

Relationships between Water Status, Leaf Chlorophyll Content and Photosynthetic Performance in Tempranillo Vineyards

G. Hailemichael¹, A. Catalina², M.R. González², P. Martín^{2*}

(1) Jimma Agricultural Research Centre, P.O. Box 192, Jimma, Ethiopia

(2) Departamento de Producción Vegetal y Recursos Forestales, Universidad de Valladolid, Avda. de Madrid 57, 34004 Palencia, Spain

Submitted for publication: April 2016

Accepted for publication: July 2016

Key words: chlorophyll fluorescence, chlorosis, photosynthesis, stress, *Vitis vinifera* L.

The aims of this work were (i) to study the relationships between leaf water potential, foliar chlorophyll content and photosynthetic performance of grapevines under field conditions, and (ii) to evaluate the feasibility of chlorophyll fluorescence parameters to assess water status in commercial vineyards. During three consecutive seasons (2011 to 2013), 20 non-irrigated “Tempranillo” vineyard subzones were monitored in Ribera del Duero, Spain. Data on foliar chlorophyll content, midday and predawn leaf water potential, net assimilation and chlorophyll fluorescence parameters were measured at fruit set, pre-véraison and véraison. The results show that the water potential indicated weak to moderate stress in the study years and decreased along the vegetative cycle, while foliar chlorophyll contents showed an increasing trend. The correlation between both variables was not consistent. Linear regression of the chlorophyll fluorescence parameters in dark-adapted leaves, F_o , F_v and F_v/F_m , over chlorophyll content was strong in all the study years (higher determination coefficients for F_v/F_m at fruit set in 2012 and 2013). In general, the water potential with low coefficients of variation showed a low effect on photosynthesis activity, although values in variables such as F_m , F_v , F_v/F_m , F_s/F_o and Φ_{PSII} registered an increasing trend when midday leaf water potential decreased in the pre-véraison and véraison stages. These variables could be useful to estimate water status in commercial vineyards for application in precision viticulture.

INTRODUCTION

Conditions such as extreme air temperature, changes in photoperiod, light intensity and quality, nutrient abundance and starvation, drought, flooding or excessive salts hinder the full expression of the genetic potential of plants and are limiting to yield and fruit quality (Ciais *et al.*, 2005). There is a lot to research on the relationships between soil water availability to the grapevine, water loss through evapotranspiration, grapevine photosynthesis, and yield and grape composition (Medrano *et al.*, 2003; Van Leeuwen *et al.*, 2009; Alves *et al.*, 2013). In temperate climates, water deficit conditions are necessary to produce high-quality red wine (Van Leeuwen *et al.*, 2009), but stress as a result of excessive water deficit may lead to significant productivity and quality losses (Ojeda *et al.*, 2002). Vine water status is dependent on soil and climate characteristics, and its spatial characterisation in vineyards is an important step in precision viticulture.

Drought limits photosynthesis, even in plants such as grapevines that are well adapted to this condition (Chaves,

1991; Adir *et al.*, 2003). Under conditions of mild water stress, stomatal conductance (g_s), internal CO_2 concentration and/or net assimilation (A_n) decline and the A_n/g_s ratio increases (Flexas *et al.*, 2000; Van Leeuwen *et al.*, 2009). When water stress is more severe, non-stomatal regulation of photosynthesis is observed, including decreased light- and CO_2 -saturated photosynthesis (Escalona *et al.*, 1999), reduced photochemistry (Flexas *et al.*, 1999; 2002a), depressed activity of photosynthetic enzymes (Maroco *et al.*, 2002) and decreased mesophyll conductance (Flexas *et al.*, 2002a).

Photosynthetic pigments allow plants to absorb energy from light, so foliar chlorophyll content is a key factor affecting the performance of plant photosynthesis (Taiz & Zeiger, 2006). Various reports have explained that drought stress significantly decreases the chlorophyll a, chlorophyll b and total chlorophyll content of different crops (Mafakheri *et al.*, 2010; Gholamin & Khatnezhad, 2011). However, the presence of low levels of chlorophyll in leaves (chlorosis)

*Corresponding author: pmartinp@pvs.uva.es

Acknowledgements: The authors acknowledge the financial support of the Spanish Ministry of Science and Education (MEC) for project RTA2014-00077-C02-02, and of the Regional Government of Castilla y Leon for project VA011A10-2. The authors are grateful to Bodegas Emilio Moro S.L., for their collaboration in conducting this research

is a general symptom of stress and might be poorly related to water status in the field when other environmental factors interact.

The analysis of chlorophyll fluorescence parameters is considered as an important approach to evaluating the health or integrity of the photosynthetic apparatus within a leaf, and provides a rapid and accurate technique for detecting and quantifying the interaction among and tolerance of plants to stress (Maxwell & Johnson, 2000; Lichtenthaler *et al.*, 2005). The intensity of water stress in vines has been significantly correlated with non-photochemical quenching and minimal, maximum, variable and steady-state fluorescence in light-adapted leaves (Souza *et al.*, 2004; Catalina *et al.*, 2011). The ratio of steady-state to minimal fluorescence in dark-adapted leaves (F_s/F_0) has also been reported to be useful in the early detection of water stress (Flexas *et al.*, 2002b). On the other hand, the fluorescence variables measured in dark-adapted leaves may be strongly affected by other factors, such as iron deficiency chlorosis (Bavaresco *et al.*, 2006; Martín *et al.*, 2007; Catalina *et al.*, 2011; Zarco-Tejada *et al.*, 2013).

Most of the studies that have linked photosynthesis and water status in grapevines, mentioned above, were carried out in single irrigation experiments, applying different water regimes to vines under controlled conditions. It was regarded as interesting for precision viticulture to contrast the impact of water status on photosynthetic activity in commercial vineyards, where variables such as soil characteristics or cultivation practices may modify its effects. The objectives of this work therefore were (i) to study the relationships between water status, foliar chlorophyll content and photosynthetic performance of grapevines under field conditions, and (ii) to evaluate the feasibility of chlorophyll fluorescence parameters to assess water status in vineyards for application in precision viticulture.

MATERIALS AND METHODS

Description of study site

The study was conducted in 2011, 2012 and 2013 on 24 non-irrigated vineyard subzones in Pesquera de Duero, Valladolid (Ribera del Duero Designation of Origin, North-Central Spain), at an altitude of 800 m above sea level. The subzones (10 m x 10 m each) were selected with different soil depths, soil texture and topography to assure maximum variability in water availability across the sites, in terms of the purposes of the study. The vineyards were planted with the 'Tempranillo' cultivar, were 10 to 14 years old and grafted on 110-Richter rootstock. Vines were spaced at 3 m x 1.5 m (2 222 plants per hectare), pruned on a double cordon Royat system, trellis trained and with vertically positioned shoots.

The soils in the study area are calcareous, very basic and poor in organic matter (Martín *et al.*, 2008). Texture ranged from medium to medium-weighted. Concentrations of active carbonate (3.3 to 15.5%) and DPTA extractable iron (2.3 to 6.4 mg/kg) were highly heterogeneous within the area. These soil properties, along with the presence of a lime-sensitive rootstock (110-Richter), led to different levels of iron deficiency chlorosis in the vineyards of the area. The subzones had a high variability of soil concentrations of extractable potassium, phosphorus and magnesium (Zarco-Tejada *et al.*, 2013).

The study area has a Mediterranean climate, with low temperatures in the winter and hot and dry summers. Mean annual temperatures were 11.2°C, 11.4°C and 11.1°C in 2011, 2012 and 2013 respectively. The rainfall registered from 1 April to 30 September was 137 mm, 185 mm and 173 mm in the first, second and third year respectively (Table 1). Without irrigation, this insufficient water supply in soils varying in texture and root explorability depth ensured a broad variability in soil water content in the study zones.

TABLE 1

Monthly mean values of maximum (T_{max}), minimum (T_{min}) and average (T_{ave}) temperature (°C), and total precipitation (P, in mm), collected during the study years from the station VA07 - Valbuena de Duero (Valladolid, Spain).

Year	Variables	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sept	Oct	Nov	Dec
2011	T_{max}	7.6	11.9	13.4	20.6	24.0	26.8	28.3	30.0	28.1	22.2	12.9	9.0
	T_{min}	1.0	-1.1	2.3	6.2	7.3	9.4	9.9	12.4	9.0	3.5	9.0	-0.8
	T_{ave}	4.1	4.6	7.4	13.1	15.7	18.2	19.6	21.2	18.4	12.6	8.1	3.5
	P	38.9	19.2	34.7	31.6	18.3	7.2	3.0	57.2	20.0	18.8	41.8	15.2
2012	T_{max}	7.8	9.2	16.6	13.0	23.1	28.1	29.8	30.8	25.2	18.5	12.2	9.1
	T_{min}	-2.1	-4.3	-0.3	2.5	6.4	9.9	10.2	11.4	9.6	4.7	2.1	0.7
	T_{ave}	2.1	2.1	8.2	7.7	15.0	19.3	20.5	21.5	17.2	11.3	6.9	4.8
	P	1.6	0.0	20.8	95.0	41.4	18.0	6.0	1.4	22.9	24.1	47.5	32.6
2013	T_{max}	8.7	9.2	11.8	15.1	17.3	24.5	32.3	30.2	26.1	19.1	11.3	8.9
	T_{min}	0.1	-1.4	2.2	2.6	3.7	8.2	13.1	11.6	9.6	7.4	1.9	-2.4
	T_{ave}	4.1	3.4	6.6	8.9	10.4	16.2	22.7	20.9	17.6	12.8	6.5	2.4
	P	50.7	36.5	137.4	47.7	36.9	22.3	7.7	4.8	53.5	69.8	10.3	55.2

Field data collection

Data on foliar chlorophyll content, midday and predawn leaf water potential, net assimilation and chlorophyll fluorescence variables were measured in the fruit set, pré-véraison (approximately 20 to 28 days after fruit set) and véraison growth stages.

Foliar chlorophyll content was recorded with a CL-01 portable colorimeter (Hansatech Instruments Ltd., Norfolk, UK). In each subzone, 30 leaves were selected by choosing the fourth or fifth leaf, counting from the first sheet of the apex. Chlorophyll content (Chl, in $\mu\text{g}/\text{cm}^2$ leaf) was calculated from the CL-01 meter readings (R) by the regression line obtained for cv. Tempranillo by González *et al.* (2005), as $\text{Chl} = 6.0817 \cdot R + 7.6084$.

Net assimilation (A_n : $\mu\text{mol CO}_2/\text{m}^2/\text{s}$) and chlorophyll fluorescence parameters were monitored using a LI-Cor 6400 portable infrared gas analyser (IRGA) equipped with a 6400-40 leaf chamber pulse amplitude modulation fluorometer (Li-Cor, Inc. Lincoln, Nebr., USA). Fluorescence parameters included minimum (F_0) and maximum (F_m) fluorescence in dark-adapted leaves (F_0' and F_m' under light conditions), variable (F_v) and steady-state fluorescence (F_s), efficiency (ΦPSII) and maximum efficiency (F_v/F_m and F_v'/F_m') of photosystem II, apparent rate of electron transport (ETR), and photochemical (qP) and non-photochemical quenching (NPQ).

Nine fully expanded leaves exposed to direct solar radiation and located at the fourth or fifth node from the apex were selected in each subzone to conduct the photosynthesis measurements. The chlorophyll fluorescence parameters of the dark-adapted leaves were obtained before dawn (01:00 to 03:00 local time), while the levels of gas exchange and fluorescence parameters of the light-adapted leaves were obtained at midday (11:00 to 13:00 local time). Leaf chamber conditions were set constant for all measurements conducted: incident photosynthetic photon flux density on the leaves was set to $1\,500\ \mu\text{mol photons}/\text{m}^2/\text{s}$ photosynthetic saturation in field-grown grapevines (Escalona *et al.*, 1999); and CO_2 levels in the analyser were held constant at $370\ \mu\text{mol CO}_2/\text{mol air}$. Temperature, air pressure and relative humidity were maintained at ambient levels. Air flow rate through the leaf chamber was maintained at $500\ \mu\text{mol}/\text{s}$. Leaf

absorbances measured at 635 nm and 465 nm were entered into the IRGA for photosynthetic measurements taken at each study site. These values were estimated from leaf greenness data, previously obtained with a CL-01 portable colorimeter as described above (González *et al.*, 2005).

Measurements of predawn leaf water potential (Ψ_{pd}) were taken during the two hours before dawn. The vine is in equilibrium with the water potential of the soil at that time (Winkel & Rambal, 1993). Measurements of midday leaf water potential (Ψ_{Md}) were performed within two hours of solar noon (13:00 to 15:00 local time). The leaf water potential was monitored with a Scholander pressure chamber (Solfranc Technologies SL, Spain). Values were measured in six exposed leaves in each subzone, on the same shoot/branch that was utilised for the photosynthesis and chlorophyll fluorescence measurements.

Statistical analysis

The relationships of the leaf water potential, foliar chlorophyll content and photosynthesis variables were tested with linear regression analysis. Analysis of variance (ANOVA) and Tukey's test were performed to test mean differences between years. Data analysis was performed with version 9.2 of the SAS software package (SAS Institute Inc., Cary NC, USA).

RESULTS AND DISCUSSION

Water status and foliar chlorophyll content

Table 2 shows the variation in the ranges of leaf water potential in the study zones in each phenological stage and year studied. It is interesting to note that predawn leaf water potential (Ψ_{pd}) always registered higher variation coefficients than midday leaf water potential (Ψ_{Md}). The variability of water status between subzones was clearly greater in 2011 than in 2012 and 2013 (Table 2), when the precipitation registered during the vegetative cycle was lower (Table 1).

In general, the records of midday and predawn leaf water potential at fruit set and véraison in the study years indicated moderate to weak water deficit, according to the ranges from Van Leeuwen *et al.* (2009), although in some stages, such as pré-véraison in 2013 (Table 2), the Ψ_{Md} values correspond to moderate to severe water deficit ranges. Normally, values

TABLE 2

Minimum, maximum and coefficient of variation (CV) of predawn and midday leaf water potential (Ψ_{pd} and Ψ_{Md} respectively, in MPa) registered in the years studied.

Year	Value	Fruit set		Pre-véraison		Véraison	
		Ψ_{Md}	Ψ_{pd}	Ψ_{Md}	Ψ_{pd}	Ψ_{Md}	Ψ_{pd}
2011	Min.	-0.521	-0.200	-0.673	-0.392	-0.809	-0.371
	Max.	-0.820	-0.534	-1.051	-0.740	-1.436	-0.941
	CV (%)	9.84	22.81	11.43	16.71	11.75	28.99
2012	Min.	-0.708	-0.423	-0.963	-0.566	-1.101	-0.619
	Max.	-0.950	-0.731	-1.404	-0.991	-1.515	-1.107
	CV (%)	8.24	13.78	10.38	13.85	7.98	13.19
2013	Min.	-0.716	-0.466	-1.022	-0.689	-0.879	-0.680
	Max.	-1.238	-0.825	-1.532	-0.979	-1.192	-1.073
	CV (%)	13.18	16.45	7.25	8.34	7.96	8.93

of leaf water potential decrease through the vegetative cycle (Table 2). Ψ_{Md} and Ψ_{Pd} decreased as the precipitation became very scanty, the soil water reserves dropped and the temperature increased from spring to summer (Medrano *et al.*, 2003).

The ANOVA of leaf water potential and foliar chlorophyll content (Chl) at fruit set and véraison was significant according to year from the explanatory variables (data not shown), evidence of the existence of a high variability in meteorological conditions in the study years. From a comparison of the means, the values of both Ψ_{Md} and Ψ_{Pd} at fruit set decreased from 2011 to 2013 (Table 3), while more negative values were registered at véraison in 2012.

The mean values of Chl at fruit set in 2012 were higher than in 2013, but at véraison the result was reversed (Table 3). Lower levels of leaf water potential (more water-stressed vines) corresponded to lower values of Chl at fruit set and véraison in 2011; however, this relationship was not repeated in other years. These results gave a hint that water status and foliar chlorophyll content appear to be independent. In fact, no consistent correlations were obtained between these variables at different growth stages and in different years of study (Table 4). Negative correlations were observed at fruit set, pre-véraison and véraison in 2011, and at véraison in 2012, but the correlation coefficient of Chl with Ψ_{Pd} at fruit set in 2013 was positive.

Foliar chlorophyll content, which is part and parcel of the vigour of plants, is influenced by soil and environmental factors. Various reports have explained that drought stress significantly decreases the chlorophyll content in crops such as chickpeas (Mafakheri *et al.*, 2010) or maize (Gholamin & Khatnezhad, 2011). In the present study, Chl was not a good indicator of the water status of grapevines. This might be due to both the existence of narrow ranges of variation in leaf water potential (slight to moderate stress), and the

influence of nutrient status (iron deficiency chlorosis), which could strongly affect the foliar chlorophyll content in the study area (Martín *et al.*, 2008).

Chlorophyll content and photosynthesis activity

Foliar chlorophyll content per unit area of leaf is considered to be an indicator of the photosynthetic capacity of plants (Taiz & Zeiger, 2006; Ling *et al.*, 2011). In the present study, the linear regression of An over Chl was significant and had a positive slope at the pre-véraison stage in 2011 (Table 5), but no significant regressions were obtained in any of the other years and at any of the other measuring times.

As Table 5 shows, the linear regressions of chlorophyll fluorescence parameters in dark-adapted leaves, Fo, Fv and Fv/Fm, over Chl were significant in all the study years (higher determination coefficients for Fv/Fm at fruit set in 2012 and 2013). The positive relationship between Fv/Fm and Chl confirms the results of Palliotti *et al.* (2009), who showed that high levels of leaf photosynthetic pigments might enhance light absorption, thus increasing the maximum quantum yield of photosystem II. In relation to this, Mandal *et al.* (2009) observed that Fv/Fm was higher in healthy leaves than in plants infected by downy mildew (with chlorosis).

Fo measured at fruit set resulted in a significant and negative regression in all study years, while the relationship between both variables was not significant at more advanced phenological stages (Table 5). Significant and positive linear regressions were obtained between Fs/Fo and Chl in all vegetative stages in 2011, and at fruit set in 2013. The regressions of qP, NPQ and ETR over Chl was not statistically significant in most years and growth stages. Fluorescence variables in light-adapted leaves, Fo', Fm', Fs and Fv'/Fm', showed significant positive regression over Chl only at pre-véraison and véraison in 2011, and at pre-véraison in 2013.

TABLE 3

Comparison of the means of foliar chlorophyll content (Chl: $\mu\text{g}\cdot\text{cm}^{-2}$) and predawn and midday leaf water potential (Ψ_{Pd} and Ψ_{Md} respectively, in MPa), measured at fruit set and véraison.

Year	Fruit set			Véraison		
	Ψ_{Md}	Ψ_{Pd}	Chl	Ψ_{Md}	Ψ_{Pd}	Chl
2011	-0.708a	-0.375a	42.6c	-1.215b	-0.600a	87.5b
2012	-0.826b	-0.577b	82.9a	-1.321c	-0.895b	95.8ab
2013	-0.960c	-0.621b	63.4b	-1.017a	-0.879b	105.8a

Means followed by the same letter are not significantly different ($p < 0.05$) according to Tukey's test

TABLE 4

Pearson correlation coefficients between foliar chlorophyll content and predawn and midday leaf water potential (Ψ_{Pd} and Ψ_{Md} respectively) at fruit set and véraison.

Year	Fruit set		Pre-véraison		Véraison	
	Ψ_{Md}	Ψ_{Pd}	Ψ_{Md}	Ψ_{Pd}	Ψ_{Md}	Ψ_{Pd}
2011	-0.41*	-0.17	-0.24	-0.52**	-0.43**	-0.05
2012	-0.22	0.11	0.08	0.09	-0.50**	0.35
2013	0.04	0.48*	0.22	-0.20	0.06	0.07

Levels of significance: * $p < 0.05$, ** $P < 0.01$; $n = 24$

TABLE 5

R-squared and slope (in brackets) of significant linear regressions of net assimilation (An) and fluorescence variables on foliar chlorophyll content at different growth stages in 2011, 2012 and 2013.

Variables	2011			2012			2013		
	Fruit Set	Pre-véraison	Véraison	Fruit Set	Pre-véraison	Véraison	Fruit Set	Pre-véraison	Véraison
An		(+)0.40**							
Fo	(-)0.40*	(-)0.34**		(-)0.44*			(-)0.71***		
Fm		(+)0.38**	(+)0.47***	(+)0.46*	(+)0.23*	(+)0.42***	(+)0.22*	(+)0.32**	(+)0.26*
Fv	(+)0.33*	(+)0.51***	(+)0.48***	(+)0.62**	(+)0.28**	(+)0.48***	(+)0.60***		(+)0.29**
Fv/Fm	(+)0.41**	(+)0.59***	(+)0.31**	(+)0.68***	(+)0.41***	(+)0.24*	(+)0.72***		(+)0.23*
Fo'		(+)0.30**	(+)0.40**					(+)0.50***	
Fm'		(+)0.3**	(+)0.40**					(+)0.40***	
Fs		(+)0.30*	(+)0.30**					(+)0.40***	
Fv'/Fm'		(+)0.40**	(+)0.20*		(+)0.20*			(+)0.20*	
ΦPSII	(+)0.30*		(+)0.20*			(+)0.30**			
qP								(-)0.20*	
NPQ									
ETR	(+)0.30*		0.30*	0.20*		0.30*			
Fs/Fo	(+)0.75***	(+)0.56**	(+)0.29*				(+)0.56**		

Level of significance: * P < 0.05, ** P < 0.01 and *** P < 0.001, n = 24

The significant relationships observed between Fo, Fv/Fm and ΦPSII and Chl, repeated over three years, might be associated with the high variability in Chl generated within the study zone by the incidence of iron stress (Martín *et al.*, 2008). A low chlorophyll concentration (chlorosis) in young leaves is a visible symptom of iron deficiency, which has a significant negative effect on photosynthesis activity (Maxwell & Johnson, 2000). A shortage of physiologically active iron leads to a decrease in the electron transport rate, as well as to a lowering of the efficiency of photosystem II (Bavaresco *et al.*, 2006; Martín *et al.*, 2007).

Water status and photosynthetic activity

It is well known that the net assimilation by leaves decreases as water potential decreases (Chaves, 1991; Lawlor, 2002). Stomatal closure is one of the first responses of plants to the lack of water in the soil (Chaves, 1991; Flexas *et al.*, 2002a). After stomatal closure, water deficit induces a decline in photosynthetic activity by reducing the availability of CO₂ in the leaf mesophyll (Chaves, 1991; Flexas *et al.*, 2002a). In the present study, linear regression relations of An over ψ_{Md} and ψ_{pd} were not consistent (Table 6). However, the relationships between water potential and some chlorophyll fluorescence parameters reflect that the water status had a significant influence on photosynthetic performance, as explained below.

Minimum fluorescence (Fo) showed a significant and positive relationship with midday leaf water potential at fruit set in 2011, while no other significant relations were obtained in the rest of the phenological stages and years (Table 6). Other chlorophyll fluorescence variables measured in dark-adapted leaves, such as Fm, Fv and Fv/Fm, showed a significant and negative relationship with midday leaf water potential at fruit set and véraison in 2011 and at pre-véraison and véraison in 2012. On the other hand, relationships

between Fm, Fv and Fv/Fm and predawn leaf water potential were weak, except for values registered at véraison in 2013.

A sustained decrease in dark-adapted Fv/Fm and increase in Fo indicate the occurrence of damage to photosystem II, which reduces the efficiency of absorbed light energy transfer from the light-harvesting complex. For Maxwell and Johnson (2000), this may result from various environmental stresses, including high temperature or water deficit. Nevertheless, Baker and Rosenqvist (2004) have stated that water stress has no major impact on the efficiency of photosystem II. In this regard, Medrano *et al.* (2003) did not detect significant differences in Fv/Fm between irrigated and non-irrigated grapevines of cv. Tempranillo. Souza *et al.* (2004) observed a tendency for higher Fo values and lower Fm in water-stressed *versus* non-stressed plants, but a slight decrease in Fv/Fm in stressed plants was confirmed only with intense water stress.

Water status showed a better relationship with chlorophyll fluorescence variables measured in the dark-adapted than in the light-adapted leaves (Table 6). Both types of variables showed a negative relationship with leaf water potential in the most cases, disagreeing with Alves *et al.* (2013), who demonstrated that ΦPSII, ETR and qP decreased significantly in non-irrigated in comparison to irrigated grapevines. On the other hand, the current results show that water status was not linked with steady-state fluorescence, as has been explained by other authors (Maxwell & Johnson, 2000; Flexas *et al.*, 2002b).

Although most of the regressions were not significant (Table 6), it can be seen that midday leaf water potential was related more than predawn leaf water potential to chlorophyll fluorescence variables in light-adapted leaves. These results confirm previous findings obtained in the same study area in 2010 (Catalina *et al.*, 2011). Flexas *et al.* (2002a) showed that light-adapted chlorophyll fluorescence variables, such

TABLE 6
R-squared and slope (in brackets) of significant linear regressions of net assimilation (An) and fluorescence variables measured in dark-adapted and light-adapted leaves on midday (Ψ_{Md}) and predawn (Ψ_{Pd}) leaf water potential at different growth stages.

Year	Stage	Ψ_{Md}	An	Fo	Fm	Fv	Fv/Fm	Fo'	Fm'	Fs	Fs/Fo	Fv'/Fm'	Φ PSII	qP	NPQ	ETR
2011	Fruit set	Ψ_{Md}	(+)0.44**	(-)0.20*	(-)0.42**	(-)0.47**	(-)0.18*	(-)0.18*	(-)0.37**	(-)0.30*	(-)0.20*	(-)0.52***	(-)0.23*	(-)0.20*	(-)0.20*	(-)0.20*
		Ψ_{Pd}	(-)0.31*		(-)0.23*											(-)0.50***
	Pre- véraison	Ψ_{Md}	(-)0.26*													
	Véraison	Ψ_{Md}	(-)0.34**	(-)0.40**	(-)0.24*	(+)0.19*	(+)0.18*					(-)0.21*				(-)0.17*
2012	Fruit set	Ψ_{Md}														
		Ψ_{Pd}														
	Pre- véraison	Ψ_{Md}	(+)0.20*	(-)0.29**	(-)0.31**	(-)0.30**	(-)0.50***	(-)0.43**	(-)0.47**	(-)0.44**						
	Véraison	Ψ_{Pd}	(+)0.16*	(-)0.26*	(-)0.29*	(-)0.20*	(-)0.17*	(-)0.18*	(-)0.34**			(+)0.17*				(-)0.18*
2013	Fruit set	Ψ_{Md}	(+)0.16*			(+)0.19*										(+)0.17*
		Ψ_{Pd}														(-)0.18*
	Pre- véraison	Ψ_{Md}	(-)0.16*													(+)0.21*
	Véraison	Ψ_{Md}	(-)0.28**	(-)0.28**	(-)0.23*	(-)0.23*									(-)0.19*	(-)0.17*

Level of significance: * P < 0.05, ** P < 0.01 and *** P < 0.001; n = 24

as F_o' , F_m' , F_v' , F_v'/F_m' and F_s , had a strong relationship with midday leaf water potential.

The regressions of F_s/F_o with ψ_{Md} at fruit set in 2011 and at pre-véraison in 2012 were significant and had a negative slope (Table 6), and slightly increased the coefficients of determination obtained with F_s . Flexas *et al.* (2002b) reported that F_s/F_o is the best modulated test for the early detection of water stress in plants. The relationship could be negative or positive, depending on the stress level. A positive correlation is observed between F_s/F_o and photosynthetic photon flux density under irrigation, but an inverse correlation develops as drought progresses (Flexas *et al.*, 1999).

Non-photochemical quenching (NPQ) resulted in a significant relationship only with ψ_{pd} at pre-véraison in 2012 and at véraison in 2013 (Table 6). NPQ reflects the thermal dissipation of excessive excitation energy in the chloroplasts, and has a strong link with water status, but not with F_v/F_m (Maxwell & Johnson, 2000). The present results disagree with those of Flexas *et al.* (1999; 2000) and Alves *et al.* (2013), who showed a strong increase in NPQ as drought progressed. It is possible that, in the current study, there were no subzones with sufficiently high levels of water stress for this.

CONCLUSIONS

The presented results have revealed that foliar chlorophyll content cannot be considered an indicator of water stress in vineyards when the spatial variation of water status is limited and/or overlapped by a variation in iron or other nutrient status. Nevertheless, a linear regression of the chlorophyll fluorescence variables on foliar chlorophyll content was significant and positive in more of the growth stages and study years.

Linear regression relationships of chlorophyll fluorescence parameters with leaf water potential showed little significance in all the study years. However, variables such as F_m , F_v , F_v/F_m , F_s/F_o and Φ_{PSII} showed an increasing trend when midday leaf water potential decreased at pre-véraison and véraison. These variables could help assess the water status of vineyards for application in precision viticulture. In future studies it would be interesting to study the use of chlorophyll fluorescence variables as indicators of yield and grape quality in vineyards with significant spatial variability in water status.

LITERATURE CITED

- Adir, N., Zer, H., Shochat, S. & Ohad, I., 2003. Photoinhibition – A historical perspective. *Photosynth. Res.* 76, 343-370.
- Alves, F., Costa, J., Costa, P., Correia, C., Gonçalves, B., Soares, R. & Pereira, J.M., 2013. Grapevine water stress management in Douro Region: Long-term physiology, yield and quality studies in cv. Touriga Nacional. In: Group of International Experts of Vitivincultural Systems for Co-Operation (ed.). Proc. 18th Int. Symp. GiESCO, July 2013, Porto, Portugal.
- Baker, N.R. & Rosenqvist, E., 2004. Applications of chlorophyll fluorescence can improve crop production strategies: An examination of future possibilities. *J. Exp. Bot.* 55, 1607-1621.
- Bavaresco, L., Bertamini, M. & Iacono, F., 2006. Lime-induced chlorosis and physiological responses in grapevine (*Vitis vinifera* L. cv. Pinot blanc) leaves. *Vitis* 45, 45-46.
- Catalina, A., González, R., González, M.R., Zarco-Tejada, P.J. & Martín, P., 2011. Iron and water stress, photosynthetic efficiency differently affects vine photosynthetic efficiency and grape composition. Proc. 34th World Congress of Vine and Wine, June 2011, Porto, Portugal.
- Chaves, M.M., 1991. Effects of water deficits on carbon assimilation. *J. Exp. Bot.* 42, 1-16.
- Ciais, P., Reichstein, M., Viovy, N., Granier, A., Ogée, J., Allard, V., Aubinet, M., Buchmann, N., Bernhofer, Chr., Carrara, A., Chevallier, F., De Noblet, N., Friend, A.D., Friedlingstein, P., Grünwald, T., Heinesch, B., Keronen, P., Knohl, A., Krinner, G., Loustau, D., Manca, G., Matteucci, G., Miglietta, F., Ourcival, J.M., Papale, D., Pilegaard, K., Rambal, S., Seufert, G., Soussana, J.F., Sanz, M.J., Schulze, E.D., Vesala, T. & Valentini, R., 2005. Europe-wide reduction in primary productivity caused by the heat and drought in 2003. *Nature* 472, 529-533.
- Escalona, J.M., Flexas, J. & Medrano, H., 1999. Stomatal and non-stomatal limitations of photosynthesis under water stress in field grown grapevines. *Aust. J. Plant Physiol.* 26, 421-433.
- Flexas, J., Bota, J., Escalona, J.M., Sampol, B. & Medrano, H., 2002a. Effects of drought on photosynthesis in grapevines under field conditions: An evaluation of stomatal and mesophyll limitations. *Funct. Plant Biol.* 29, 461-471.
- Flexas, J., Briantais, J.M., Cerovic, Z., Medrano, H. & Moya, I., 2000. Steady-state and maximum chlorophyll fluorescence responses to water stress in grapevine leaves. A new remote sensing system. *Remote Sens. Environ.* 73, 282-297.
- Flexas, J., Escalona, J.M. & Medrano, H., 1999. Water stress induces different photosynthesis and electron transport rate regulation in grapevine. *Plant Cell & Environ.* 22, 39-48.
- Flexas, J., Escalona, J.M., Evain, S., Gulías, J., Moya, I., Osmond, C.B. & Medrano, H., 2002b. Steady-state chlorophyll fluorescence (F_s) measurements as a tool to follow variations of net CO_2 assimilation and stomatal conductance during water-stress in C_3 plants. *Physiol. Plant.* 114, 231-240.
- Gholamin, R. & Khayatnezhad, M., 2011. The effect of end season drought stress on the chlorophyll content, chlorophyll fluorescence parameters and yield in maize cultivars. *Sci. Res. Essay* 6, 5351-5357.
- González, R., Nunez, L.C., Martín, P., Berjon, A. & Zarco-Tejada, P.J., 2005. Estimación de la absorbancia de radiación PAR en hojas de vid a partir de su contenido en clorofila. In: Asociación Portuguesa de Ciencias Hortícolas (ed.). Proc. 5th Congreso Ibérico de Ciencias Hortícolas, Actas Portuguesas de Horticultura, Vol. 2, May 2005, Oporto, Portugal. pp. 384–389.
- Lawlor, D.W., 2002. Limitation to photosynthesis in water-stressed leaves: Stomata vs. metabolism and the role of ATP. *Ann. Bot.* 89, 871-885.
- Lichtenthaler, H.K., Buschmann, C. & Knapp, M., 2005. How to correctly determine the different chlorophyll fluorescence parameters and the chlorophyll fluorescence decrease ratio R_{Fd} of leaves with the PAM fluorometer. *Photosynthetica* 43, 379-393.
- Ling, Q., Huang, W. & Jarvis, P., 2011. Use of a SPAD-502 meter to measure leaf chlorophyll concentration in *Arabidopsis thaliana*. *Photosynth. Res.* 107, 209-214.
- Mafakheri, A.B., Siosemardeh, P.C., Bahramnejad, Y., Struik, T. & Sohrabi, S., 2010. Effect of drought stress on yield, proline and chlorophyll contents in three chickpea cultivars. *Aust. J. Crop Sci.* 4, 580-585.
- Mandal, K., Saravanan, R., Maiti, S. & Kothari, I.L., 2009. Effect of downy mildew disease on photosynthesis and chlorophyll fluorescence in *Plantago ovata* Forsk. *J. Plant Dis. Prot.* 116, 164-168.
- Maroco, J.P., Rodrigues, M.L., Lopes, C. & Chaves, M.M., 2002. Limitations to leaf photosynthesis in field-grown grapevine under drought-metabolic and modelling approaches. *Funct. Plant Biol.* 29, 451-459.

- Martín, P., González, M.R., Núñez, L.C., González, R. & Zarco-Tejada, P.J., 2007. Influencia de la clorosis férrica sobre el rendimiento fotosintético del viñedo cv. Tempranillo. In: GIENOL (ed). Actas de las IX Jornadas de los Grupos de Investigación Enológica, May 2007, Badajoz, Spain. pp. 437 – 439.
- Martín, P., Zarco-Tejada, P.J. & González, M.R., 2008. Diagnostico nutricional y recomendaciones de abonado en suelos calizos de la Ribera de Duero. *Vida Rural* 270, 26-32.
- Maxwell, K. & Johnson, G.N., 2000. Chlorophyll fluorescence – A practical guide. *J. Exp. Bot.* 51, 659-668.
- Medrano, H., Escalona, J.M., Cifre, J., Bota, J. & Flexas, J., 2003. A ten-year study on the physiology of two Spanish grapevine cultivars under field conditions: Effects of water availability from leaf photosynthesis to grape yield and quality. *Funct. Plant Biol.* 30, 607-619.
- Ojeda, H., Andara, C., Kraeva, E., Carbonneau, A. & Deloire, A., 2002. Influence of pre- and postveraison water deficit on synthesis and concentration of skin phenolic compounds during berry growth of *Vitis vinifera* cv. Syrah. *Am. J. Enol. Vitic.* 53, 261-267.
- Pallioti, A., Silvestroni, O. & Petoumenou, D., 2009. Photosynthetic and photoinhibition behavior of two field grown grapevine cultivars under multiple summer stresses. *Am. J. Enol. Vitic.* 60, 189-198.
- Souza, R.P., Machado, E.C., Silva, J.A.B., Lagôa, A.M.M.A. & Silveria, J.A.G., 2004. Photosynthetic gas exchange, chlorophyll fluorescence and some associated metabolic changes in cowpea (*Vigna unguiculata*) during water stress and recovery. *Environ. Exper. Bot.* 51, 45-56.
- Taiz, L. & Zeiger, E., 2006. *Plant physiology*. Sinauer Associates Inc., Massachusetts, USA.
- Van Leeuwen, C., Tregoat, O., Choné, X., Bois, B., Pernet, D. & Gaudillère, J.-P., 2009. Vine water status is a key factor in grape ripening and vintage quality for red Bordeaux wine. How can it be assessed for vineyard management purposes? *J. Int. Sci. Vigne Vin* 43, 121-134.
- Winkel, T. & Rambal, S., 1993. Influence of water stress on grapevines growing in the field: From leaf to whole-plant response. *Aust. J. Plant Physiol.* 20, 143-157.
- Zarco-Tejada, P.J., Catalina, A., González, M.R. & Martín, P., 2013. Relationships between net photosynthesis and steady-state chlorophyll fluorescence retrieved from airborne hyperspectral imagery. *Remote Sens. Environ.* 136, 247-258.