mass  
407 transfer test under comparable operational conditions ( $41.1 \pm 0.4 \text{ g CH}_4 \text{ m}^{-3} \text{ h}^{-1}$ ),  
408 supported a PCO<sub>2</sub> of  $78.5 \pm 6.3 \text{ g CO}_2 \text{ m}^{-3} \text{ h}^{-1}$  along with a mineralization of  $70.5 \pm$   
409  $4.7\%$ . As discussed in Section 3.1, the performance indicators of this study were  
410 comparable to those typically reported for poorly water soluble compounds ( $< 70\%$ )  
411 (Kraakman et al., 2011).

412 Additionally, the observed O<sub>2</sub>:CH<sub>4</sub> molar consumption at this stage was  $1.5 \pm 0.1$ ,  
413 which corresponded to the theoretical value ( $\sim 1.5$ -2) reported by Karthikeyan et al.  
414 (2015). The pH remained stable at  $7.2 \pm 0.2$  throughout the whole operation and the  
415 presence of nitrite was not detected, thus avoiding culture inhibition (**Fig. 5b**). This  
416 supports the hypothesis that dissimilatory nitrate reduction, where NO<sub>3</sub><sup>-</sup> is used as the  
417 electron acceptor for energy production, did not prevail over assimilatory nitrate  
418 reduction when N consumption and N supply are balanced. These outcomes have  
419 important implications for the application of this biotechnology at pilot or industrial  
420 scale under long-term operation. Biomass concentration was maintained at  $3.0 \pm 0.9 \text{ g}$   
421  $\text{TSS L}^{-1}$  from day 10 onwards, accounting for a biomass production of  $20.0 \pm 8.3 \text{ g TSS}$   
422  $\text{m}^{-3} \text{ h}^{-1}$  (**Fig. 5c**).

423 Process operation under sequential N feast-famine cycles from day 35 onward  
424 supported a stable CH<sub>4</sub>-EC of  $40.5 \pm 1.4 \text{ g CH}_4 \text{ m}^{-3} \text{ h}^{-1}$  over 15 cycles. Remarkably, the  
425 N starvation periods (24 h) did not entail a significant decrease in the system  
426 performance as previously reported by García-Pérez et al. (2018). These authors  
427 observed a deterioration in the EC from the fifth cycle onwards when longer N  
428 deprivation periods were applied (48 h) during the treatment of diluted CH<sub>4</sub> emissions

429 in a BCB. Interestingly, an increase in the mineralization ratio (from  $2.1 \pm 0.1$  to  $2.3 \pm$   
430  $0.1 \text{ g CO}_2 \text{ g}^{-1} \text{ CH}_4$ ) was recorded during the N-deprivation periods (**Fig 5a**), which  
431 suggested a higher carbon flux towards formaldehyde oxidation to  $\text{CO}_2$  for the  
432 regeneration of reducing equivalents needed in the PHB pathway (Khosravi-Darani et  
433 al., 2013).

434 The implementation of feast-famine N cycles was initiated after a first limitation  
435 period lasting 48 h, which induced a PHB accumulation up to  $23.2 \pm 0.3\%$  (mg PHB  
436  $\text{mg}^{-1} \text{ TSS} \times 100$ ) in the cells. **Fig 5b** illustrates the dynamics of N addition and rapid N  
437 uptake by *M. hirsuta* during the 15 cycles. The implementation of 15 N feast-famine  
438 cycles supported an average PHB accumulation of  $14.5 \pm 2.9\%$  (**Fig. 5c**). The  
439 determination of the partitioning coefficients revealed that most of the electrons derived  
440 from methane were used for energy production by the cells ( $f_e = 0.80$ ) during the  
441 accumulation phase (**Table S1**). Although values ranging 0.52-0.94 were reported by  
442 López et al. (2018) when cultures were supplemented with volatile fatty acids, the  $f_e$   
443 found in batch assays were typically lower ( $\sim 0.32$ ). During each cycle, the presence of  
444 N in the cultivation broth triggered the co-consumption of PHB and  $\text{CH}_4$ , which  
445 entailed a decrease of  $\sim 5\%$  in PHB accumulation (up to  $9.1 \pm 3.5\%$ ).

446 < **Figure 5** >

447 According to Bordel et al. (2019c), the depletion of the stored PHB in the  
448 presence of both  $\text{CH}_4$  and nitrogen occurs through anaplerotic reactions. These reactions  
449 provide intermediates that are necessary for the synthesis of building blocks such as  
450 glyoxylate and succinyl-CoA into the serine and TCA cycle, respectively. In this  
451 context, a previous work demonstrated that the consumption of the accumulated PHB  
452 by *Methylocystis parvus* did not support growth in the absence of  $\text{CH}_4$  (Pieja et al.,

453 2011b). In fact, it has been reported that PHB storage in the presence of CH<sub>4</sub> could  
454 provide bacteria a competitive advantage.

455 Repeated N cycles of 24-h in a sequencing batch reactor resulted in a similar  
456 PHB content in a methanotrophic mixed culture (~15%) (Pieja et al., 2012). Although  
457 the figures recorded were nearly a third than those achieved batchwise by *M. hirsuta*  
458 using biogas as CH<sub>4</sub> source (up to 45%) (López et al., 2018; Rodríguez et al., 2020), no  
459 previous study has been carried out under strictly continuous operation mode aiming at  
460 biogas valorization. It must be emphasized that the carbon mass balance conducted  
461 presented errors of 0.4 and 5.4% during the growth and accumulation phase,  
462 respectively, which validated the results and analyses carried out (Table S1).

463 PHB productivities ranging from 0.04 to 0.06 kg PHB m<sup>-3</sup> d<sup>-1</sup>, corresponding to non-  
464 N limited and N limiting conditions, respectively, were obtained. An estimation based  
465 on the global stoichiometry of the process and the CH<sub>4</sub> uptake rate (Eq. 6) led to a  
466 theoretical productivity value of ~0.26 kg PHB m<sup>-3</sup> d<sup>-1</sup>. The productivities herein  
467 recorded remained below this value likely due to the substantial impact observed from  
468 PHB consumption during the growth phase on the overall yield of the process. In this  
469 regard, PHB consumption lowered this value nearly by 60%, i.e. from 0.12 (expected)  
470 to 0.05 g PHB produced g<sup>-1</sup> CH<sub>4</sub> (Table S1). Thus, PHB depletion would result in a  
471 PHB productivity of 0.10 kg PHB m<sup>-3</sup> h<sup>-1</sup>, which would match satisfactorily with the  
472 experimental results. In addition, productivities slightly below the theoretical figures  
473 could be also explained by the short time available for methanotrophs to accumulate  
474 PHB during the N deprived period applied. A previous study using a similar gas-liquid  
475 contactor in batch mode and natural gas as a CH<sub>4</sub> source found that the maximum PHB  
476 accumulation (30.5%) occurred over 84 h (Rahnama et al., 2012). In this context, a  
477 strategy with an extended N limiting period was carried out (24h with N:48 without N).

478 However, process operation with such long N limitation resulted in an EC and PHB  
479 content decrease after the first complete cycle (data not shown), which ultimately  
480 resulted in system collapse.

481 In a biorefinery context, in which a medium size municipal solid waste (MSW) plant  
482 treats over 300 ton d<sup>-1</sup> of residues with an organic fraction of 46% (IEA Bioenergy), 1  
483 ton of VS typically would yield 121.7 m<sup>3</sup> CH<sub>4</sub>. Thus, considering repeated N cycles of  
484 24h:24h, a removal efficiency of 70% and a PHB yield of 0.54 g PHB g<sup>-1</sup> CH<sub>4</sub>, it can be  
485 predicted that 72.1 ton of MSW would be necessary for the production of 1 ton of PHB.  
486 This is, 6.9 kg of PHB can be produced out of 1 ton of MSW. In this regard, a recent  
487 geographical analysis conducted by Pérez et al. (2020) revealed that a combined  
488 scenario, i.e. PHB and cogeneration from biogas, in countries in which energy costs are  
489 high, would achieve PHB production costs as low as 1.5 euro kg<sup>-1</sup>. It also came out that  
490 in those regions where energy production is not economically favorable, biogas could  
491 be fully exploited for PHB production with competitive production costs (4.1 euro kg<sup>-1</sup>).

492 Finally, it is also worth mentioning that methane content in the exhaust gas from the  
493 reactor was 3.0 ± 0.0 % v v<sup>-1</sup> as a result of the high dilution ratio when using air as O<sub>2</sub>  
494 source. Therefore, this CH<sub>4</sub> content would not match the minimum concentration  
495 required for CH<sub>4</sub> combustion, which is above 35-40% (Haubrichs and Widmann, 2006).

496

497

498

## 499 **4 Conclusions**

500 This work demonstrated for the first time the technical feasibility of PHB  
501 production from biogas in a continuous bubble column bioreactor equipped with  
502 internal gas recirculation. This work provided valuable insights into the operational  
503 conditions supporting a sustained CH<sub>4</sub> bioconversion into PHB. The implementation of  
504 internal gas recirculation led to the decoupling of the turbulence in the cultivation broth  
505 and gas EBRT, providing outstanding CH<sub>4</sub>-ECs and CH<sub>4</sub>-REs with values up to 4 times  
506 higher than in the absence of gas recirculation. An EBRT of 60 min and a R of 30 were  
507 identified as the optimal conditions for maximizing substrate utilization. N-NO<sub>3</sub><sup>-</sup> supply  
508 to the culture broth must match N demand when using biogas as a CH<sub>4</sub> source in order  
509 to prevent nitrite accumulation and the subsequent inhibition of methanotrophic activity.  
510 Finally, the N feast-famine strategy applied for PHB production (24h:24h) under  
511 optimal mass transfer conditions conferred a stable CH<sub>4</sub> oxidation (stage I) and a  
512 continuous PHB production (stage II) with a CH<sub>4</sub>-RE of 70% and PHB productivities up  
513 to 0.06 kg PHB m<sup>-3</sup> d<sup>-1</sup>. These findings highlight the potential of methanotrophic  
514 bacteria as an effective/feasible platform for PHB production within a biogas bio-  
515 refinery concept. Furthermore, the use of biogas as a low-cost and “green” alternative to  
516 conventional carbon sources for biopolymer production would boost their viability in  
517 terms of environmental and economic impact. Further studies should explore process  
518 robustness under N limiting conditions ranging from 24 to 48 h to achieve higher PHB  
519 accumulations.

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529

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693 **Tables**

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695 **Table 1.** Operational conditions during the CH<sub>4</sub> mass transfer optimization test.

Test No.	EBRT (min)	Inlet gas flow (ml min <sup>-1</sup> )	R	Virtual EBRT (min)	Inlet load (g CH <sub>4</sub> m <sup>-3</sup> h <sup>-1</sup> )	Organic loading rate (g COD L <sup>-1</sup> d <sup>-1</sup> )
1.1	30	83	0	30	202 ± 22	19.4 ± 2.1
1.2			2.5	9		
1.3			5	5		
1.4			10	3		
1.5			15	2		
1.6			30	1		
2.1	60	42	0	60	86 ± 6	8.3 ± 0.6
2.2			2.5	17		
2.3			5	10		
2.4			10	5		
2.5			15	4		
2.6			30	2		
3.1	120	21	0	120	51 ± 4	4.9 ± 0.4
3.2			2.5	34		
3.3			5	20		
3.4			10	11		
3.5			15	8		
3.6			30	4		

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700 **Figure captions**

701 **Fig. 1** (a) Image and (b) Schematic representation of the experimental set-up. (1) Air  
702 compressor, (2) Mixing chamber, (3) Rotameters, (4) Biogas cylinders, (5) Bubble  
703 column bioreactor, (6) Condenser, (7) Liquid sampling port, (8) Gas recirculation pump,  
704 (9) Thermostatic bath, (10) Gas sampling ports, (11) Magnetic stirrer, (12) Mass flow  
705 controller.

706 **Fig. 2** Influence of R on the (a) CH<sub>4</sub>-EC and on the (b) CH<sub>4</sub>-RE at EBRTs of 30 min, 60  
707 min and 120 min.

708 **Fig. 3** Influence of the virtual gas residence time on the CH<sub>4</sub>-EC.

709 **Fig. 4** Time course of (a) CH<sub>4</sub>-EC and PCO<sub>2</sub>; (b) N-NO<sub>2</sub><sup>-</sup> and N-NO<sub>3</sub><sup>-</sup> concentration in  
710 the culture broth, and N-NO<sub>3</sub><sup>-</sup> concentration in the mineral salt medium; (c) biomass  
711 concentration expressed as TSS.

712 **Fig. 5** Time course of (a) CH<sub>4</sub>-EC and PCO<sub>2</sub>; (b) N-NO<sub>2</sub><sup>-</sup> and N-NO<sub>3</sub><sup>-</sup> concentration in  
713 the culture broth, and N-NO<sub>3</sub><sup>-</sup> concentration in the mineral salt medium; (c) PHB (%)  
714 and biomass concentration expressed as TSS (hexagons).

**“Biogas valorization via continuous polyhydroxybutyrate production  
by *Methylocystis hirsuta* in a bubble column bioreactor”**

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Ceará, Brazil

## Supplementary Material

### Material and methods

#### S1. Electric power demand

The power demand of the internal gas recirculation and air compressors for each operational condition at the mass transfer test (2.2.2) was estimated according to the following formula (Estrada et al. 2011):

$$P \text{ (kW)} = \frac{\Delta P \cdot (Q + Q_R)}{\eta} \quad (\text{Eq.S1})$$

where  $\Delta P$  represents the pressure drop (kPa),  $Q + Q_R$  represent the real flow entering the column (inlet gas flow and internal recirculation flow, respectively) and  $\eta$  stands for the efficiency of both compressors (70%).

#### S2. Carbon footprint emissions

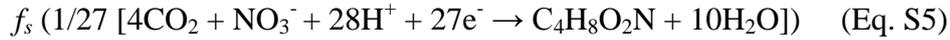
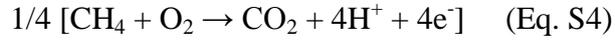
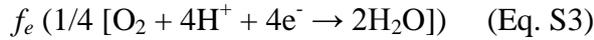
Two scenarios were evaluated to study the environmental impact in terms of CO<sub>2</sub> equivalents (kg CO<sub>2</sub> y<sup>-1</sup>) of direct and indirect emissions on a 1-year basis: (1) The biogas produced in the anaerobic digester was completely vented; (2) The biogas produced in the anaerobic digester was treated in the BCB. Both scenarios were assessed at the different operating conditions of the mass transfer test.

Conversion factors for methane emission and electricity production of 25 kg CO<sub>2</sub> kg<sup>-1</sup> CH<sub>4</sub> and 0.35 kg CO<sub>2</sub> kWh<sup>-1</sup> were used, respectively.

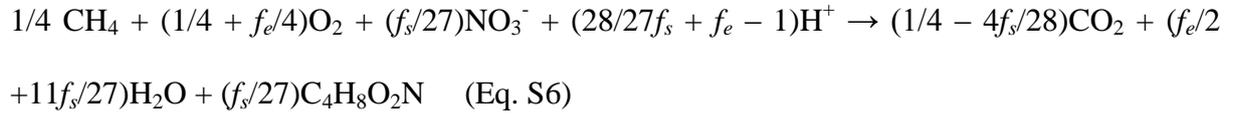
$$\text{Carbon footprint reduction (\%)} = \frac{CO_2 \text{ eq}_{(1)} - CO_2 \text{ eq}_{(2)}}{CO_2 \text{ eq}_{(1)}} \quad (\text{Eq.S2})$$

### S3. Substrate partitioning coefficients ( $f_e$ and $f_s$ ) and carbon distribution

As described by Rostkowski et al. (2012), cell synthesis in methanotrophs occurs in three half reactions: (1) the reduction of the electron acceptor ( $O_2$ ) into  $H_2O$  (Eq. S3); (2) the oxidation of the electron donor ( $CH_4$ ) into  $CO_2$  (Eq. S4) and (3) cell growth. The latter has been adjusted according to the formula used for cell mass ( $C_4H_8O_2N$ ) (Khosravi-Darani et al., 2013) in the present work.

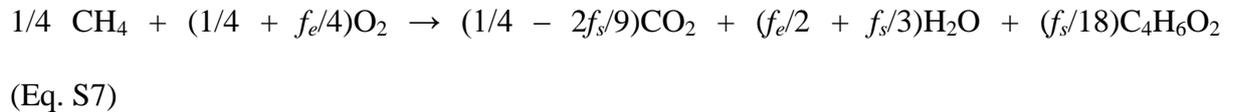


The global reaction for cell mass synthesis using  $NO_3^-$  as a nitrogen source is given by Eq. S6.

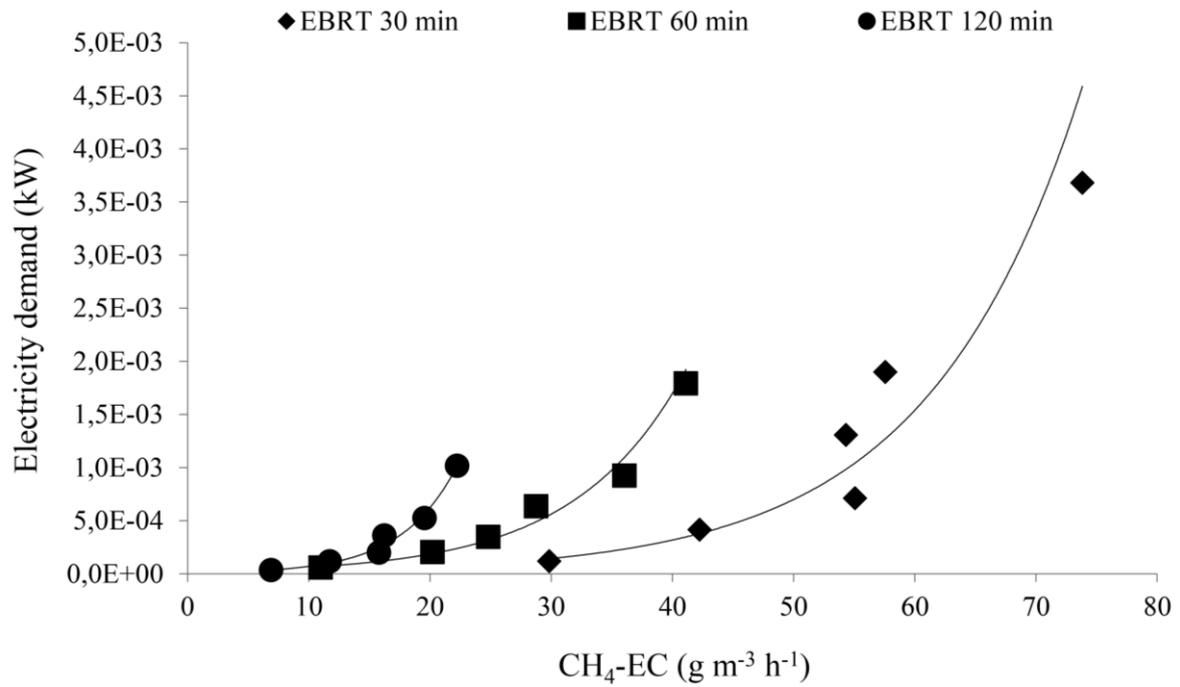


$f_e$  and  $f_s$  represent the fraction of electrons from the substrate that are utilized for energy generation and for biomass synthesis, respectively. The sum of  $f_e$  and  $f_s$  is equal to 1.

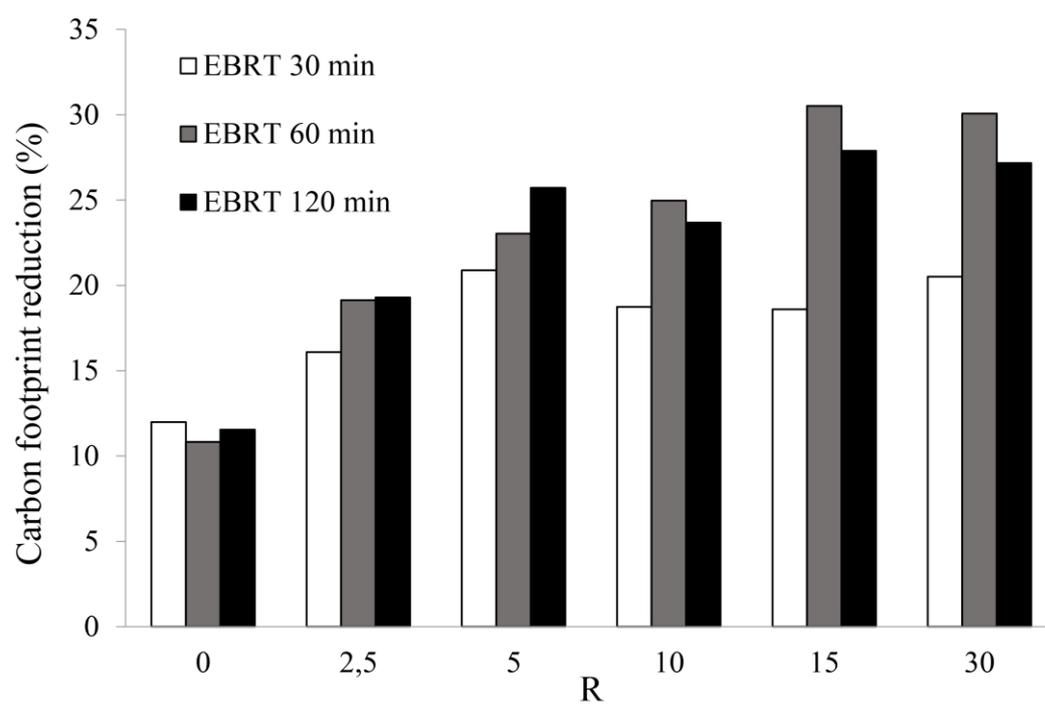
Likewise, biopolymer ( $C_4H_6O_2$ ) synthesis can be expressed as:



## Results and discussion



**Figure S1.** Power demand at different EBRTs. Each point of the curve represents the different internal recirculation rates assayed (from left to right: 0, 2.5, 5, 10, 15 and 30).



**Fig. S2** Carbon footprint mitigation (%) resulting from the biological treatment at lab scale of biogas (comparison between Scenario 1 and 2).

**Table S1.** Electron fractions ( $f_e$  and  $f_s$ ) and carbon distribution during a N feast-famine cycle (24h:24h)

Phase	$f_e$	$f_s$	$Y_X$ (g biomass g <sup>-1</sup> CH <sub>4</sub> ) $Y_{PHB}$ (g PHB g <sup>-1</sup> CH <sub>4</sub> )	Carbon balances (C-g)							
				CH <sub>4</sub> (in)	CO <sub>2</sub> (in)	CH <sub>4</sub> (out)	CO <sub>2</sub> (out)	Biomass (out)	PHB (out)	TOC (out)	Balance error (%)
Growth	0.44	0.56	0.55	2.70 ± 0.11	1.07 ± 0.03	0.89 ± 0.03	2.41 ± 0.11	0.64 ± 0.06	-0.19 ± 0.02	0.04 ± 0.00	0.45
Accumulation	0.80	0.20	0.24	2.73 ± 0.07	1.05 ± 0.04	0.90 ± 0.02	2.55 ± 0.13	0.21 ± 0.01	0.32 ± 0.04	0.00 ± 0.00	5.38

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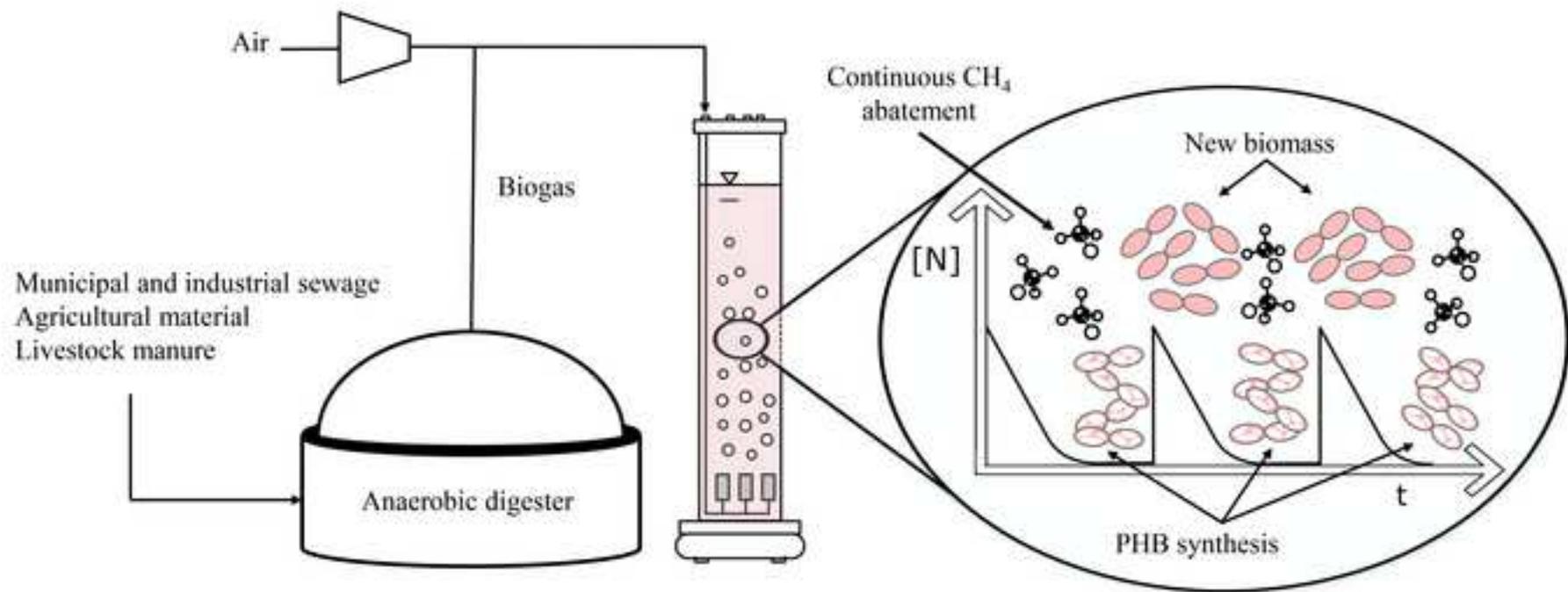


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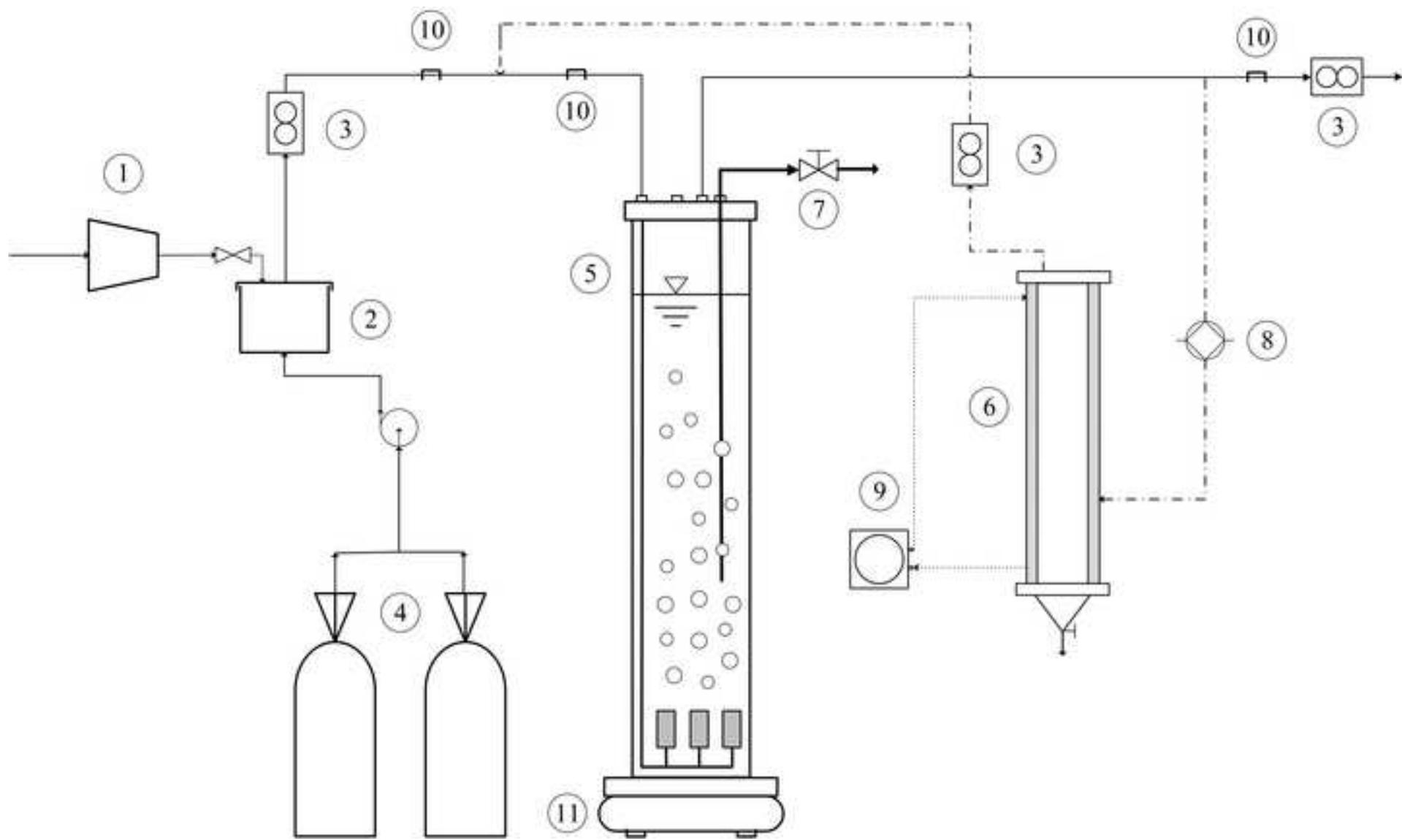


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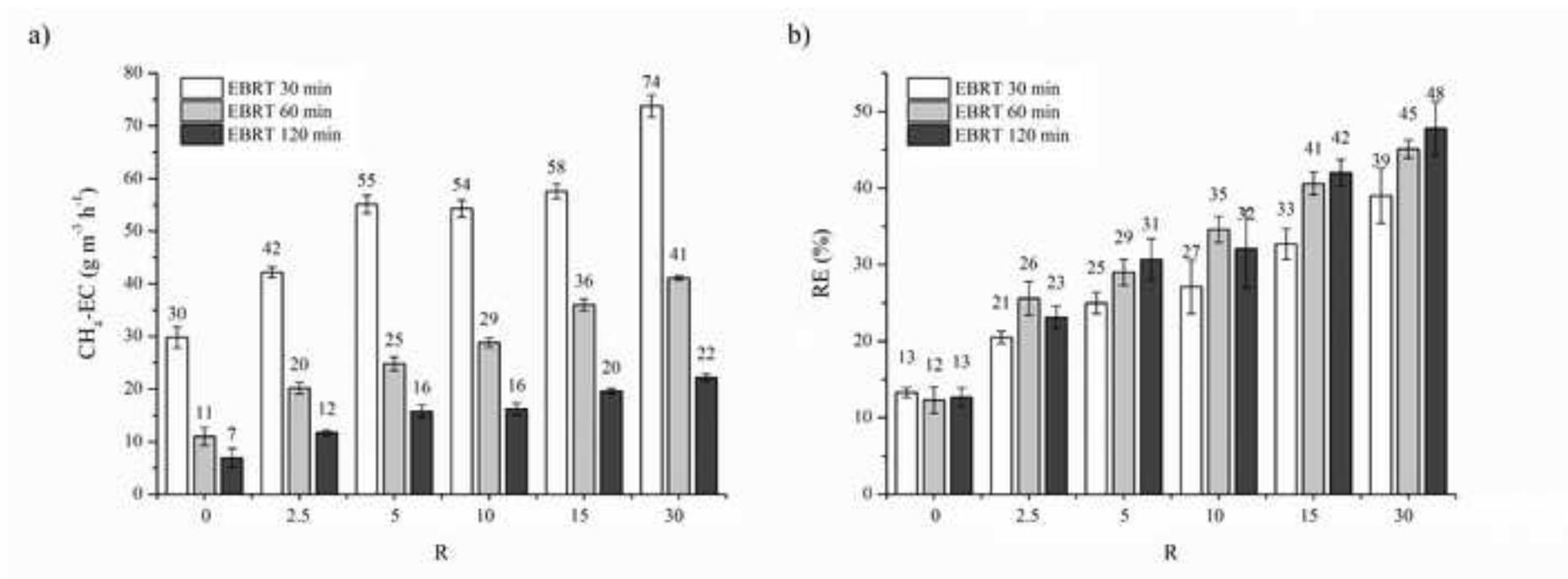


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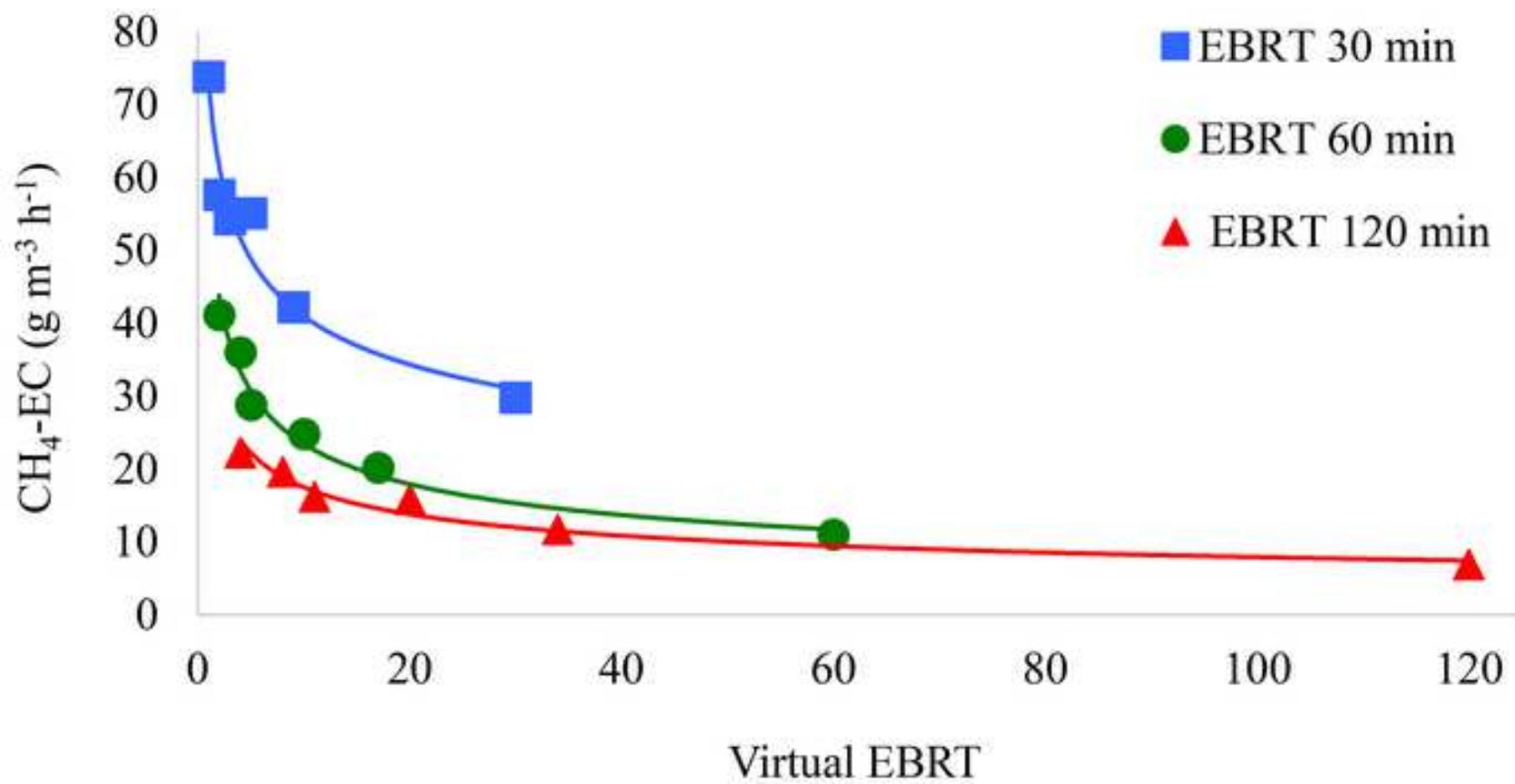


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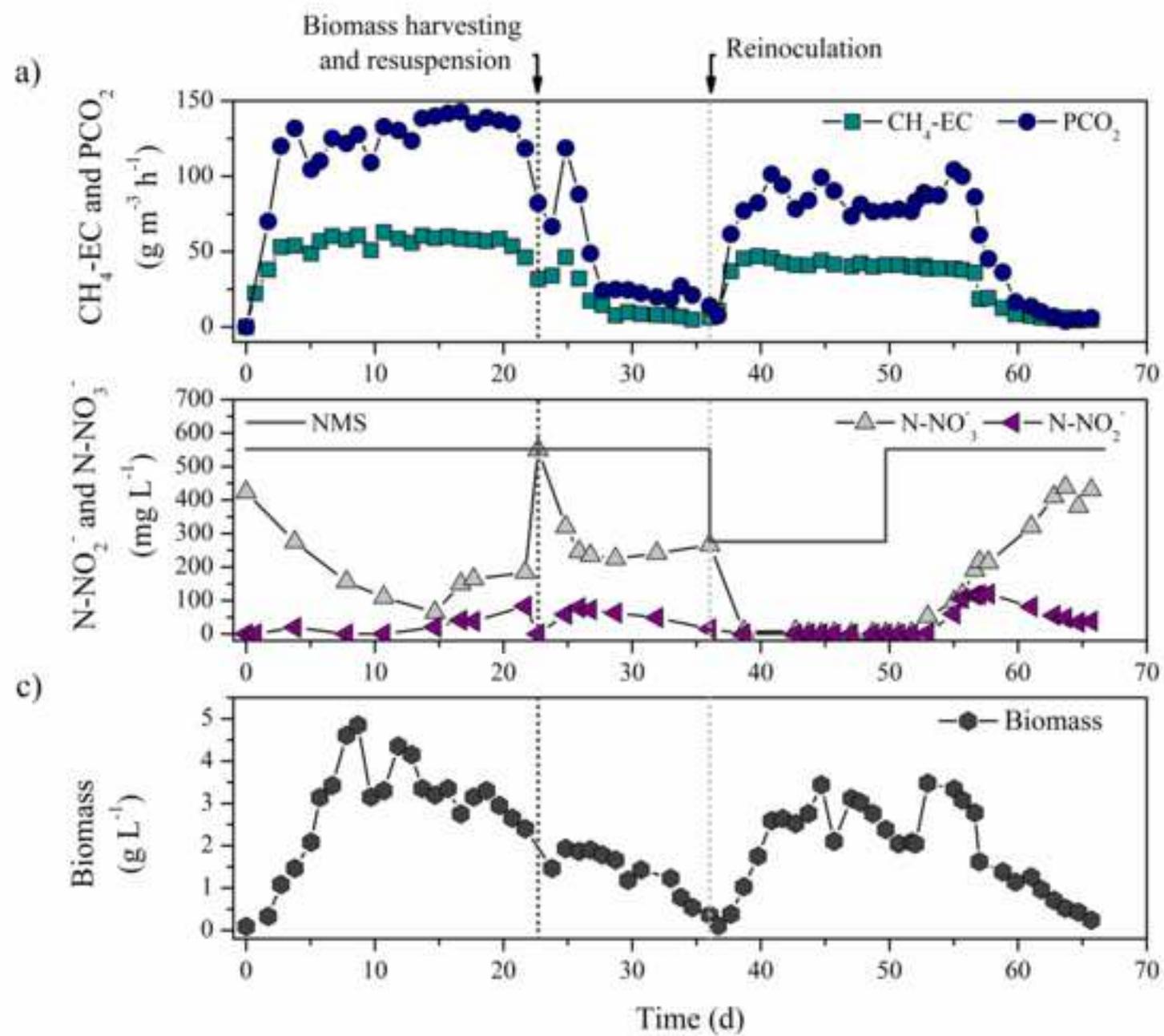


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