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Rhodamine B removal with activated carbons obtained from lignocellulosic waste



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ABSTRACT

By-products from the wax production process from carnauba palm (leaves), from the extraction of oil from macauba seeds (endocarp) and from pine nut production (shell) have been assessed for activated carbon production, using H₃PO₄ or CaCl₂ for their chemical activation. The resulting activated charcoals have been thoroughly characterized by elemental and thermal analysis, X-ray diffraction, infrared spectroscopy, electron scanning microscopy and N₂ adsorption behavior. Subsequently, their adsorption capacity for the removal of rhodamine B (RhB) from aqueous solutions has been evaluated by studying different parameters: contact time, pH, adsorbent dose, initial dye concentration and solution temperature. The adsorption of RhB followed Freundlich's model in all cases. Kinetic studies indicate that the pseudo-second order model can be used for describing the dynamics of the adsorption process. Thermodynamic parameters have also been evaluated, indicating its endothermic and spontaneous nature. Finally, a preliminary analysis of the impact of cellulose content in the carbon precursor materials has been conducted, by using a mixture of native cellulose with one of the lignocellulosic materials.

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1. Introduction

Synthetic dyes are widely used in industries, producing a large amount of toxic, carcinogenic, mutagenic, bio-accumulative wastewaters that pollute the environment (Chen et al., 2010) and which have been deemed as one of the most important groups of contaminants for the aquatic environment (Hosseini et al., 2011). Dyes are difficult to remove by conventional wastewater purification treatments due to their high stability against light, temperature, chemical and microbial attacks (Forgacs et al., 2004). Wastewater from industries also are contaminated by heavy metals (Lo et al., 2012), causing severe poisoning due to their high solubility in water and their non-biodegradability (Zaini et al., 2010). One of the most widely used products for the removal of these pollutants are activated carbons (activated charcoals), which can be produced from agricultural residues

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such as pecan nut shell (Bansode et al., 2004; Klasson et al., 2009; Ng et al., 2003), leather waste (Kong et al., 2013), olivewaste cake (Baccar et al., 2009), grass clippings, horse manure (Hao et al., 2013), rice husk waste (de Luna et al., 2013), etc.

The adsorption technique is used for the treatment of industrial effluents, and activated carbon is extensively used for the removal of many pollutants, but it has the disadvantage of having a high associated cost. Consequently, other low-cost materials have been proposed as alternatives, in particular residual biomass, which is a natural biodegradable renewable product, with a minimum associated cost and which also has good adsorption properties for contaminants, either metals or chemical compounds (Paraskeva et al., 2008).

The adsorption involves the separation of a substance, followed by its accumulation or concentration on the surface of another material (Metcalf and Eddy, 2003). Activated charcoal of plant origin is a versatile adsorbent of great economic interest for the treatment of water, either for human consumption or for industrial purposes.

The aim of this research has been to find an efficient way to



produce, characterize and use activated charcoals obtained from three little exploited lignocellulosic waste materials (from *Copernicia prunifera, Acrocomia aculeata* and *Pinus pinea*) for the adsorption of contaminants in wastewaters, taking Rhodamine-B (RhB) as a representative -and particularly harmful- example of industrial pollutant dye. The effect of impregnation of the raw materials with H₃PO₄ and CaCl₂ has been assessed, together with the impact of variations of the pH, concentration of adsorbate, contact time, temperature and adsorbent dose on the efficiency of the adsorption process. Rhodamine-B adsorption mechanisms have been analyzed through isotherms, kinetic and thermodynamic studies. Moreover, the effect of the cellulose content in the precursor materials on the RhB removal capability of the resulting activated charcoals has been evaluated by its artificial increase with native cellulose for the first time.

2. Materials and methods

2.1. Precursor materials

Carnauba palm (*C. prunifera*) leaves from Ceara (Brazil), macauba palm (*A. aculeata*) endocarp from Minas Gerais (Brazil) and stone pine (*P. pinea*) nut shells from Valladolid (Spain) have been selected as precursor lignocellulosic materials (da Silva Lacerda et al., 2015). Commercial native cellulose was supplied by Merck. Phosphoric acid (H₃PO₄, 85%) and hydrochloric acid (HCl, 37%) were purchased from Panreac, and calcium chloride (CaCl₂, anhydrous, >93%) from Sigma Aldrich.

2.2. Activated carbon preparation

The chemical activation of the lignocellulosic raw materials was carried out using phosphoric acid and calcium chloride. Prior to the impregnation of the materials, they were washed with deionized water until constant pH and then were grounded on a Retsch ZM-100 ultra centrifugal mill to a particle size <0.25 mm. Subsequently, the materials were impregnated with solutions of phosphoric acid (1 M) and calcium chloride (1 M), using 8 g of each lignocellulosic product in 100 mL of the respective solutions (H₃PO₄-1 M and CaCl₂-1 M) for 24 h. After impregnation, the samples were filtered and dried at room temperature for 24 h. The carbonization of the materials was conducted in a Mufla Digital oven mod. 10-PR/300 at 500 °C for 1 h under N₂ atmosphere. The materials resulting from carbonization were washed with 50 mL of hydrochloric acid (1 M) for 1 h and sonicated for 10 min in a JP Selecta ultrasonic cleaning bath, and then filtered and washed with deionized water until chlorides complete elimination and constant pH were attained. The charcoals were finally dried to remove all moisture. Aforementioned washing process is essential in order to remove the chemical activating agents, allowing higher porosity (Peláez-Cid and Teutli-León, 2012).

2.3. Physico-chemical characteristics of the activated carbons

The behavior of the impregnated materials against pyrolysis was studied in a Mettler Toledo TGA/SDTA851e/SF/1100 thermal analyzer, by heating the samples from 25 to 830 °C at a 25 °C/min rate under nitrogen atmosphere. The sample weights were in the 9.5–10.5 mg range, each of them determined with 0.01 mg precision.

C, H and N contents were measured using a Leco CHN-600 Elemental Analyzer. Since ash and metal contents are low for these lignocellulosic materials, the oxygen content was calculated by subtraction, following a procedure analogous to that of Pastor-Villegas et al. (2006).

The content of cellulose, hemicellulose and lignin was determined for each of the precursor materials as follows: the insoluble lignin content was calculated according to ANSI/ASTM standard (ANASI and ASTM, 1977) and holocellulose (hemicellulose + cellulose) was obtained after delignification of the sample and its content was determined using the technique described by Browning (1967). Cellulose content was determined according to ANSI/ASTM procedure (ANSI and ASTM, 1977) and hemicellulose content was then calculated by subtraction of cellulose content to that of holocellulose.

Structural characterization of the charcoals by X-ray powder diffraction was carried out in a Bruker D8 Advance Bragg-Brentano diffractometer, in reflection geometry. The diffractograms were recorded with 2θ sweep angles in the 5–70° range at 0.02 (2θ /sec) speed with 2 s exposure time.

The functional groups present on the surface were determined by infrared spectroscopy using a Thermo Nicolet 380 FT-IR apparatus equipped with a Smart Orbit Diamond ATR system.

The pore structure of the samples was characterized by nitrogen adsorption-desorption at 77 K using a Micromeritics ASAP 2020 surface area and porosity analyzer. Prior to the analysis, samples were degassed for 2 h under vacuum at 393 K.

The surface morphology was studied by scanning electron microscopy (SEM) in a FEI Quanta 200F apparatus.

2.4. RhB adsorption assays

Batch adsorption experiments were conducted to investigate the adsorption of Rhodamine B on the various activated carbons. All determinations were performed in triplicate biological replications, and the reported values correspond to the average.

To analyze the effect of pH on RhB adsorption, experiments were carried out at initial pH values ranging from 2 to 10. The initial pH was controlled by the addition of dilute HCl or NaOH solutions. The effect of the dose of adsorbent was studied by varying the amount of carbon in the solution (from 2 to 8 g/L). The influence of contact time was assessed by using the same dye concentration and changing the contact time from 5 to 240 min. The impact of the initial dye concentration was determined by isotherms at 25, 35 and 45 °C, varying the initial dye concentrations from 65 to 140 mg/ L while keeping the dose of adsorbent and the contact time fixed. Finally, with a view to determining to what extent the amount of cellulose in the precursor materials affected the RhB adsorption capacity, a preliminary study has been conducted by mixing different amounts of native cellulose (0 wt.%, 20 wt.% and 50 wt.%) with one of the lignocellulosic materials (pine nut shell) and comparing the percentage of removal of RhB for each the resulting activated charcoals. The aim of adding such native cellulose was to artificially increase the cellulose content prior to the activation phase (with H₃PO₄).

Rhodamine B concentrations in the solution before and after adsorption were determined with a Hitachi U-2001 UV–vis spectrophotometer at 552 nm. The amount adsorbed per unit mass of biosorbent (mg/g) is calculated using the following equation (Eq. (1)):

$$q_t = [V(C_0 - C_t)]/m$$
(1)

where C_0 (mg/L) is the initial concentration, C_t (mg/L) is the concentration in the liquid phase at time t, V (L) the volume of the solution and m (g) the mass of adsorbent. In equilibrium conditions, $q_t = q_e$ and $C_t = C_e$.

3. Results and discussion

3.1. Activated carbon characterization

The C, H, O and N content (% dry matter) of the precursor materials and their corresponding activated charcoals are summarized in Table S1. As expected, the carbon content is increased by carbonization (e.g., for carnauba leaves, it increases from 45.40% to 72.03% when impregnated with CaCl₂ or to 67.61% when impregnated with H₃PO₄). The low nitrogen content for macauba endocarp (0.03%) and pine nut shell (0.03%) ensures low NO_x emissions during the activated carbons preparation, while higher emissions can be expected for carnauba leaves (3.40%). The highest oxygen content in the precursor materials corresponds to carnauba leaves (48.7%), followed by pine nut shell (45.4%) and macauba endocarp (41.2%). These values are similar to those of other precursor materials or hardwoods such as those of white oak (42%) or yellow poplar (47.3%) (Jagtoyen and Derbyshire, 1998). It is worth noting that during the carbonization process, oxygen is consumed and this reduces its content in the carbonized materials to values between 1794% and 2693%

The cellulose, hemicellulose and lignin contents are summarized in Table S2. The highest cellulose content corresponds to carnauba leaves (37.5%), followed by pine nut shell (30.9%) and macauba endocarp (29.7%).

For all the charcoals under study, the X-ray powder diffraction patterns (Figure S2) show the presence of broad peaks, indicating that most of the samples are amorphous, which is a typical characteristic of porous adsorbents (Bouchelta et al., 2008). However, some peaks arise sporadically in the profile for the activated carbon obtained from carnauba leaves treated with H₃PO₄ (C_carnauba_H₃PO₄) and to a lesser extent in those of the carbons obtained from carnauba leaves treated with CaCl₂ (C_carnauba_CaCl₂) and from pine nut shell treated with H₃PO₄ (C_pine_nut_shell_H₃PO₄), indicating the presence of a small fraction of crystalline material.

The chemical structure and functional groups of the prepared activated carbons have been studied by ATR-FTIR spectroscopy (Figure S3). The spectra can be interpreted on the basis of the characteristic vibration frequencies of the most common functional groups and structural components. The band at 2356 cm⁻¹, present in the coal derivatives obtained from macauba (for both impregnations), can be assigned to carbon-oxygen groups due to the presence of the ketone group or to CO₂ contamination. The band at around 1600 cm⁻¹, present in all the activated carbons, can be attributed to aromatic rings or to C=C bonds and involves the presence of carbonyl groups and the aromatization of the precursor (Guo and Rockstraw, 2007). All charcoals also exhibit wide bands in the 1000–1300 cm^{-1} range, with a maximum at 1160 cm^{-1} , due to absorption caused by oxidized carbons and to the presence of C-O stretching vibrations in acids, alcohols, phenols, ethers and esters (Freeman and Gimblett, 1988). Although this band was observed in all the spectra, it is more intense in the H₃PO₄ impregnated materials, because it is also characteristic of phosphorus and carbon-phosphorous compounds (Freeman et al., 1988), with the presence of hydrogen bonded P–O groups (Phan et al., 2006; Santhi et al., 2014) of phosphates or polyphosphates, the stretching vibrations of C–O bonds in (aromatic) P–O–C groups (Santhi et al., 2014) and in O=P–OH groups. For all H₃PO₄ impregnated carbons, a shoulder can also be observed at 985 cm⁻¹, characteristic of the phosphate group, which is also associated with asymmetric stretching vibrations of C–O–P groups for pentavalent phosphorus (Brasquet et al., 2000). The peak at ca. 760 cm⁻¹ corresponds to C–Cl vibrations, for carbons activated with CaCl₂, and the band at 680 cm⁻¹ can be ascribed to C=O asymmetric vibration (Phan et al., 2006).

According to the shape of the nitrogen adsorption-desorption isotherms (Figure S4), they can be identified as type IV with hysteresis cycles, in accordance with the IUPAC classification (Sing, 1985). Type IV isotherms are typical of mesoporous solids (Zhou et al., 2009), where adsorption occurs in multilayers. This process is reflected in the central area of the upward isotherm as more layers on the solid surface are absorbed. Another property of type IV isotherms is the possibility of developing hysteresis loops, i.e., the adsorption and desorption curves do not match. Depending on the hysteresis curve type, in accordance with IUPAC, all our carbons exhibit a type H₄ hysteresis, which is typical of complex materials containing both micropores and mesopores, such as activated carbons. Figure S5 shows the pore size distribution according to the Barrett-Joyner-Halenda (BJH) (Barrett et al., 1951) method. The highest concentration of pores occurs in the 20–50 Å range, confirming the presence of micropores and mesopores.

The textural parameters for the carbonaceous materials are summarized in Table 1. The materials impregnated with H₃PO₄ have higher Brunauer-Emmett-Teller (BET) surface for macauba endocarp (371.12 m²/g) and pine nut shell (296.01 m²/g), whereas in the case of carnauba leaves the BET is higher for the carbon impregnated with CaCl₂ (402.43 m^2/g). Pore volumes range from 0.12 to 0.24 cm³/g, and the highest value corresponds again to C_carnauba_CaCl₂. The textural parameters for the activated carbons under study are within the ranges reported by other authors for different lignocellulosic residues, with surface areas ranging from 228 to 2410 m²/g and pore volumes in the 0.033–1.4 cm³/g range (Ioannidou and Zabaniotou, 2007; Zuo et al., 2009). It is worth noting that CaCl₂ activated carbons have larger average pore sizes (18.25-23.16 Å) in comparison with those activated with H_3PO_4 (17.26–19.97 Å), so it can be inferred that $CaCl_2$ activation produces larger pores.

The surfaces of all the raw materials and active charcoals have been analyzed by scanning electron microscopy (see Figure S6). Although the micropores and mesopores are not visible, the micrographs allow to observe the shape and location of macropores on the surface of the materials. By combining the pore size distribution obtained from nitrogen adsorption and the SEM micrographs, it is possible to obtain complete information on pores

Table 1

Textural parameters for the carbonaceous materials and their precursors, obtained from the N2 adsorption isotherm using standard conditions at 77 K.

	PPP (21)	P 1 4 24 2	24	- · · · · · · · · · · · · · · · · · · ·
Materials	BET surface area (m ² /g)	Pore volume (cm ³ /g)	Micropore volume (cm ³ /g)	Pore size (A)
Carnauba leaves	0.8581	0.000359	0.000359	21.246
Macauba endocarp	0.6028	0.002019	-0.000111	133.99
Pine nut shell	0.8608	0.00316	-0.000163	146.97
C_carnauba_CaCl ₂	430.695	0.249375	0.121191	23.160
C_carnauba_H ₃ PO ₄	402.430	0.200946	0.123078	19.973
C_macauba_CaCl ₂	265.493	0.121151	0.082070	18.252
C_macauba_H ₃ PO ₄	371.126	0.160181	0.126290	17.264
C_pine_nut_shell_CaCl ₂	290.053	0.136993	0.082738	18.892
C_pine_nut_shell_H ₃ PO ₄	296.017	0.132521	0.100583	17.907

development during carbonization and activation. In the precursor materials no macropores are observed in the SEM micrographs. With regard to the activated carbons obtained from carnauba leaves, macropores are found primarily within the fiber, in analogy with those reported for the activation of different fibers by other authors (Brasquet et al., 2000; Phan et al., 2006). In the case of the carbons derived from macauba endocarp and pine nut shell, macropores appear distributed all throughout the material surface. The macropores for CaCl₂ activated carbons show more variability and are larger than those of the materials activated with H₃PO₄. The distribution and shape of the pores in the activated carbons significantly vary depending on the raw material used.

3.2. Effect of pH on RhB adsorption capacity

The pH of the solution can either promote or prevent the dye adsorption, since it can affect the surface charge of the adsorbent and the adsorbate species in solution (Lin et al., 2013). Fig. 1 depicts the effect of the pH of the solution on RhB adsorption on the activated charcoals under study in the 2-10 pH range. As the pH increases from 2 to 8, a slight increase on the RhB adsorption capacity can be observed (confirmed by a one-way ANOVA), which is more pronounced for those carbons activated with H₃PO₄. At low pH values, the high concentration of H⁺ ions promotes the protonation of the functional groups of the adsorbent, charging it more positively and leading to repulsion of the cationic dye molecules. Moreover, at low pH values, the H⁺ ions compete with RhB molecules for the adsorption sites. On the other hand, when the pH of the solution is increased, the surface of the carbons gets negatively charged due to OH⁻ adsorption and deprotonation of functional groups, thus improving the adsorption by electrostatic attraction (Ahmad and Kumar, 2010). The decrease in the absorbance when the pH is higher than 8 is due to the formation of hydrated ions of RhB, which lead to a larger molecular structure (dimer) that hinders the entry of the dye into the pores of the material. This same behavior -with an increase in the absorption capacity and a subsequent decrease as the pH is increased- has also been reported in a study of RhB removal by activated carbon produced from Acacia nilotica leaves with H₂SO₄ (Santhi et al., 2014). Talebnia et al. (2010) also reported an optimal pH around 7 for the adsorption of RhB on montmorillonite $((Na,Ca)_{0.33}(Al,Mg)_2(Si_4O_{10})(OH)_2 \cdot nH_2O)$. The highest adsorption capacity the activated charcoals presented herein occurs at pH 8. Since the work system is designed to operate in natural waters at pH in the 6–8 interval, no pH adjustment was required for the tests on the effect of adsorbent dose discussed in next section.



Fig. 1. Effect of the pH of the solution on RhB adsorption (adsorbent dose = 2 g/L; temperature = 25 °C, [RhB]₀ = 65 mg/L; contact time = 120 min).



Fig. 2. Effect of activated carbon dosage on RhB removal ($[Rh-B]_0 = 80 \text{ mg/L}$, contact time = 120 min; pH = 7; temperature = 25 °C).

3.3. Effect of activated carbon dosage on adsorption

Fig. 2 shows the percentage of RhB removal as a function of the dose of activated charcoal. As expected, for all the activated carbon atoms under study, dye adsorption increased with increasing carbon concentration (e.g., from 87.6% to 99.85% for C_carnauba_-CaCl₂). This increase in the removal percentage can be attributed to the increase in the available surface area and availability of more adsorption sites (Garg et al., 2004). On the basis of the results presented in Fig. 2, it can inferred that the most efficient activated charcoal dose is 4 g/L, since further increase in dose (6 or 8 g/L) does not lead to a significant increase in the removal of the dye. This can be attributed to overlapping or aggregation of adsorption sites, resulting in a decrease in total adsorbent surface area available to the pollutant and an increase in diffusion path length (Garg et al., 2004). The same behavior has also been reported by other authors, in relation to the absorption of RhB (Namasivayam et al., 2001) and to the adsorption of other dyes (Wang et al., 2010).

3.4. Impact of contact time on RhB removal

As it can be observed from Fig. 3, the adsorption is very rapid at first and it gradually decreases till equilibrium is reached after 120 min, for all the activated carbons under study. In general, it can be assumed that the dyes removal mechanism, by adsorption, consists of the following steps (Lazaridis and Asouhidou, 2003): (i) diffusion across the fluid phase surrounding the adsorbent particles, i.e., external diffusion or film diffusion; (ii) diffusion in the liquid contained in the pores and/or along the pore walls, which is so-called internal diffusion or intra-particle diffusion; and (iii) adsorption and desorption between the adsorbate and active sites. i.e., mass action. The decrease in the amount of dve adsorption versus time may be due to aggregation processes of the dye molecules. This aggregation can prevent migration of the adsorbate, saturate the adsorption sites and generate a resistance to diffusion of dye molecules. Other authors have also reported a similar behavior in the absorption of RhB, with a rapid initial phase followed by a gradually slower one (Fernandez et al., 2014; Gad and El-Sayed, 2009; Wang and Zhu, 2007).

3.5. Effect of dye initial concentration and solution temperature

Fig. 4 depicts the effect of varying the initial RhB concentration



Fig. 3. Effect of contact time on RhB adsorption for the different carbons activated with (*a*) CaCl₂ and (*b*) H₃PO₄ (pH = 7; temperature = 25 °C, [RhB]₀ = 80 mg/L; absorbent dose = 2 g/L). Fitting of the data conducted by kinetic models.

on the adsorption for the different activated carbons at 25, 35 and 45 °C. An increase in the adsorption capacity with increasing dye concentration is observed. The largest increase occurs for C_carnauba_CaCl₂, for which it changes from 32.61 to 42.10 mg/L. This increase in adsorption capacity is probably due to the increase in the driving force of the concentration gradient, which decreases resistance to the mass transfer of ions between the aqueous medium and the sorbent (Aksu and Dönmez, 2003). This behavior has been widely reported in the literature (Gad and El-Sayed, 2009; Kadirvelu et al., 2005; Santhi et al., 2014).

Moreover, the increase in temperature also resulted in an increase in the adsorption capacity for all activated carbons. Amongst the different materials under study, C_macauba_CaCl₂ is the one with the most significant increase in the adsorption capacity, which changed from 33.44 mg/L at 25 °C to 34.87 mg/L at 45 °C. This increase in adsorption capacity with temperature is due to the fact that the process is endothermic, leading to an increase in the intraparticle diffusion speed of the dye in the pores or to the formation of new adsorption sites (Karthikeyan et al., 2005). Wang and Zhu (2007) also reported the endothermic nature of the RhB adsorption process on a commercial activated carbon pretreated with acid.

3.6. Kinetic study

The experimental data have been fitted by nonlinear regression to the two most commonly used kinetic models in the literature: the pseudo-first order model and the pseudo-second order model, which are described below.

The pseudo-first order model (Lagergren, 1898) considers that the occupation rate of adsorption sites is proportional to the number of unoccupied sites (see Eq. (2)). By integrating this equation under conditions q = 0 at t = 0 and $q = q_t$ at t = t, Eq. (3) is obtained, where q_t (mg/g) is the absorption of the dye at time t, q_e (mg/g) is the adsorption capacity in equilibrium, t (min) is the exposure time and k_1 (h⁻¹) is the adsorption rate constant:

$$\frac{dq}{dt} = k_1(q_e - q) \tag{2}$$

$$q_t = q_e \left(1 - e^{-k_1 t} \right) \tag{3}$$

The pseudo-second order model (Ho, 1995) describes how the adsorption mechanism follows a second-order reaction. The absorption rate is then proportional to the square of the number of unoccupied sites (see Eq. (4)). By integrating this equation under the following conditions, q = 0 at t = 0 and $q = q_t$ at t = t, Eq. (5) is obtained, where k_2 (g/mg·h) is the rate constant of the pseudo-second order adsorption equation:

$$\frac{dq}{dt} = k_2 (q_e - q)^2 \tag{4}$$

$$l_t = \frac{t}{1/k_2 q_e^2 + t/q_e}$$
(5)

The adsorption kinetic fittings are shown in Fig. 3, and all the calculated kinetic parameters are summarized in Table 2. C_carnauba_H₃PO₄ presents the fastest rate of dye removal (k_1) , followed C_carnauba_CaCl₂ > C_pine_nut_shell_CaCl₂ > C_macbv $auba_CaCl_2 > C_pine_nut_shell_H_3PO_4 > C_macauba_H_3PO_4.$ Although correlation coefficients higher than 0.99 were obtained for all activated carbons under study for the pseudo-first order model, the calculated adsorption capacities $(q_{e_{cal}})$ were significantly different from the experimental values $(q_{e_{exp}})$ (Table 2). A similar situation has been reported by Ahmad et al. (2012), who observed that for very quick adsorption processes the r^2 values were similar for both the pseudo-first and the pseudo-second order models, and then attention should be paid to $q_{e_{cal}}$ values so as to distinguish which fitting is the most suitable. Consequently, it can be concluded that the fitting of the dye adsorption with a pseudofirst order is not good.

On the other hand, when the pseudo-second order kinetic model is chosen, not only the correlation coefficients are higher than 0.99 (Table 2), but the calculated adsorption values (q_{e_cal}) are also in good agreement with the experimental data (q_{e_exp}). This better fitting to a pseudo-second-order model suggests that the adsorption rate is more dependent on the availability of adsorption sites than on the dye concentration in the solution. Similar phenomena have been observed for RhB adsorption on different biosorbents (Kadirvelu et al., 2005; Wang et al., 2010).

3.7. Adsorption isotherms

The adsorption data for the different activated carbons have been fitted by non-linear Langmuir and Freundlich models. SigmaPlotTM v.10 (Systac Software Inc) software was used for these fittings.



Fig. 4. Effect of RhB initial concentration on the adsorption by (*a*) C_carnauba_CaCl₂; (*b*) C_carnauba_H₃PO₄; (*c*) C_macauba_CaCl₂; (*d*) C_macauba_H₃PO₄; (*e*) C_pine_nut_shell_CaCl₂; (*f*) C_pine_nut_shell_H₃PO₄ (pH = 7; adsorbent dose = 2 g/L; contact time = 120 min) at three different temperatures (25, 35 and 45 °C). The data have been fitted according to Langmuir y Freundlich non-linear models.

Table 2

Kinetic parameters for rhodamine B adsorption for the different activated carbons under study.

Samples	q_{e_exp}	Pseudo-first order model			Pseudo-second order model		
		k_1 (min)	$q_{\rm e_cal} ({ m mg g}^{-1})$	r ²	$k_2 (\mathrm{g} \; \mathrm{mg}^{-1} \; \mathrm{min}^{-1})$	$q_{\rm e_{cal}} ({ m mg~g^{-1}})$	r ²
C_carnauba_CaCl ₂	33.315	0.769	36.186	0.998	0.236	36.363	0.998
C_carnauba_H ₃ PO ₄	32.429	0.851	32.365	0.999	0.431	32.467	0.999
C_macauba_CaCl ₂	31.371	0.850	31.197	0.999	0.350	31.347	0.999
C_macauba_H ₃ PO ₄	31.099	0.856	30.948	0.999	0.280	31.055	0.999
C_pine_nut_shell_CaCl ₂	31.855	0.861	31.687	0.999	0.299	31.847	0.999
C_pine_nut_shell_H ₃ PO ₄	31.184	0.920	31.035	0.999	0.332	31.152	0.999

The Langmuir model (Eq. (6)) assumes that adsorption occurs in a monolayer, where the active sites are identical and energetically equivalent, that there are no interactions between the adsorbed molecules and that no transmigration of the adsorbed molecules takes place on the adsorption surface (Langmuir, 1918):

$$q_e = \frac{k_L q_{\max} C_e}{1 + k_L C_e} \tag{6}$$

where $q_e(mg/g)$ is the adsorption capacity in equilibrium, $C_e(mg/L)$ is the pollutant concentration in equilibrium, k_L (L/mg) is the

Langmuir constant related to the adsorption energy, which quantitatively reflects the affinity between the adsorbent and adsorbate, and q_{max} (mg/g) is the theoretical maximum adsorption capacity.

The essential characteristics of the Langmuir adsorption isotherm can be expressed as a dimensionless constant, called *separation factor* or *equilibrium parameter* (R_L), which is defined by Eq. (7)) (Hall et al., 1966):

$$R_L = \frac{1}{1 + k_L C_0} \tag{7}$$

where C_0 is the initial dye concentration (mg/L). The R_L value indicates the shape of the isotherm, which can be irreversible ($R_L = 0$), favorable ($0 < R_L < 1$), linear ($R_L = 1$) or unfavorable ($R_L > 1$).

Freundlich isotherm is an empirical equation (Eq. (8)) which considers the possibility of multilayer adsorption in multiple layers with a heterogeneous energy distribution of the active sites, accompanied by interactions between the adsorbed molecules (Freundlich, 1906):

$$q_e = k_F \cdot C_e^{1/n} \tag{8}$$

where $k_{\rm F}$ (mg g⁻¹(mg L⁻¹)^{1/n}) is the Freundlich isotherm constant, which is an indicator of the adsorptive capacity of an adsorbent for a solute, and *n* is a measure of the intensity of the adsorption or surface heterogeneity (a value closer to zero represents a more heterogeneous surface).

The fittings of the experimental data to the Langmuir and Freundlich isotherm models at different temperatures are depicted in Fig. 4. The correlation coefficients (r^2) and the constants obtained from the application of the Langmuir and Freundlich models are summarized in Table 3. In light of these data, Freundlich model has a better fit to the experimental data than Langmuir model in all

cases. This reveals the heterogeneity of the surface of the activated carbons under study.

The R_L values (Table 3) confirm that the dye adsorption is favorable, with values below 1 for all the activated carbons. The maximum adsorption (q_{max}) for the different coals decrease in the following order: C_carnauba_CaCl₂ > C_carnauba_H₃PO₄ > C_macauba_CaCl₂ > C_pine_nut_shell_CaCl₂ > C_pine_nut_shell_H₃PO₄ > C_macauba_H₃PO₄. All the charcoals activated with CaCl₂ showed higher q_{max} values than those activated with H₃PO₄, as expected for the larger size of their pores (discussed above).

Table S3 summarizes a comparison of the absorption capacity for the activated charcoals presented herein with others found in the literature (for a more general comparison of other lignocellulosic products as adsorbents, the interested reader is referred to, for example, the work by Parab et al. (2009)). The activated carbons prepared from the selected lignocellulosic waste materials have adsorption capacities between 33.08 and 39.98 mg/g, comparable to the values reported for commercial activated carbon (Bhadusha and Ananthabaskaran, 2012) and above the average of the values reported in the literature (which vary from 1.6 to 60.8 mg/L). Nonetheless, it should be noted that an accurate comparison is not feasible in this case, given the differences on the nature of the

Table 3

Adsorption isotherm param	neters for rhodamine B ads	sorption for the different	activated carbons under study.
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$ \frac{q_{max}}{(mg~r)} - \frac{k_{L}(L~gg^{-1})}{r^2} - \frac{r^2}{R_L} - \frac{g}{r} - \frac$	Sample	T (°C)	Langmuir constants				
C_carnauba_CCl2 25 9.218 38.881 0.976 32.16.07 2.057 45 39.983 52.488 0.975 2.966.07 2.093 C_carnauba_H3PO4 25 35.779 2.459 0.994 5086.06 0.797 45 35.958 2.497 0.995 5.016.06 0.602 7 35 33.830 2.661 0.996 4.705.06 0.671 7 31.23 0.998 4.005.06 0.672 0.661 0.661 7 31.33 3.497 0.998 3.586.06 0.662 7 31.33 3.697 0.998 3.586.06 0.662 7 31.33 3.697 0.998 3.586.06 0.662 7 3.23 0.998 3.586.06 0.562 7 3.3363 3.697 0.998 3.586.06 0.562 7 3.3365 3.697 0.997 3.295.06 0.563 7 3.53 3.685 3.795 0.997 3.295.06 0.553 7 3.53 3.2732 6.899 0.997 1.815.06 0.583 7 3.53 3.2732 6.899 0.997 1.815.06 0.			$q_{ m max} \ ({ m mg~g}^{-1})$	K_L (L mg ⁻¹)	r ²	R_L	σ
$ \begin{array}{c ccr} & 35 & 39.732 & 43.164 & 0.975 & 2.90E-07 & 2.093 \\ 45 & 39.983 & 52.488 & 0.975 & 2.38E-07 & 2.159 \\ 2.carnauba_H_3PO_4 & 25 & 35.279 & 2.459 & 0.994 & 5.08E-06 & 0.801 \\ 45 & 35.998 & 2.497 & 0.995 & 5.01E-06 & 0.809 \\ 2.macauba_CaCl_2 & 25 & 32.959 & 1.505 & 0.996 & 8.31E-06 & 0.671 \\ 35 & 33.830 & 2.661 & 0.996 & 4.70E-06 & 0.671 \\ 35 & 33.830 & 2.661 & 0.9998 & 4.00E-06 & 0.360 \\ 35 & 33.363 & 3.973 & 0.998 & 3.58E-06 & 0.365 \\ 35 & 33.363 & 3.973 & 0.998 & 3.58E-06 & 0.365 \\ 35 & 33.363 & 3.973 & 0.998 & 3.58E-06 & 0.626 \\ 2.macauba_H_3PO_4 & 25 & 33.065 & 2.688 & 0.998 & 4.65E-06 & 0.462 \\ 25 & 33.065 & 2.688 & 0.998 & 4.65E-06 & 0.462 \\ 35 & 33.065 & 2.688 & 0.998 & 4.65E-06 & 0.462 \\ 35 & 33.080 & 1.105 & 0.997 & 3.28E-06 & 0.558 \\ 3.997 & 3.08E-06 & 0.558 & 0.997 & 3.28E-06 & 0.558 \\ 3.998 & 4.117 & 0.997 & 3.04E-06 & 0.601 \\ 35 & 33.080 & 1.105 & 0.997 & 1.31E-06 & 0.588 \\ 3.999 & 0.997 & 1.31E-06 & 0.588 & 0.998 \\ 4.5 & 30.080 & 1.105 & 0.997 & 1.31E-06 & 0.588 \\ 0.997 & 1.31E-06 & 0.588 & 0.987 & 0.708 & 0.588 \\ 0.997 & 1.31E-06 & 0.588 & 0.987 & 0.708 & 0.588 & 0.595 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.997 & 0.703 & 0.998 & 0.533 & 0.3256 & 0.998 & 0.613 & 0.997 & 0.703 & 0.997 & 0.703 & 0.998 & 0.533 & 0.3267 & 2.1989 & 0.998 & 0.533 & 0.3267 & 2.1980 & 0.998 & 0.533 & 0.3267 & 2.1980 & 0.998 & 0.533 & 0.3267 & 2.1980 & 0.999 & 0.325 & 0.998 & 0.533 & 0.3267 & 2.1980 & 0.999 & 0.335 & 0.399 & 0.334 & 0.999 & 0.335 & 0.9$	C_carnauba_CaCl ₂	25	39.218	38.881	0.976	3.21E-07	2.057
$ \begin{array}{c c} +6 & 39.983 & 52.488 & 0.975 & 2.38E.07 & 2.159 \\ C_{carnauba_H_3PO_4} & 25 & 35.79 & 2.499 & 0.994 & 5.08E.06 & 0.891 \\ 35 & 35.796 & 2.479 & 0.996 & 5.04E.06 & 0.079 \\ 45 & 35.998 & 2.497 & 0.996 & 8.31E.06 & 0.0642 \\ 35 & 33.830 & 2.661 & 0.996 & 4.70E.06 & 0.671 \\ 45 & 34.217 & 3.123 & 0.998 & 3.58E.06 & 0.350 \\ c_macauba_H_3PO_4 & 25 & 32.651 & 3.492 & 0.998 & 3.58E.06 & 0.350 \\ c_macauba_H_3PO_4 & 25 & 33.605 & 2.688 & 0.997 & 2.95E.06 & 0.662 \\ 45 & 33.691 & 4.241 & 0.997 & 2.95E.06 & 0.652 \\ 35 & 33.665 & 3.795 & 0.997 & 3.29E.06 & 0.553 \\ c_pine_nut_shell_H_3PO_4 & 25 & 32.498 & 4.113 & 0.998 & 3.03E.06 & 0.553 \\ c_pine_nut_shell_H_3PO_4 & 25 & 32.498 & 4.113 & 0.998 & 3.03E.06 & 0.550 \\ 45 & 33.09 & 10.05 & 0.997 & 1.31E.06 & 0.589 \\ \hline \\ Sample & T^{(*C)} & \hline T(m t - 1)^{(n)} & n & r^2 & \sigma \\ (m t - 1)^{(n)} & n & r^2 & \sigma \\ (m t - 1)^{(n)} & n & r^2 & \sigma \\ (m t - 1)^{(n)} & n & r^2 & \sigma \\ (m t - 1)^{(n)} & n & r^2 & \sigma \\ (m t - 1)^{(n)} & n & r^2 & \sigma \\ (m t - 1)^{(n)} & 1 & 0.998 & 0.997 & 0.635 \\ C_carnauba_H_3PO_4 & 25 & 3.062 & 23.463 & 0.997 & 0.635 \\ C_carnauba_H_3PO_4 & 25 & 3.062 & 23.463 & 0.997 & 0.635 \\ C_carnauba_H_3PO_4 & 25 & 3.062 & 23.463 & 0.997 & 0.635 \\ C_carnaub_H_3PO_4 & 25 & 3.062 & 23.463 & 0.997 & 0.635 \\ C_carnaub_H_3PO_4 & 25 & 29.755 & 21.599 & 0.998 & 0.539 \\ C_carnaub_H_3PO_4 & 25 & 29.755 & 21.599 & 0.998 & 0.539 \\ C_macaub_H_3PO_4 & 25 & 29.755 & 21.599 & 0.998 & 0.539 \\ C_macaub_H_3PO_4 & 25 & 29.555 & 30.064 & 0.999 & 0.325 \\ C_macaub_H_3PO_4 & 25 & 29.555 & 30.6655 & 0.999 & 0.326 \\ C_macaub_H_3PO_4 & 25 & 29.555 & 30.664 & 0.999 & 0.325 \\ C_macaub_H_3PO_4 & 25 & 29.555 & 30.664 & 0.999 & 0.326 \\ C_macaub_H_3PO_4 & 25 & 29.655 & 30.0664 & 0.999 & 0.326 \\ C_macaub_H_3PO_4 & 25 & 29.655 & 30.0664 & 0.999 & 0.326 \\ C_macaub_H_3PO_4 & 25 & 29.505 & 30.664 & 0.999 & 0.326 \\ C_macaub_H_3PO_4 & 25 & 29.505 & 30.664 & 0.999 & 0.326 \\ C_pine_nut_shell_H_3PO_4 & 25 & 29.505 & 30.664 & 0.999 & 0.326 \\ C_pine_nut_shell_H_3PO_4 & 25 & 29.505 & 30.664 & 0.999 & 0.326 \\ C_pine_$		35	39.732	43.164	0.975	2.90E-07	2.093
$\begin{array}{c} \mbox{C} c_{arrauba,HyP0_4} & 25 & 35.279 & 2.459 & 0.994 & 5.081-06 & 0.891 \\ 35 & 35.796 & 2.479 & 0.995 & 5.011-06 & 0.809 \\ \mbox{C} macauba,CaCl_2 & 25 & 32.959 & 1.505 & 0.996 & 3.311-06 & 0.672 \\ 35 & 33.830 & 2.661 & 0.996 & 4.701-06 & 0.671 \\ \mbox{A} & 25 & 32.2651 & 3.492 & 0.998 & 3.581-06 & 0.365 \\ \mbox{C} macauba,HyP0_4 & 25 & 32.651 & 3.492 & 0.998 & 3.581-06 & 0.522 \\ \mbox{A} & 35 & 33.636 & 3.973 & 0.998 & 3.581-06 & 0.525 \\ \mbox{C} macauba,HyP0_4 & 25 & 33.666 & 2.688 & 0.998 & 4.651-06 & 0.661 \\ \mbox{A} & 35 & 33.694 & 4.241 & 0.997 & 2.951-06 & 0.553 \\ \mbox{C} macauba,HyP0_4 & 25 & 33.606 & 2.688 & 0.998 & 4.651-06 & 0.661 \\ \mbox{A} & 35 & 33.671 & 4.131 & 0.998 & 3.031-06 & 0.555 \\ \mbox{C} macauba,HyP0_4 & 25 & 32.498 & 4.117 & 0.997 & 3.041-06 & 0.569 \\ \mbox{A} & 5 & 33.080 & 11.055 & 0.997 & 1.131-06 & 0.569 \\ \mbox{S} & 35 & 35.051 & 23.211 & 0.977 & 0.703 \\ \mbox{C} & 5 & 35.051 & 23.211 & 0.997 & 0.703 \\ \mbox{C} & 5 & 35.051 & 23.211 & 0.997 & 0.703 \\ \mbox{A} & 5 & 35.051 & 23.211 & 0.997 & 0.703 \\ \mbox{C} & 5 & 30.0267 & 21.899 & 0.998 & 0.543 \\ \mbox{C} & 5 & 30.267 & 21.899 & 0.998 & 0.533 \\ \mbox{C} & 5 & 30.0267 & 21.899 & 0.998 & 0.533 \\ \mbox{C} & 5 & 30.0267 & 21.899 & 0.998 & 0.533 \\ \mbox{C} & 5 & 30.0267 & 21.899 & 0.998 & 0.533 \\ \mbox{C} & 5 & 30.0267 & 21.899 & 0.998 & 0.533 \\ \mbox{C} & 5 & 30.0267 & 21.899 & 0.998 & 0.533 \\ \mbox{C} & 5 & 30.0267 & 21.899 & 0.998 & 0.533 \\ \mbox{C} & 5 & 30.039 & 28.655 & 0.999 & 0.034 \\ \mbox{C} & 5 & 30.031 & 35.855 & 0.999 & 0.034 \\ \mbox{C} & 5 & 30.033 & 35.1259 & 0.999 & 0.333 \\ \mbox{C} & 5 & 30.333 & 35.1259 & 0.999 & 0.333 \\ \mbox{C} & 1.99 & 0.335 & 0.599 & 0.034 \\ \mbox{C} & 1.99 & 0.335 & 0.599 & 0.034 \\ \mbox{C} & 1.99 & 0.335 & 0.599 & 0.034 \\ \mbox{C} & 5 & 30.034 & 35.335 & 0.999 & 0.034 \\ \mbox{C} & 5 & 30.048 & 34.108 & 0.999 & 0.335 \\ \mbox{C} & 5 & 30.048 & 34.108 & 0.999 & 0.335 \\ \mbox{C} & 1.99 & 0.325 & 0.999 & 0.336 \\ \mbox{C} & 1.99 & 0.995 & 0.336 & 0.147 & 3.336 & 0.999 & 0.336 \\ \mbox{C} & $		45	39.983	52.488	0.975	2.38E-07	2.159
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	C_carnauba_H ₃ PO ₄	25	35.279	2.459	0.994	5.08E-06	0.891
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		35	35.796	2.479	0.996	5.04E-06	0.779
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		45	35.998	2.497	0.995	5.01E-06	0.809
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	C_macauba_CaCl ₂	25	32.959	1.505	0.996	8.31E-06	0.642
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		35	33.830	2.661	0.996	4.70E-06	0.671
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		45	34.217	3.123	0.998	4.00E-06	0.530
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	C_macauba_H ₃ PO ₄	25	32.651	3.492	0.998	3.58E-06	0.365
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		35	33.363	3.973	0.998	3.15E-06	0.522
$\begin{array}{c c} C_{pine_nut_shell_CaCl_2} & 25 & 33.606 & 2.688 & 0.998 & 4.65E-06 & 0.462 \\ 35 & 33.685 & 3.795 & 0.997 & 3.29E-06 & 0.553 \\ 45 & 33.971 & 4.131 & 0.998 & 3.03E-06 & 0.559 \\ 35 & 32.732 & 6.489 & 0.997 & 1.41E-06 & 0.601 \\ 35 & 32.732 & 6.489 & 0.997 & 1.41E-06 & 0.569 \\ 45 & 30.08 & 11.055 & 0.997 & 1.31E-06 & 0.569 \\ \hline \\ tite{tite{tite{tite{tite{tite{tite{tit$		45	33.694	4.241	0.997	2.95E-06	0.568
$ \begin{array}{c c} 35 & 33.685 & 3.795 & 0.997 & 3.29E-06 & 0.553 \\ 33.087 & 33.971 & 4.131 & 0.998 & 3.03E-06 & 0.555 \\ 32.498 & 4.117 & 0.997 & 3.04E-06 & 0.601 \\ 35 & 32.732 & 6.899 & 0.997 & 1.81E-06 & 0.589 \\ 45 & 33.080 & 11.055 & 0.997 & 1.81E-06 & 0.560 \\ \hline \\ $	C_pine_nut_shell_CaCl ₂	25	33.606	2.688	0.998	4.65E-06	0.462
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		35	33.685	3.795	0.997	3.29E-06	0.553
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		45	33.971	4.131	0.998	3.03E-06	0.555
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	C_pine_nut_shell_H ₃ PO ₄	25	32.498	4.117	0.997	3.04E-06	0.601
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		35	32.732	6.899	0.997	1.81E-06	0.589
$\begin{split} \text{Sample} & & & & & & & & & & & & & & & & & & &$		45	33.080	11.055	0.997	1.13E-06	0.560
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Sample	<i>T</i> (°C)	Fre	eundlich constants			
C_carnauba_CaCl2 25 35.062 23.463 0.997 0.703 35 35.601 23.211 0.997 0.635 A5 35.953 23.285 0.998 0.613 C_carnauba_H3P04 25 29.755 21.599 0.998 0.539 C_arnauba_CaCl2 25 29.755 21.989 0.998 0.539 C_macauba_CaCl2 25 27.459 21.832 0.999 0.333 C_macauba_A13PO_4 25 29.306 26.191 0.999 0.332 A5 30.039 28.644 0.999 0.332 A5 30.141 36.693 0.999 0.399 A5 30.333 35.1259 0.999 0.326 A5 30.333 35.1259 0.999 0.326 A5 30.333 35.1259 0.999 0.328 A5 30.488 34.108 0.999 0.328 A5 30.488 34.108 0.999 0.359 A5 30.488 34.108 0.999 0.359 A5			K _F (m	(mg g^{-1}) $(\text{mg L}^{-1})^{1/n})$	n	r ²	σ
35 35.601 23.211 0.997 0.635 45 35.953 23.285 0.998 0.613 25 29.755 21.599 0.998 0.539 35 30.267 21.989 0.998 0.533 C_macauba_CaCl2 25 27.459 21.832 0.997 0.607 C_macauba_CaCl2 25 27.459 21.832 0.999 0.333 35 29.306 26.191 0.999 0.325 A5 30.039 28.644 0.999 0.325 A5 30.333 35.1259 0.999 0.326 C_macauba_H_3PO_4 25 29.555 38.655 0.999 0.326 C_pine_nut_shell_CaCl2 25 29.629 30.664 0.999 0.326 C_pine_nut_shell_H_3PO_4 25 30.488 34.108 0.999 0.312 C_pine_nut_shell_H_3PO_4 25 30.488 34.108 0.999 0.325 C_pine_nut_shell_H_3PO_4 25 30.406	C_carnauba_CaCl ₂	25	35	.062	23.463	0.997	0.703
4535.95323.2850.9980.613C_carnauba_H_3PO42529.75521.5990.9980.5393530.26721.9890.9980.543A530.43121.9360.9970.607C_macauba_CaCl22527.45921.8320.9990.333C_macauba_ACaCl22529.30626.1910.9990.325C_macauba_H_3PO42529.55538.6550.9990.094A530.14136.9830.9990.326C_pine_nut_shell_CaCl22529.62930.6640.9990.326A530.14733.5360.9990.326C_pine_nut_shell_H_3PO42529.62930.6640.9990.312C_pine_nut_shell_H_3PO42530.48834.1080.9990.312C_pine_nut_shell_H_3PO42530.40649.3140.9990.325C_pine_nut_shell_H_3PO43530.40649.3140.9990.325S30.40649.3140.9990.325S30.40649.3140.9990.325		35	35	.601	23.211	0.997	0.635
C_arnauba_H_3PO4 25 29.755 21.599 0.998 0.539 35 30.267 21.989 0.998 0.543 A5 30.431 21.936 0.997 0.607 C_macauba_CaCl2 25 27.459 21.832 0.999 0.333 35 29.306 26.191 0.999 0.555 C_macauba_H_3PO4 25 29.555 38.655 0.999 0.054 C_macauba_H_3PO4 25 29.555 38.655 0.999 0.054 C_macauba_H_3PO4 25 29.629 36.644 0.999 0.333 C_pine_nut_shell_CaCl2 25 29.629 30.664 0.999 0.326 C_pine_nut_shell_H_3PO4 25 29.629 30.664 0.999 0.328 35 30.147 33.536 0.999 0.312 A5 30.488 34.108 0.999 0.312 A5 30.488 34.108 0.999 0.329 C_pine_nut_shell_H_3PO4 25<		45	35	.953	23.285	0.998	0.613
35 30.267 21.989 0.998 0.543 45 30.431 21.936 0.997 0.607 C_macauba_CaCl2 25 27.459 21.832 0.999 0.333 35 29.306 26.191 0.999 0.355 C_macauba_H_3PO_4 25 29.555 38.655 0.999 0.094 35 30.141 36.983 0.999 0.333 C_macauba_H_3PO_4 25 29.555 38.655 0.999 0.094 35 30.141 36.983 0.999 0.333 C_pine_nut_shell_CaCl2 25 29.629 30.664 0.999 0.326 C_pine_nut_shell_H_3PO_4 25 29.629 30.664 0.999 0.328 45 30.488 34.108 0.999 0.312 45 30.488 34.108 0.999 0.329 C_pine_nut_shell_H_3PO_4 25 29.506 38.957 0.999 0.392 35 30.406 49.314 <td< td=""><td>C_carnauba_H₃PO₄</td><td>25</td><td>29</td><td>.755</td><td>21.599</td><td>0.998</td><td>0.539</td></td<>	C_carnauba_H ₃ PO ₄	25	29	.755	21.599	0.998	0.539
45 30.431 21.936 0.997 0.607 C_macauba_CaCl2 25 27.459 21.832 0.999 0.333 35 29.306 26.191 0.999 0.355 C_macauba_H_3PO_4 25 30.039 28.644 0.999 0.094 C_macauba_H_3PO_4 25 29.555 38.655 0.999 0.094 35 30.141 36.983 0.999 0.333 C_pine_nut_shell_CaCl2 25 29.629 30.664 0.999 0.288 35 30.147 33.536 0.999 0.312 45 30.488 34.108 0.999 0.312 C_pine_nut_shell_H_3PO_4 25 29.506 38.957 0.999 0.329 C_pine_nut_shell_H_3PO_4 25 30.406 49.314 0.999 0.329		35	30	.267	21.989	0.998	0.543
C_macauba_CaCl2 25 27.459 21.832 0.999 0.333 35 29.306 26.191 0.999 0.325 45 30.039 28.644 0.999 0.094 25 29.555 38.655 0.999 0.094 35 30.141 36.983 0.999 0.332 C_pine_nut_shell_CaCl2 25 29.629 30.664 0.999 0.326 C_pine_nut_shell_H_3PO_4 25 30.147 35.36 0.999 0.326 C_pine_nut_shell_H_3PO_4 25 29.629 30.664 0.999 0.326 C_pine_nut_shell_H_3PO_4 25 30.488 34.108 0.999 0.312 35 30.406 34.108 0.999 0.326		45	30	.431	21.936	0.997	0.607
35 29.306 26.191 0.999 0.325 45 30.039 28.644 0.999 0.155 C_macauba_H_3PO4 25 29.555 38.655 0.999 0.032 5 30.141 36.983 0.999 0.326 C_pine_nut_shell_CaCl2 25 29.629 30.664 0.999 0.326 5 30.147 33.536 0.999 0.326 6_pine_nut_shell_H_3PO4 5 30.488 34.108 0.999 0.326 C_pine_nut_shell_H_3PO4 25 29.506 38.957 0.999 0.399 C_pine_nut_shell_H_3PO4 25 30.406 49.314 0.999 0.408	C_macauba_CaCl ₂	25	27	.459	21.832	0.999	0.333
45 30.039 28.644 0.999 0.155 C_macauba_H_3PO4 25 29.555 38.655 0.999 0.094 35 30.141 36.983 0.999 0.339 45 30.333 35.1259 0.999 0.326 C_pine_nut_shell_CaCl2 25 29.629 30.664 0.999 0.288 35 30.147 33.536 0.999 0.312 45 30.488 34.108 0.999 0.326 C_pine_nut_shell_H_3PO_4 25 29.506 38.957 0.999 0.322 35 30.406 49.314 0.999 0.408		35	29	.306	26.191	0.999	0.325
C_macauba_H_3PO_4 25 29.555 38.655 0.999 0.094 35 30.141 36.983 0.999 0.339 45 30.333 35.1259 0.999 0.326 C_pine_nut_shell_CaCl_2 25 29.629 30.664 0.999 0.288 35 30.147 33.536 0.999 0.312 45 30.488 34.108 0.999 0.312 C_pine_nut_shell_H_3PO_4 25 29.506 38.957 0.999 0.392 35 30.406 49.314 0.999 0.408		45	30	.039	28.644	0.999	0.155
35 30.141 36.983 0.999 0.339 45 30.333 35.1259 0.999 0.326 C_pine_nut_shell_CaCl2 25 29.629 30.664 0.999 0.288 35 30.147 33.536 0.999 0.312 45 30.488 34.108 0.999 0.325 C_pine_nut_shell_H ₃ PO ₄ 25 29.506 38.957 0.999 0.329 35 30.406 49.314 0.999 0.408	C_macauba_H ₃ PO ₄	25	29	.555	38.655	0.999	0.094
45 30.333 35.1259 0.999 0.326 C_pine_nut_shell_CaCl2 25 29.629 30.664 0.999 0.288 35 30.147 33.536 0.999 0.312 45 30.488 34.108 0.999 0.359 C_pine_nut_shell_H3PO4 25 29.506 38.957 0.999 0.302 35 30.406 49.314 0.999 0.408		35	30	.141	36.983	0.999	0.339
C_pine_nut_shell_CaCl2 25 29.629 30.664 0.999 0.288 35 30.147 33.536 0.999 0.312 45 30.488 34.108 0.999 0.359 C_pine_nut_shell_H3PO4 25 29.506 38.957 0.999 0.392 35 30.406 49.314 0.999 0.408		45	30	.333	35.1259	0.999	0.326
35 30.147 33.536 0.999 0.312 45 30.488 34.108 0.999 0.359 C_pine_nut_shell_H ₃ PO ₄ 25 29.506 38.957 0.999 0.329 35 30.406 49.314 0.999 0.408	C_pine_nut_shell_CaCl ₂	25	29	.629	30.664	0.999	0.288
45 30.488 34.108 0.999 0.359 C_pine_nut_shell_H3PO4 25 29.506 38.957 0.999 0.392 35 30.406 49.314 0.999 0.408		35	30	.147	33.536	0.999	0.312
C_pine_nut_shell_H ₃ PO ₄ 25 29.506 38.957 0.999 0.392 35 30.406 49.314 0.999 0.408		45	30	.488	34.108	0.999	0.359
35 30.406 49.314 0.999 0.408	C_pine_nut_shell_H ₃ PO ₄	25	29	.506	38.957	0.999	0.392
		35	30	.406	49.314	0.999	0.408
45 <u>31.1571</u> <u>58.790</u> <u>0.999</u> <u>0.393</u>		45	31	.1571	58.790	0.999	0.393

Table 4

Thermodynamic darameters for Kinb adsorbuon for the different activated carbons under study	Thermodynamic	parameters for	RhB adsorption	for the different	activated car	rbons under study.
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Material	T (°C)	Thermodynamic parameters			
		$\Delta G^0 (kJ mol^{-1})$	$\Delta H^0 (kJ mol^{-1})$	ΔS^0 (J mol ⁻¹ K ⁻¹)	r ²
C_carnauba_CaCl ₂	25	-9.069	11.778	69.82	0.963
	35	-9.641			
	45	-10.471			
C_carnauba_H ₃ PO ₄	25	-2.229	0.604	9.509	0.999
	35	-2.325			
	45	-2.419			
C_macauba_CaCl ₂	25	-1.013	28.921	100.965	0.916
	35	-2.506			
	45	-3.011			
C_macauba_H ₃ PO ₄	25	-3.098	7.679	36.244	0.972
	35	-3.532			
	45	-3.819			
C_pine_nut_shell_CaCl ₂	25	-2.449	17.032	65.709	0.903
	35	-3.415			
	45	-3.750			
C_pine_nut_shell_H ₃ PO ₄	25	-3.506	38.916	142.377	0.999
	35	-4.946			
	45	-6.353			

precursors, on the activation agents and on the operational conditions amongst the different studies.

3.8. Thermodynamic parameters

The thermodynamic parameters for the adsorption process have been estimated from the Langmuir constants (mmol/g). The enthalpy change (ΔH^0), the entropy change (ΔS^0) and the change in Gibbs free energy (ΔG^0) for the adsorption process can be calculated from the K_L constant and the temperature (*T*) using the following equations (Singh, 1988):

$$\Delta G^0 = -RT \cdot lnK_L \tag{9}$$

$$\Delta G^0 = \Delta H^0 - T \cdot \Delta S^0 \tag{10}$$

$$lnK_{L} = \left(\Delta S^{0} / RT\right) + \left(\Delta H^{0} / R\right)$$
(11)

where *R* is the ideal gas constant (8.314 \times 10⁻³ kJ/mol K), *T* is the temperature in Kelvin, and *K*_L is the distribution coefficient.

The thermodynamic parameters are summarized in Table 4. It



Fig. 5. Effect of additional native cellulose dose on RhB removal using H_3PO_4 -activated carbons obtained from pine nut shell ([RhB]₀ = 80 mg/L, pH = 7, activated carbon dose = 2 g/L; temperature = 25 °C).

can be observed that all the activated carbons under study have negative ΔG^0 values, so the processes are spontaneous in nature. It is also worth noting that ΔG^0 values become more negative with increasing temperature, which implies that the process is more spontaneous at higher temperatures. ΔH^0 was positive for all coals, indicating that the adsorption process is endothermic, and have values below 40 kJ mol⁻¹, characteristic of a physical-type absorption (Bhatnagar et al., 2009). The maximum monolayer RhB adsorption capacity (q_{max}) increases when the temperature is increased from 25 to 45 °C, further confirming the endothermic nature of the process. The positive value of ΔS^0 suggests increased randomness at the solid/solution interface during the adsorption of RhB in the different activated charcoals.

3.9. Preliminary assessment of the effect of cellulose content on RhB removal

From a comparison of the components for the lignocellulosic materials under study (Table S2) with the textural characteristics of their corresponding activated carbons (Table 1), it can be inferred that the BET surface area and pore volume increase with cellulose content. Cagnon et al. (2009), when using physical activation for the different individual components (cellulose, lignin and hemicellulose), concluded that the activated carbon obtained from cellulose had a higher pore volume (0.32 cm³/g) than those obtained from lignin (0.30 cm³/g) and hemicellulose (0.24 cm³/g). Khezami et al. (2005) also reported a higher mesopore content for the chemical activation (with KOH) of cellulose (0.049 cm³/g) in comparison to lignin (0.032 cm³/g) and hemicellulose (0.048 cm³/g).

Consequently, we have conducted a preliminary study aimed at assessing the impact of cellulose content on the RhB removal by adsorption. Pine nut shell has been selected for these tests, provided that its cellulose content is similar to that of macauba endocarp and lower than that of carnauba leaves (so there is more room for improvement). As indicated in the experimental section (subsection 2.4), the cellulose content was artificially increased prior to activation with H₃PO₄ by adding different amounts of native cellulose (0 wt.%, 20 wt.% and 50 wt.%) to the pine nut shell lignocellulosic material. The respective RhB removal efficiencies are depicted Fig. 5. It can be observed that the RhB removal significantly increases from 78% for the activated carbon obtained from pure pine nut shell to 97% when 50 wt.% of native cellulose was added for the same operational conditions. This improvement, as noted above, would be associated to the higher mesopore ratio for

higher cellulose content, which favors RhB adsorption. A detailed study on the suitability of different combinations of native cellulose with lignocellulosic materials for activated charcoals production will be the objective of a further research.

4. Conclusions

Adsorption studies have been conducted for the different activated carbons obtained from the three selected lignocellulosic waste materials (carnauba palm leaves, macauba endocarp and pine nut shell) to assess their capability to remove Rhodamine B in the aqueous phase. The adsorption process fits a pseudo-second order model, which suggests that the adsorption rate is more dependent on the availability of adsorption sites than on dye concentration, and the equilibrium data fit the Freundlich isotherm model, associated to multilayer adsorption and a heterogeneous energy distribution of the active sites. The adsorptive capacity depends both on the lignocellulosic material used and on the activation procedure, attaining better results for CaCl₂ than H₃PO₄ activation as a consequence of enhanced mesopore formation. From the thermodynamic parameters (ΔH^0 , ΔS^0 and ΔG^0) it can be concluded that the physical adsorption process is endothermic and spontaneous for all the activated carbons, so they can be regarded as promising adsorbents for pollutants removal from aqueous solutions. Finally, the preliminary study on the impact of cellulose content (by adding native cellulose to the precursor) shows that its increase leads to an enhancement of the adsorption properties, associated to improved textural properties.

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Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.jenvman.2015.03.007.

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