

1 **Effect of different extrusion treatments and particle size distribution on the physico-**  
2 **chemical properties of rice flour**

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18 **Abstract**

19 Rice flour is an interesting alternative for developing gluten free products, but its features do not  
20 meet the process requirements. The objective of this study was to modify the functional  
21 properties of rice flour by combining extrusion and size fractionation. Different extrusion  
22 conditions were applied to vary the severity of the treatment on the flour constituents. Extrusion  
23 and mechanical fractionation of the rice flours modified their behavior affecting hydration,  
24 thermal and pasting features, besides their susceptibility to enzymatic hydrolysis. Thermal  
25 properties (temperature and enthalpy) increased with the intensity of the extrusion and that effect  
26 was intensified with the greatest particle size of the flours. Fine flours with stronger extrusion  
27 showed the highest susceptibility to enzymatic hydrolysis and extrusion process increased that  
28 effect. Overall the combination of both physical treatment maybe an attractive alternative for  
29 obtaining clean label rice flours with modified features.

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32 **Keywords:** extrusion, rice flour, particle size, thermal properties, hydration, enzymatic  
33 hydrolysis.

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36 **1. Introduction**

37 Lately, there is an increasing interest for gluten free products that has prompted extensive  
38 research focused on improving the quality of gluten free products. The vast majority of studies  
39 have been centered in the substitution of wheat flour for different blends of gluten free flours,  
40 starches, protein isolates, hydrocolloids (Houben, Hochstotter, & Becker, 2012) and the  
41 enzymatic improvement of those formulations (Rosell, 2009). Nevertheless, the potential of the  
42 flours from gluten free cereals has been scarcely exploited. Physical treatments have the benefit  
43 over the chemical ones of changing starch functionalities keeping the Green label (Jacobs &  
44 Delcour, 1998).

45 Rice flour functional properties are fully dependent on genotype and environmental conditions  
46 (Yeh, 2004) and besides that, postharvest treatments could be an alternative for modulating flour  
47 functional features. It is well known that rice grinding significantly affects rice flour properties,  
48 like water binding capacity and swelling power (Perdon, Siebenmorgen, Mauromoustakos,  
49 Griffin, & Johnson, 2001). Recently (de la Hera, Gómez & Rosell, 2013a) showed that particle  
50 size fractionation of rice flour might be advisable for selecting specific physico-chemical  
51 properties like different hydration properties and enzymatic starch hydrolysis. Moreover, those  
52 fractionated flours showed different processing behaviour more suitable for bread or cake  
53 making depending on the particle size (de la Hera, Martínez & Gómez, 2013b; de la Hera,  
54 Martínez, Oliete & Gómez, In press).

55 Thermal treatments are highly attractive to modify the functional properties of the cereal flours.  
56 Extrusion cooking is considered high-temperature-short-time (HTST) during which flours are  
57 submitted to high temperatures and mechanical shearing at relatively low levels of moisture  
58 content (Camire, Camire, & Krumhar, 1990). This treatment allows starch pregelatinization,

59 denaturation of protein, enzyme (in)activation, and Maillard reactions, the extent of which are  
60 dependent on the severity of the extrusion. Those changes at the constituents' level modify the  
61 rheological behavior of flour (Hagenimana, Ding, & Fang, 2006). During extrusion, the starch  
62 properties are dependent on the temperature, initial moisture content and the screw speed (Wen,  
63 Rodis, & Wasserman, 1990). Raising the intensity of the treatment is possible to break down the  
64 amylopectin chains (Mercier & Feillet, 1975). In fact, (Colonna, Doublier, Melcion,  
65 Demonredon, & Mercier, 1984) described that extruded wheat starches have amylose and  
66 amylopectin chains of lower molecular weight than the ones obtained by drum drying due to the  
67 shear effect, and that gave low thickening ability at low temperature (Doublier, Colonna, &  
68 Mercier, 1986).

69 The extrusion also promotes important nutritional changes in the flours, like increase in the  
70 soluble fiber content and reduction in the lipid oxidation tendency, the content of antinutritional  
71 factors and the microbial population (Camire et al, 1990). Besides, it could be obtained an  
72 increase in the content of resistant starch in rice flours (Hagenimana et al, 2006), which is  
73 dependent on the treatment intensity, and it is always higher than the one obtained by other  
74 thermal treatments (Alsaffar, 2011). Extrusion cooking is responsible for gelatinization and  
75 degradation of starch and also for changing the extent of molecular associations between  
76 components, e.g. the amylose– lipid complex that can affect the *in vitro* starch digestibility of  
77 the flours (Hagenimana et al, 2006).

78 Despite the impact of the extrusion on the molecular level, little attention has been paid to the  
79 variation of the functional properties of the flours by hydrothermal treatments (Clerici, Arioldi,  
80 & El-Dash, 2009), even though physically modified flours are considered to be natural materials  
81 with high safety (Jacobs et al, 1998). In fact, Clerici et al (2009) included 10% of extruded acid-

82 modified rice flours for making gluten free breads. When using rice flours extruded in the  
83 presence of different amount of lactic acid, gluten free breads presented crust and crumb colour  
84 and texture values similar to those of wheat bread, although specific volume was rather low.  
85 Considering the influence of the flour fractionation on the functional properties of the rice flours  
86 (de la Hera et al, 2013b), and the molecular changes induced by extrusion cooking, the  
87 combination of both physical treatments could modify rice flour functional properties keeping  
88 the green label. The aim of this study was to modify the functional properties of rice flour by  
89 combining extrusion and size fractionation. With that purpose, different extrusion conditions  
90 were applied to vary the severity of the treatment on the flour constituents. The impact of  
91 processing on the flours was also followed by assessing the susceptibility of the flours to  
92 enzymatic hydrolysis.

93

## 94 **2. Materials and methods**

### 95 **2.1 Materials**

96 Rice flours were provided by Harinera Los Pisones (Zamora, Spain) that carried out the  
97 extrusion treatment in a single screw extruder Bühler Basf (Bühler S.A., Uzwil, Switzerland).  
98 The length to diameter (L/D) ratio for the extruder was 20:1. Rice flour was subjected to  
99 different extrusion intensities (barrel temperature and moisture content of the mass feed)  
100 yielding three types of extruded flours (1-3). Rice flour 1 and 2 were extruded at a maximum  
101 barrel temperature of 110°C with a feed rate of 700kg/h. For flours 1 and 2 feed moisture  
102 content and screw speed was 17% and 30%, and 453rpm and 397rpm, respectively. The  
103 diameter of the die hole used in those flours was 8mm. Rice flour 3 was extruded at a maximum  
104 barrel temperature of 140°C with a feed-rate of 500kg/h and feed moisture content of 25%. The

105 screw speed was 340rpm and the diameter of the die hole was 6 mm. The same rice flour (rice  
106 flour 0) without any treatment was used as a control.

107 Extruded product was dried by convection air and then ground with a compression roller till  
108 particle size was lower than 200 microns. Ground extrudates were sifted in a Bühler MLI 300B  
109 (Bühler AG, Uzwil, Switzerland) with screens of 132 and 200 microns to obtain fine (f) – lower  
110 than 132 $\mu$ m- and coarse (c) – 132 $\mu$ m-200  $\mu$ m- extruded flours.

111 Flours were stored in air-tight plastic containers and held at 4°C until analysis.

112

## 113 **2.2 Methods**

### 114 **2.2.1. Flours characterization**

115 Flours were analyzed following AACC method (AACC, 2012) for protein (AACC, 46-30.01)  
116 with a Leco TruSpec device (Leco, St. Joseph, MI, USA). The particle size distribution was  
117 measured using a particle size analyzer with laser diffraction Helos & Rodos (Sympatec,  
118 Clausthal-Zellerfeld, Germany) following AACC method (AACC, 55-40.01). Determinations  
119 were carried out in duplicate.

### 120 **Free sugars**

121 The glucose content was measured using a glucose oxidase-peroxidase (GOPOD) kit. The  
122 absorbance was measured using an Epoch microplate reader (BIOTEK EPOCH, Izasa,  
123 Barcelona, Spain) at 510 nm. In all cases four replicates were assayed for each experimental  
124 point.

### 125 **Damage starch**

126 The content of damaged starch was determined according to AACC 76-30A method (AACC,  
127 2012). A fungal enzyme from *Aspergillus oryzae* (A6211, Sigma Chemical Co., St. Louis, MO,

128 USA) was used in that analysis. Three determinations were made for each sample. Damaged  
129 starch was expressed as percentage of flour weight on dry basis.

### 130 **Hydration properties**

131 Hydration properties included swelling and water binding capacity (WBC) (Nelson, 2001).  
132 Swelling volume or the volume occupied by a known weight of flour was evaluated by mixing  
133 5g ( $\pm 0.1$ mg) of flour with 100ml distilled water and allowing it to hydrate during 16h.  
134 Water binding capacity defined as the amount of water retained by the flour after it has been  
135 subjected to centrifugation was measured as described the method 56.30 (AACC, 2012).  
136 Determinations were carried out in duplicate.

### 137 **Emulsifying properties**

138 Flour suspension (360 mL) of 0.5% (w/v) starch concentration was mixed with commercial  
139 sunflower oil (Langosta, F. Faiges S.L, Daimiel, Ciudad Real, Spain) (36 mL). The content was  
140 stirred for one min with a beater (Taurus Bapi 350 CP/CM, Taurus, Oliana, Lérida, Spain) to  
141 disperse the sample in the oil. The suspensions were then centrifuged at 800xg for 10 min. The  
142 emulsifying capacity (EC) was calculated as:

$$143 \quad EC = (ev/tv) * 100 \quad (\text{Eq. 2})$$

144 where  $ev$  is the emulsion volume and  $tv$  is total volume.

145 Emulsion stability (ES) against high temperatures, were determined in the emulsions that were  
146 heated in a water bath at 80°C for 30 min, and centrifuged at 800xg for 10 min. ES was  
147 calculated as:

$$148 \quad ES = (fev/iev) * 100 \quad (\text{Eq. 3})$$

149

150 where *fev* is the final emulsion volume and *iev* is initial emulsion volume. Determinations were  
151 carried out in duplicate.

### 152 **Foaming properties**

153 Aliquots (150mL) of 4% w/v suspension were whipped at moderate speed for one min using a  
154 beater (Taurus Bapi 350 CP/CM, Taurus, Oliana, Lérida, Spain). Foam volumes were recorded  
155 after 30 s. The foam capacity (FC) was calculated as follows:

$$156 \quad FC = (ifv / tsv) * 100 \quad (\text{Eq. 4})$$

157 where *ifv* is the initial foam volume and *tsv* is the total suspension volume.

158 The foam stability (FS) was calculated as the foam volume after 20 min.

$$159 \quad FS = (ffv / tsv) * 100 \quad (\text{Eq. 5})$$

160 where *ffv* is the foam volume after 20 min and *tsv* is total suspension volume. Results were the  
161 average of two determinations.

### 162 **Pasting characteristics**

163 Pasting properties of flours were analyzed using the standard method (AACC, 2012), (AACC,  
164 61-02.01) with a Rapid Visco Analyser (RVA-4) (Newport Scientific Pty Ltd., Warriewood,  
165 Australia) controlled by Thermocline software (Newport Scientific Pty. Limited, Warriewood,  
166 Australia) for Windows.

### 167 **Thermal properties**

168 Analyses were performed in a differential scanning calorimeter DSC-7 (Perkin-Elmer,  
169 Waltham, MA, USA), using aluminum pans (PE 0219-0062). The equipment was calibrated  
170 with Indium and an empty pan was used as a reference. Flour (3 mg) was loaded into the  
171 aluminum pan and distilled water (10 $\mu$ L) was added with the help of a Hamilton micro syringe.  
172 Samples were hermetically sealed and allowed to stand for 1 h at room temperature before



173 heating in the DSC. The calorimeter scan conditions were set as follows: samples were kept at  
174 30°C for 2 min, heated from 30 to 110°C at 5°C/min. Onset temperature ( $T_o$ ), peak temperature  
175 ( $T_p$ ), gelatinization temperature range ( $T_p - T_o$ ), peak height index ( $\Delta H_g / T_p - T_o$ ) as well as the  
176 enthalpy of starch gelatinization ( $\Delta H_g$ ) (expressed as mJ/mg of sample) were determined. All  
177 samples were run in quadruplicate.

### 178 **Colour of flours**

179 Colour was measured using a Minolta CN-508i spectrophotometer (Minolta, Co.LTD, Tokyo,  
180 Japan) with the D65 standard illuminant and the 2° standard observer. Results were expressed in  
181 the CIELab colour space. Colour determinations were made 5×2 times on each sample of flour.

### 182 **Enzymatic hydrolysis of starch**

183 Starch hydrolysis was measured following the method described by Gularte and Rosell (2011)  
184 with minor modifications. Briefly, for free sugars removal, flour sample (100 mg) suspended in  
185 two milliliters of 80% ethanol was kept in a shaking water bath at 85°C for five minutes, and  
186 then centrifuged for 10 min at 1000×g. The pellet was incubated with porcine pancreatic  $\alpha$ -  
187 amylase (10 mg/ml) (Type VI-B,  $\geq 10$  units/mg solid, Sigma Chemical, St. Louis, MO, USA)  
188 and amyloglucosidase (3300 U/ml) (Sigma Chemical, St. Louis, MO, USA) in 10 ml of 0.1M  
189 sodium maleate buffer (pH 6.0) in a shaking water bath at 37 °C (0.25–16 h). Aliquots of 200  $\mu$ l  
190 were withdrawn during the incubation period. Aliquots were mixed with 200  $\mu$ l of ethanol  
191 (96%) to stop the enzymatic reaction and the sample was centrifuged for 5 min at 10000×g and  
192 4 °C. The precipitate was washed twice with 50% ethanol (100  $\mu$ l) and the supernatants were  
193 pooled together and kept at 4 °C for further glucose determination.

194 The remnant starch after 16 h hydrolysis was solubilized with 2ml of 2M KOH using a Polytron  
195 ultraturrax homogenizer IKA-T18 (IKA works, Wilmington, NC, USA) during 1min at speed 3.

196 The homogenate was diluted with 8ml 1.2M sodium acetate pH 3.8 and incubated with 100µl  
197 amyloglucosidase (3300 U) at 50 °C for 30 min in a shaking water bath. After centrifuging at  
198 2000×g for 10 min, supernatant was kept for glucose determination.

199 The glucose content was measured using a glucose oxidase-peroxidase (GOPOD) kit. The  
200 absorbance was measured using an Epoch microplate reader (Biotek Instruments, Winooski, VT,  
201 USA) at 510 nm. Starch was calculated as glucose (mg)×0.9. Replicates (n=2-4) were carried  
202 out for each determination.

203 Experimental data were fitted to a first-order equation (Goñi et al., 1997):

$$204 C_t = C_\infty (1 - e^{-kt}) \quad \text{Eq. 5}$$

205 Where  $C_t$  is the concentration of product at time  $t$ ,  $C_\infty$  is the concentration at the end point, and  $k$   
206 is the pseudo-first order rate constant. Although this equation requires the estimation of an  
207 accurate  $C_\infty$ , it was useful because long reaction times were applied to determine resistant starch  
208 after complete enzymatic hydrolysis. The plot of  $\ln [(C_\infty - C_t) / C_\infty] = -kt$  against  $t$  was used to  
209 estimate the slope that corresponded to  $-k$ .

210 However, as recently suggested Butterworth, Warren, Grassby, Patel and Ellis (2012), the linear  
211 plot of  $\ln (dC/dt)$  against  $t$  was also represented to calculate the slope ( $-k$ ), and the intercept on  
212 the  $y$  axis was used for calculating the  $\ln(k C_\infty)$ . This plot was used to demonstrate if the data  
213 were of logarithmic form and the rate constant remained unchanged along the whole hydrolysis  
214 reaction, as recommended Poulsen, Ruitter, Wisser, Jorgen and Iversen (2003).

### 215 **2.2.2. Statistical analysis**

216 Multiple analyses of variance were used to determine the individual effects of thermal treatment  
217 and particle size of flours. Fisher's least significant differences test was used to calculate the  
218 means with their 95% confidence intervals. Several correlations were also run. The statistical

219 analysis was performed with the Statgraphics Plus Centurion XVI software (Statpoint  
220 Technologies, Inc., Warrenton, VA, USA).

221

### 222 **3. Results and Discussion**

223 Rice flour was subjected to different extrusion treatments that differed on the maximum barrel  
224 temperature and feed moisture content in order to obtain different extrusion intensities. In  
225 addition, resulting extruded flours after grinding were separated in two fractions depending on  
226 their particle size obtaining coarse extruded flour (132 $\mu$ m-200 $\mu$ m) and fine extruded flour  
227 (<132 $\mu$ m). Overall eight samples were obtained from each batch, which differed on the level of  
228 extrusion (identified as 1-3) and the particle size (coarse, fine).

229

#### 230 **3.1 Damage starch and free sugars**

231 To get a complete picture of the effect of extrusion and particle size a multiple analysis of  
232 variance was applied to the experimental results (Table 1). The extrusion intensity and particle  
233 size had a significant effect on the content of free sugars, which increased with the extrusion  
234 intensity and with the reduction of the particle size. Nevertheless, no significant differences were  
235 observed between the free sugars content of the control and the mild extrusion treatment (flour  
236 1). Thus the hydrolysis responsible of the sugar release required a minimum barrel temperature  
237 and also sufficient feed moisture content, since flours 1 and 2 were extruded at the same  
238 temperature and with different moisture feeding. Extrusion induced a progressive increase of the  
239 damage starch content with the intensity raise, likely due to damage produced by the shears  
240 force and the heat during extrusion (Camire et al, 1990). Conversely, damage starch decreased  
241 with the particle size, showing coarser flours the greatest amount of damage starch, which

242 agrees with the trend observed by de la Hera et al. (2013a), when studying the features of  
243 different particle size fractions of rice flours.

244

### 245 **3.2 Hydration, emulsifying and foaming properties**

246 Hydration, emulsifying and foaming properties were significantly affected by the extrusion  
247 process and the particle size of the flours (Table 1). Hydrations properties (WBC and swelling)  
248 increased with the extrusion intensity and also with the particle size of the flour. Those effects  
249 were partially attributed to the increase in the amount of damage starch since it was found a  
250 positive correlation between the amount of damage starch and WBC ( $r=0.88$ ) and with the  
251 swelling ( $r=0.88$ ). Moreover, the cooking produced during extrusion led to gelatinized starch  
252 that would have higher WBC and swelling, as occurred with the water absorption index  
253 (Hagenimana et al, 2006). Camire et al. (1990) proposed that the breakage of the starch granule  
254 integrity led to a poorly ordered molecular phase with hydroxyl groups prone to bind water  
255 molecules.

256 The extrusion significantly reduced the emulsifying capacity of the flours, with the exception of  
257 flour 3, compared to the control flour, and an increase of the EC was observed with the severity  
258 of the extrusion (flour 3). That effect must result from the protein and starch changes during  
259 extrusion process. Considering the proteins, extrusion forces the unfolding and aggregation due  
260 to protein crosslinking involving SH/SS interchange, oxidation and hydrophobic interactions  
261 (Rosell & Foegeding, 2007), which might result in a decrease of the EC of the flour. As the  
262 extrusion intensity increase, starch modification might partially mask the consequence of protein  
263 denaturation and EC increase due to gelatinized starch has greater number of hydroxyl groups  
264 available to form hydrogen bonds with the proteins leading to better emulsion capacity.

265 Emulsion stabilities were higher in the flours obtained from lower extrusion intensities (flour 1  
266 and 2), likely the denaturation of the rice proteins during extrusion increased the stability. The  
267 particle size of the rice flours did not affect significantly the EC, but the ES significantly  
268 increased with the particle size. A reduction in the particle size of the flour improves the  
269 emulsifying properties of the flours (Aluko, Mofolasayo, & Watts, 2009), but it seems that some  
270 time is needed for displaying that effect since only ES was affected by the particle size. In the  
271 present study, it was observed a significant positive relationship between the ES and the free  
272 sugars content ( $r=0.93$ ,  $P<0.001$ ), which could reduce the total charge of the proteins leading the  
273 formation of interfacial protein membranes that stabilize the emulsion (Aluko et al, 2009).

274 Extrusion improved the foaming capacity of the rice flours, but there was no trend with the  
275 extrusion severity. The foam stability could be only measured in flour 3 (50.00 for fine flour and  
276 45.18 for coarse flour), because very unstable foams were obtained with the other flours. The FC  
277 has been attributed to its microstructure, size and distribution of the gas cells and the interfacial  
278 properties (Zhang, Bai, & Zhang, 2011). Hydrothermal treatments can improve the foaming  
279 properties, like it has been reported with corn kernels (Boladea, Usman, Rasheed, Benson, &  
280 Salifou, 2002). Nevertheless, the minor effect observed in the extruded rice flours could be  
281 attributed to the low protein content of the rice flour, since usually protein isolates show great  
282 foaming capacity that improves with the hydrothermal treatments (Wang & Johnson, 2001). The  
283 effect of extrusion on foam formation followed an opposite trend to the one observed on the  
284 emulsion formation, which suggests that different mechanisms are involved during interfacial  
285 membrane formation at the air-water and oil-water interfaces. Concerning the particle size, the  
286 major FC was observed in fine flours, which was explained by the greater availability of  
287 lowering interfacial components in those flour fractions, as proposed Aluko et al. (2009).

288

289 **3.3. Colour**

290 Luminosity ( $L^*$ ) of the flours significantly decreased with the extrusion intensity (Table 1), and  
291 increased the  $a^*$  and  $b^*$ . Nevertheless, whereas the effect was steady with the extrusion severity  
292 in the case of the luminosity and  $b^*$ , no trend was observed in the case of  $a^*$ . The extrusion  
293 process could lead to Maillard reactions and a reduction of the lipids oxidation due to enzymes  
294 inactivation that induces the formation of melanoidins and the pigments protection, which in  
295 turn produces the modification of the flours color (Camire et al, 1990). Moreover, higher  $L^*$  and  
296 lower  $a^*$  y  $b^*$  were obtained in the fine flours. It has been proposed that fine flours had higher  
297 surface area that favors the contact of the constituents with the oxygen promoting the pigments  
298 oxidation (Atwell, 2001).

299

300 **3.4. Pasting characteristics**

301 Pasting plots of the extruded flours are displayed in Figure 1. When rice flours were suspended  
302 in water, the major viscosity (initial viscosity at 50°C) was observed in the flour 3 at both  
303 particle sizes, due to the high amount of damage starch content and pregelatinized starch (Chao-  
304 Chi Chuang & Yeh, 2004). Concerning the extrusion treatment, the viscosity during heating and  
305 cooling decreased with the extrusion intensity, obtaining minimum viscosity in flour 3 (with the  
306 highest intensity). Nevertheless, when particle size was taken into account, a progressive  
307 decrease of viscosity was observed in the coarse flours, but fine flours from treatment 1 and 2  
308 did not show any difference in the pasting profile (Figure 1 b). Since flour 1 and 2 were treated  
309 at maximum barrel temperature of 110°C and the unique variation was the feed moisture

310 content, it seems that the effect on pasting profiles was independent on the feed moisture content  
311 during extrusion when small particle size were subjected to this treatment.

312 Peak viscosity was significantly dependent on the damage starch content ( $r=-0.79$ ), which agree  
313 with previous studies connecting peak viscosity with gelatinized and damage starch that was  
314 related to the polymerization degree of the starch granules (Barres, Verges, Tayeb, & Della  
315 Valle, 1990). The reduction observed in the final viscosity and setback was displaying the  
316 extension of the effect on the amylose chains, which might lose the ability to retrograde during  
317 cooling due to their fragmentation during extrusion. This effect agrees with previous results of  
318 Doublier et al. (1986).

319

### 320 **3.5. Differential Scanning Calorimetry (DSC)**

321 The effect of extrusion treatment and particle size on the thermal properties of the starch is  
322 shown in Table 2. In the range of temperature tested, flours exhibited one endothermic peak  
323 corresponding to amylopectin gelatinization, with the exception of flour 3. The absence of an  
324 endothermic peak for flours 3 indicated total gelatinization of amylopectin. Indeed, these results  
325 agree with those previously discussed regarding the very small pasting curve observed in the  
326 extruded flours 3. The extrusion treatment significantly modified the gelatinization temperatures  
327 of the flours, and those temperatures were also dependent on the particle size of the flours.  
328 Gelatinization temperatures were sifted to higher values when flours were treated at increasing  
329 extrusion intensity, but the temperature range was not affected. Higher gelatinization  
330 temperature indicated that more energy is required to initiate gelatinization of the starch  
331 suggesting that extrusion is affecting the outer and more amorphous part of the granule and is

332 progressing to the core of the granule till no crystalline structure is left for gelatinization (flour  
333 3).

334 When comparing extruded flours, the gelatinization enthalpy was significantly reduced due to  
335 the intensity of the extrusion, which was expected since extrusion induces starch gelatinization  
336 and an increase of the damage starch content (Chiu & Solarek, 2009), leading to a reduction of  
337 the native starch granules able to gelatinize (Biliaderis, Page, Maurice, & Juliano, 1986).  
338 Nevertheless, the mild extrusion treatment (flour 1) gave significantly higher gelatinization  
339 enthalpy compared to the untreated flour. Taking into account that treatment 1 was applied using  
340 lower feed moisture content (insufficient to complete gelatinization), the higher enthalpy of this  
341 sample could be attributed to a reorientation of the structure of the amorphous region to  
342 resemble that of the crystalline region (Camire et al, 1990). The extrusion process modifies the  
343 crystalline structure of the starch granule affecting the temperature at which swelling starts  
344 (Camire et al, 1990). Fine flours showed lower gelatinization temperatures than the  
345 corresponding coarse flours, but without affecting the peak high index and the gelatinization  
346 enthalpy (Table 2).

347

### 348 **3.6. Starch hydrolysis**

349 The susceptibility of the extruded flours to the enzymatic hydrolysis was analyzed. Figure 2  
350 shows the kinetic plots of the extruded flours and the effect of the particle size. The enzymatic  
351 hydrolysis profiles were dependent on the particle size, and fine flours showed faster hydrolysis  
352 and reached higher asymptotic values than course flours. de la Hera et al. (2013) observed lower  
353 hydrolysis rate in the coarse flours when studied the effect of particle size distribution on the  
354 rice flour functionality. This result could be attributed to the high surface area of the fine flours



355 that increase the water diffusion and enzyme accessibility. The hydrolysis curves were fitted to a  
356 first order kinetics according to Goñi, García-Alonso and Saura-Calixto (1997) and also to  
357 Butterworth, Warren, Grassby, Patel & Ellis (2012) to obtain the kinetic parameters (Table 3).  
358 As it was observed in the plots, there was an increase in the equilibrium concentration ( $C_{\infty}$ )  
359 parallel to the extrusion intensity (except flour 2f). Regarding the rate of the hydrolysis,  $k$ , there  
360 was no general trend with the extrusion intensity. There was great agreement with the  
361 equilibrium concentration estimated from both fitted methods, indicating that the kinetic  
362 parameters can be fitted to a logarithmic function and that the rate constant did not vary along  
363 the hydrolysis reaction (Poulsen, Ruitter, Visser, & Iversen, 2003).

364 Resistant starch was also quantified to determine the potential impact of the extrusion on the  
365 structural level of starch. Although there was no clear tendency about the resistant starch  
366 content, the highest extrusion intensity gave the flours with the lower level of RS (flour 3). This  
367 finding disagrees with previous observations of Hagenimana et al. (2006), who found an  
368 increase in RS content with the treatment severity. Those authors attributed the increase in RS to  
369 the formation of amylose-lipid complexes during the extrusion, which retarded the enzymatic  
370 digestion. Therefore, results divergence might be explained because of the lower content of  
371 amylose of the flours in the present study compared with the reported ones. In addition  
372 Chinnaswamy & Hannah (1990) reported a change in the percentage of amylose/amylopectin  
373 ratio in extruded corn flours that was ascribed to both chains fragmentation, being more intense  
374 in the former. That fact could affect the starch hydrolysis rate. Moreover, Hagenimana et al.  
375 (2006) stated that the susceptibility of the extruded starches to be enzymatically hydrolyzed was  
376 directly related to the intensity of the extrusion treatment.

377 The particle size did not significantly affect the hydrolysis rate, but fine flours showed higher  
378 values of  $C_{\infty}$  and lower amount in RS. Al-Rabadi, Torley, Williamsa, Brydena, & Gidley (2011)  
379 stated that fine extruded flours of barley and sorghum had major digestibility than the coarser  
380 ones. The most compact structure, besides the smaller surface area of coarser flours, could  
381 hinder the accessibility of the enzymes within the starch structure, since diffusion of the enzyme  
382 is the first stage in the enzymatic hydrolysis (Al-Rabadi et al. 2011; Ghaid, Al-Rabadi, Gilbert,  
383 & Gidley, 2009).

384

#### 385 **4. Conclusion**

386 Extrusion and mechanical fractionation of the rice flours modified their behavior affecting  
387 hydration, thermal and pasting features, besides their susceptibility to enzymatic hydrolysis. The  
388 severity of the extrusion treatment was accompanied by an increase in the amount of damage  
389 starch and free sugars content, the former contributing to the Maillard reaction, which affected  
390 the luminosity of the flours. In parallel, hydration ability increased with the extrusion intensity,  
391 leading higher viscosity in cold solution, which might be very interesting for some food  
392 applications. Thermal properties (temperature and enthalpy) increased with the intensity of the  
393 extrusion and that effect was intensified with the greatest particle size of the flours. Fine flours  
394 with stronger extrusion showed the highest susceptibility to enzymatic hydrolysis and extrusion  
395 process increased that effect.

396

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402

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487

488

489 **Table 1:** Significant individual effects of extrusion treatment (1-3) and particle size (coarse,  
 490 fine) on free sugars, damaged starch, emulsifying, foaming and colorimetric properties of rice  
 491 flours.

	Media	SE	Extrusion treatment				Particle size	
			3	2	1	0	c	f
Free Sugars (%)	21.47	4.57	49.91c	19.58b	9.49a	6.92a***	18.74a	24.21b**
Damaged Starch (%)	18.08	2.83	32.49d	23.33c	9.85b	6.66a***	19.98b	16.18a**
WBC (g water/g solid)	2.42	0.39	4.94d	2.13c	1.35b	1.26a***	2.46b	2.39a*
Swelling (mL/g)	3.50	0.79	8.60d	2.90c	1.55b	0.95a***	3.85b	3.15a**
EC	86.21	0.34	86.34bc	85.70ab	84.92a	87.58c**	86.60a	85.82a
ES	112.21	0.52	110.90a	113.23b	113.20b	111.49a***	110.80a	113.61b***
FC	24.85	1.61	30.04c	21.44b	29.97c	17.94a***	22.92a	26.77b***
<i>L*</i>	91.0	0.67	88.62a	90.43b	90.55b	94.36c***	89.66a	92.31b***
<i>a*</i>	0.19	0.08	0.55d	0.16b	0.23c	-0.17a***	0.23b	0.15a***
<i>b*</i>	9.63	0.55	11.13c	10.37b	10.22b	6.82a***	10.83b	8.44a***

492 Particle size: coarse (c), fine (f). \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ .

493 WBC, water binding capacity; EC, emulsifying capacity; ES, emulsion stability; FC, foaming  
 494 capacity.

495

496 **Table 2.** Significant individual effects (extrusion treatment and particle size) on thermal  
 497 properties.

	Media	SE	Extrusion treatment				Particle Size	
			3	2	1	0	c	f
T <sub>o</sub> (°C)	68.0	1.2	n.d	71.1c	67.4b	65.5a***	70.3b	65.7a***
T <sub>p</sub> (°C)	74.0	1.0	n.d.	76.4c	74.4b	71.2a***	75.6b	72.4a***
T <sub>c</sub> (°C)	80.6	0.9	n.d.	82.5b	81.4b	77.9a**	81.8b	79.3a**
T <sub>p</sub> -T <sub>o</sub> (°C)	6.0	0.4		5.3a	7.0a	5.8a	5.3a	6.7a
ΔH (J/g)	2.383	0.229	n.d.	1.550a	3.075c	2.525b***	2.483a	2.283a
PHI								
(J/g*°C)	0.392	0.040	n.d.	0.275a	0.450b	0.450b*	0.433a	0.350a

498

499 Particle size: coarse (c), fine (f). \**P*<0.05; \*\**P*<0.01; \*\*\**P*<0.001.

500 n.d.: Not detected.

501 T<sub>o</sub>, gelatinization onset; T<sub>p</sub>, peak temperature; T<sub>c</sub>, conclusion temperature, T<sub>p</sub>-T<sub>o</sub>, gelatinization

502 range, ΔH, enthalpy and PHI, peak high index.

503



504 **Table 3.** Kinetic parameters extracted from first-order and LOS plots of different flours.

505

	$k$ ( $\text{min}^{-1}$ ) by first order eq.	$k$ ( $\text{min}^{-1}$ ) by LOS	$C_{\infty}$ (%)	$C_{\infty}$ (%) by LOS	Resistant starch (%)
0f	0.043	0.044	137.48	150.63	5.52
0c	0.053	0.051	124.39	130.72	5.91
1f	0.074	0.071	358.42	388.85	3.29
2f	0.061	0.059	338.79	364.30	4.01
3f	0.145	0.143	387.35	727.46	2.23
1c	0.054	0.053	134.98	143.25	6.10
2c	0.076	0.073	143.38	156.65	5.75
3c	0.061	0.059	230.75	243.55	2.11

506

507  $k$ , kinetic constant;  $C_{\infty}$ , equilibrium concentration

508 Numbers in sample codes are referred to extrusion intensity and letters are associated to coarse

509 (c) or fine (f) flour.

510

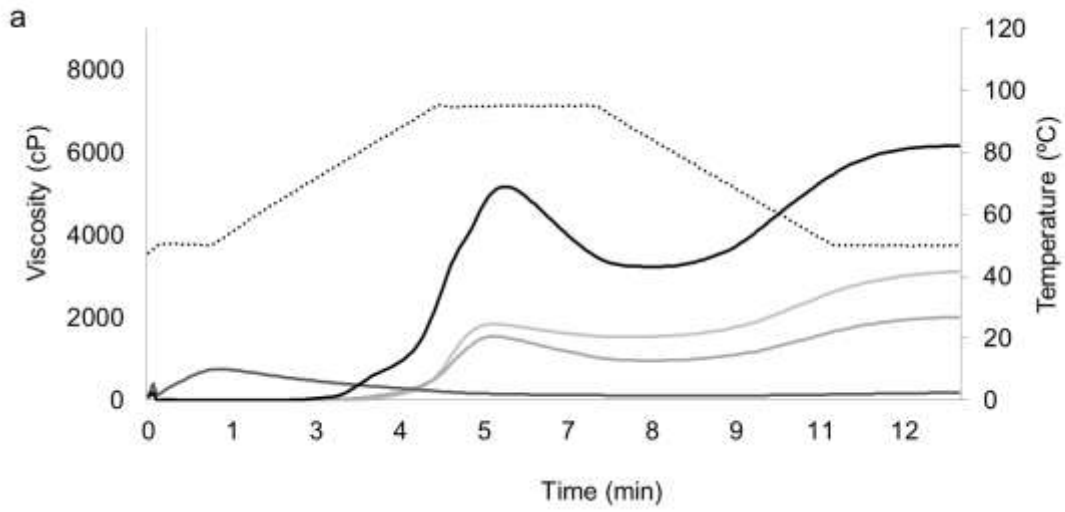
511 **FIGURE CAPTIONS**

512 **Figure 1.** Effect of extrusion treatment on the pasting properties of rice flours with different  
513 particle size. Flour 0 (black line), flour 1 (clear grey line), flour 2 (intermediate tone grey line),  
514 flour 3 (dark grey line). Temperature profile (discontinuous line). Coarse flours (a), fine flours  
515 (b).

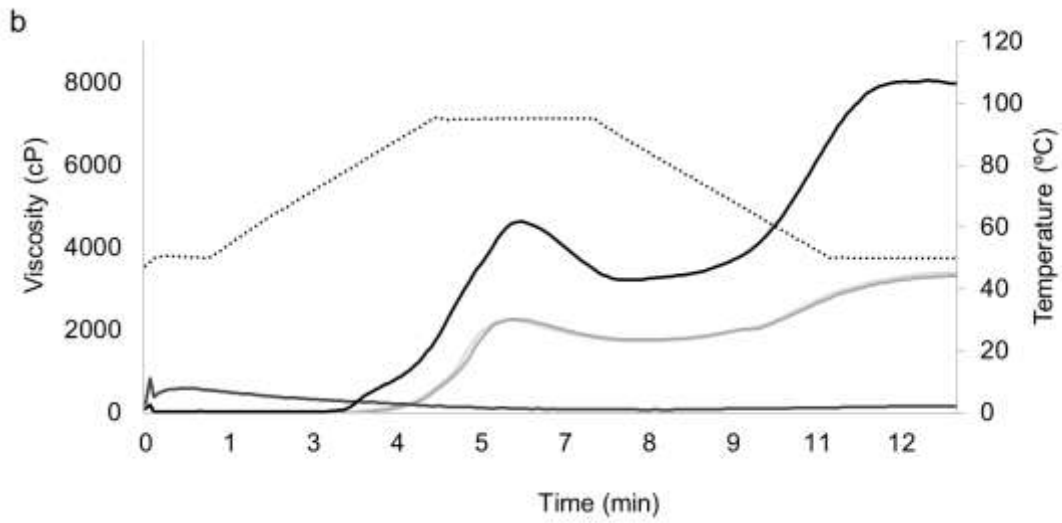
516 **Figure 2.** Effect of extrusion treatment on the enzymatic hydrolysis of rice flours with different  
517 particle size. Flour 0 (black line), flour 1 (clear grey line), flour 2 (intermediate tone grey line),  
518 flour 3 (dark grey line). Coarse flours (a), fine flours (b).

519

520 Figure 1



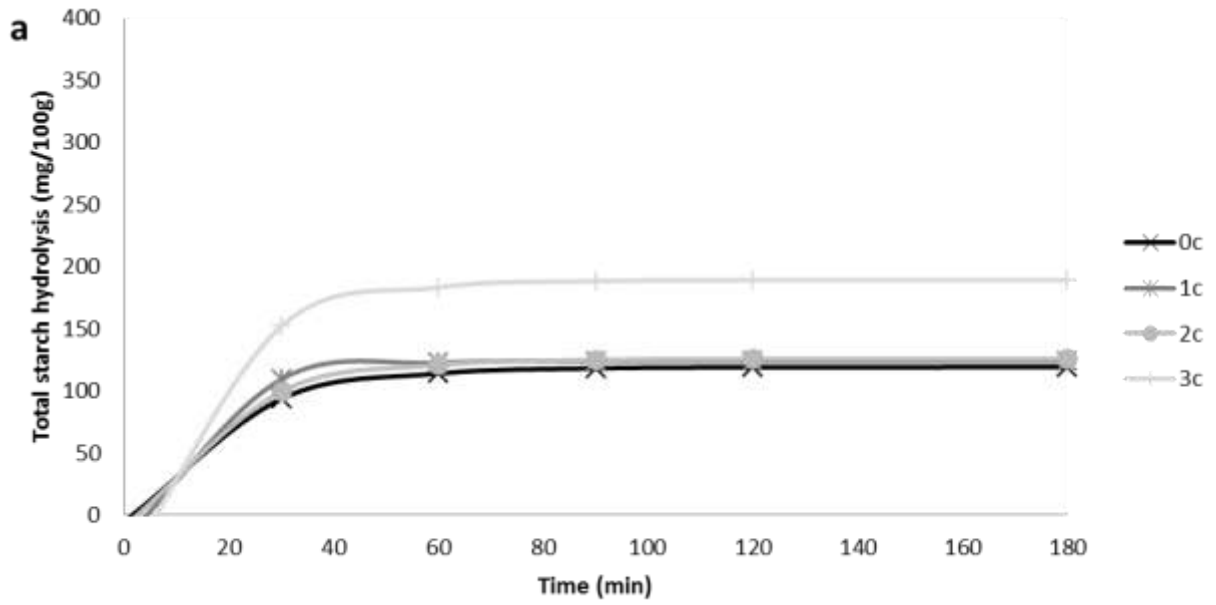
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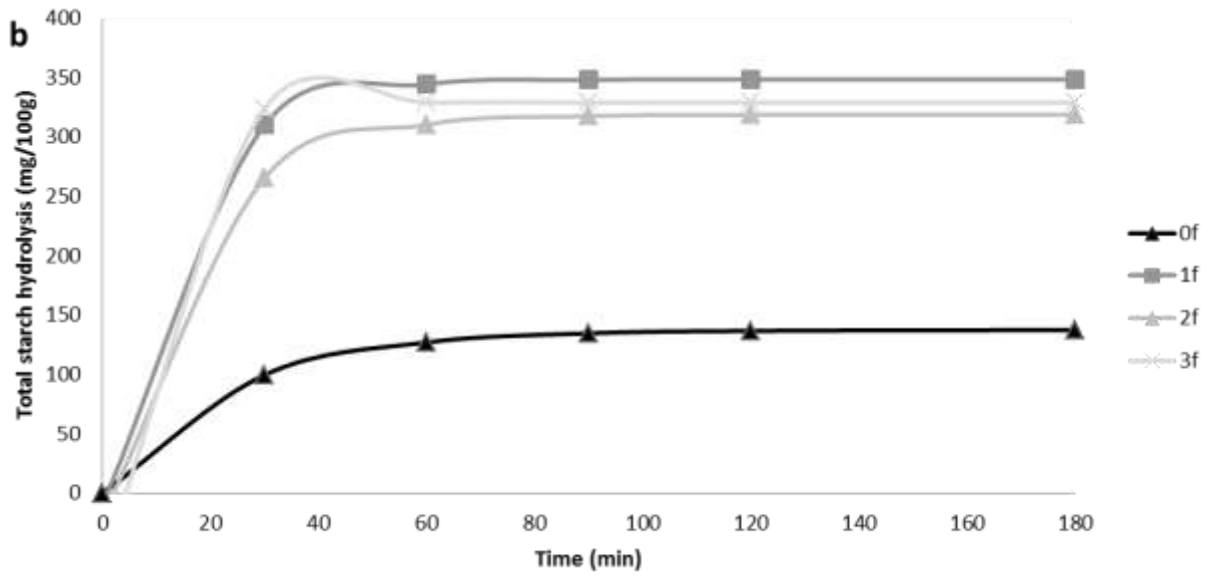
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524 Figure 2



525



526