## Advanced nanomedicine and cancer: challenges and

## opportunities in clinical translation

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### Abstract

Cancer has reached pandemic dimensions in the whole world. Although current medicine offers multiple treatment options against cancer, novel therapeutic strategies are needed due to the low specificity of chemotherapeutic drugs, undesired side effects and the presence of different incurable types of cancer. Among these new strategies, nanomedicine arises as an encouraging approach towards personalized medicine with high potential for present and future cancer patients. Therefore, nanomedicine aims to develop novel tools with wide potential in cancer treatment, imaging or even theranostic purposes. Even though numerous preclinical studies have been published with successful preliminary results, promising nanosystems have to face multiple obstacles before adoption in clinical practice as safe options for patients with cancer. In this MiniReview, we provide a short overview on the latest advances in current nanomedicine approaches, challenges and promising strategies towards more accurate cancer treatment.

**Keywords:** cancer therapy; nanomedicine; nanoparticles; clinical translation chemotherapy;

## **1. Introduction**

Cancer is considered one of the hardest health-related threats worldwide. In fact, cancer provokes 8.97 million deaths thereby being the second leading cause of death only ischemic heart disease, according to the World Health Organization (WHO) (WHO, 2020). Formerly thought of as a single mass, tumors are composed by cancer cells surrounded by non-cancer cells within the extracellular matrix (ECM) such as immune cells, adipocytes and cancer stem cells (CSCs), thereby forming the complex tumor microenvironment (TME) (Hinshaw and Shevde, 2019). Cancer is characterized by aberrant cell proliferation compared to normal cell growth (Krieghoff-Henning *et al.*, 2017). Those cells first affected determine the type of disease, being known more than 100 different types. Healthy cells are not strong enough to compete with cancer cells for nutrients from the bloodstream (DeBerardinis *et al.*, 2016; Vander Heiden *et al.*, 2017; Vazquez *et al.*, 2016). Thus, healthy cells are overcrowded by tumor cells, which are able to widely spread. Since the high amount of nutrients cannot be provided by the vasculature, some cancer cells die but most of them are able to divide in this environment where not all nutrient requirements are fulfilled (Krieghoff-Henning *et al.*, 2017).

The WHO published in 2018 the list of the most frequent cancers through the Global Cancer Observatory (GLOBOCAN) registry (WHO, 2020). Among the new cases of cancer diagnosed in 2018 (18.08 million in total), lung (2.09 million cases together with

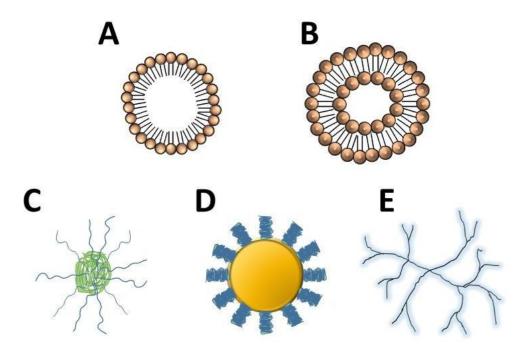
trachea and bronchus), breast (2.09 million cases) and prostate (1.28 million cases) were the most frequent. In men, lung (1.37 million cases) and prostate (1.28 million cases) cancers ranked the first and second positions, followed by stomach and liver cancer (and intrahepatic bile ducts) with 0.68 and 0.60 million cases, respectively. In women, breast cancer is the most frequent with 2.09 million cases followed by lung (0.72 million cases), cervix uteri (0.57 million cases) and colon (0.58 million cases) cancers. However, it is worthy to highlight that colon and rectal cancers (colorectal cancer) would be the third most frequent cancer overall (1.80 million cases).

Although the administration of free chemotherapeutic drugs remains as the gold standard for cancer treatment, this therapeutic strategy still presents inherent challenges. Among the most important are the lack of specific treatments and poor drug accumulation in the tumors (Creixell and Peppas, 2012; Han *et al.*, 2017; Jain and Stylianopoulos, 2010). As a consequence, undesired side effects in healthy tissues occur, especially in the heart (Octavia *et al.*, 2012), bone marrow (Daniel and Crawford, 2006), gastrointestinal tract (Mitchell, 2006), and nervous system (Grothey, 2003).

### 2. Biomaterials and cancer

During the last years, scientific research tries to develop novel biomedical devices in order to improve not only the diagnosis of cancer, but also its treatment (Brouillard *et al.*, 2021; Shi *et al.*, 2017; Wong *et al.*, 2020). In this context, materials' engineering emerges as a new medical technology aimed at the obtention of proper tailor-made biomaterials. According to the definitions of N.A. Peppas and D.F. Williams, a biomaterial is "substance other than food or drugs contained in therapeutic or diagnostic systems that is in contact with tissues or biological fluids" and "a substance that has been engineered to take a form which, alone or as part of a complex system, is used to direct, by control of interactions with components of living systems, the course of any therapeutic or

diagnostic procedure, in human or veterinary medicine" (Peppas and Langer, 1994; Williams, 2009). Biomaterials are therefore materials which are synthesized from a biological source or have the final aim of interacting with the biological system in order to assist in diagnosis, therapy or to restore physiological functionality (Aamodt and Grainger, 2016; Huebsch and Mooney, 2009). Over the years, advanced biomaterials have emerged as one of the most promising tools to increase therapeutic efficiency and biocompatibility, and possess the ability to mimic properties and features found in natural macromolecules in order to be used in multiple biomedical applications, such as drug delivery (Gonzalez-Valdivieso *et al.*, 2019), gene delivery (Piña *et al.*, 2020), nanovaccines (Gonzalez-Valdivieso *et al.*, 2020), *in vivo* imaging (Oliveira *et al.*, 2019), early diagnosis (Beri *et al.*, 2018), tissue engineering (Taraballi *et al.*, 2018), regenerative medicine (Biggs *et al.*, 2018), 3D bioprinting (Murphy and Atala, 2014), biosensors (Tao *et al.*, 2019) or cell harvesting (Pierna *et al.*, 2013).



**Figure 1.** Schematic representation of different structures used in nanomedicine. A: Lipid nanoparticle; B: Liposome; C: Micelle; D: Gold nanoparticle; E: Dendrimer.

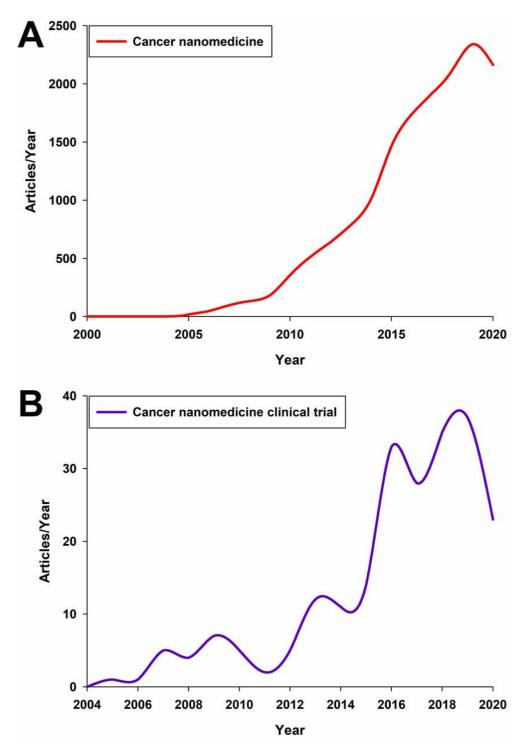
### 3. Nanomedicine

Nanomedicine combines biomaterials, nanotechnology, biomedicine and pharmaceutical science, thereby involving nanopharmaceutics, nanodevices for imaging purposes and theranostics (Eifler and Thaxton, 2011; Spencer *et al.*, 2015; Wagner *et al.*, 2006; Wong *et al.*, 2020). Although nanomedicine is relatively new, this research field evolves very fast and is being increasingly used not only for biomedical applications, but also in many other fields of our daily life (Henderson and Shankar, 2017; MacEwan and Chilkoti, 2017; Wong *et al.*, 2020). In the medical world, "nano" materials represent one of the most promising approaches not only for therapeutic purposes, but also diagnostics and theranostics approaches (Davis *et al.*, 2008; Eifler and Thaxton, 2011; Jain and Stylianopoulos, 2010; Liu *et al.*, 2020; MacEwan and Chilkoti, 2017; Rai *et al.*, 2021). According to the criteria from the US Food and Drug Administration (FDA), nanodrugs are those products that range from 1 to 100 nm, and due to their small size and high

surface area exhibit key differences compared to bulk materials or materials outside of this range that exhibit related dimension-dependent properties (Bobo *et al.*, 2016; Sainz *et al.*, 2015; Wolfram *et al.*, 2015). Biomedical nanodevices include liposomes, dendrimers, polymer nanoparticles, micelles, nanocrystals, metals, other inorganic materials and proteins (Behzadi *et al.*, 2017; Shi *et al.*, 2017). Figure 1 depicts different shapes used in nanomedicine. To overcome the abnormal physiology of tumor tissue, nanomedicine takes advantage of the so-called Enhanced Permeability and Retention (EPR) effect to improve drug accumulation within the tumor thereby reaching therapeutic drug amounts (Maeda, 2015; Matsumura and Maeda, 1986).

The options for the design and functionalization of nanomaterials are hugely varied and the list of potential applications increases more and more, so current tendency in nanomedicine points to tailor-made devices (Tran et al., 2017). Nonetheless, it is important to notice that nanomedicine-based treatments are not miracle cures, as they still have challenges to overcome. As instance, bioavailability is a typical challenge when developing a nanodevice (Yang et al., 2013). As the most important biological fluid, blood contains more than 3000 different proteins. Once injected in the bloodstream, nanodevices interact with plasma proteins and, as a consequence, its surface is usually covered by various biomolecules (especially proteins) and the so-called corona is formed (Shemetov et al., 2012; Tenzer et al., 2013). This adsorption of proteins not only alters the particle size, stability and surface properties (Hühn et al., 2013), but also affects its behavior and distribution within the body (Aggarwal et al., 2009; Karmali and Simberg, 2011; Monopoli et al., 2012). Even though the composition of the protein corona varies, it is well known that more abundant proteins first bind to nanoparticles surface and then the composition is changed during the circulation time (Aggarwal et al., 2009). Thus, non-specific interactions with serum proteins are one limiting factor in terms of circulation time. Opsonins are some of the corona-forming proteins and are recognized by reticuloendothelial systems (RES) and the mononuclear phagocyte system (MPS), leading to rapid blood clearance and high liver and spleen accumulation, so this could be an interesting strategy when needed targeted delivery to these organs (Owens and Peppas, 2006). On the other hand, dysopsonins such as apolipoproteins and albumin, inhibit phagocytic uptake, increase blood circulation time (Walkey *et al.*, 2012). Covering the nanoparticles surface with polymers such as polyethylene glycol (PEG) is the most used approach in order to develop long-circulating nanoparticles and therefore prevent rapid clearance, as pegylation reduces protein adsorption through hydrophilicity and steric repulsion effects thereby resulting in longer blood circulation time and lower accumulation in the liver (Gref *et al.*, 2000; Owens and Peppas, 2006). Furthermore, it has been demonstrated that increased density of PEG on the surface of gold nanoparticles can decrease the amount and change the types of the protein corona and reduce macrophage uptake *in vitro* (Walkey *et al.*, 2012).

Blood circulation time is directly related to the efficient extravasation of a nanoparticle. Thus, short blood circulation half-life may be sufficient for tissues with relatively large blood flow. Contrary, longer circulation half-lives are necessary to progressively extravasate in poorly perfused tissues (Shi *et al.*, 2017). Moreover, nanosystems biodistribution within the body is dramatically affected by their size. Thus, nanodevices smaller than 10 nm are cleared by renal filtration (Owens and Peppas, 2006). Contrary, it is well known that molecules bigger than 100 nm are accumulated in the liver, which is one of the most problematic side effects of current drugs. Different sizes of nanodevices are preferred for overcoming biological barriers and achieving their target. As instance, 11–30 nm size nanoparticles are suitable for liver and brain, whereas nanodevices with 31–80 nm diameter are appropriate for lungs, tumors and inflamed tissue (Souris *et al.*, 2010). Therefore, nanosystems between 10 and 100 nm are preferred due to their accurate accumulation and effect in desired tissues and organs.



**Figure 2.** Scientific articles published referring to "Cancer nanomedicine" (A) or "Cancer nanomedicine clinical trial" to 2020 (B). The number of articles published in each year was identified by searching the terms referred in the legend in the PubMed database (https://pubmed.ncbi.nlm.nih.gov/), queried on 29 November 2020.

Nanomedicine has emerged as an encouraging approach against cancer (Figure 2). In fact, there are 110 clinical trials involving the application of nanotechnology for cancer

treatment nowadays, according to the NIH (NIH, 2020). The development of accurate drug delivery systems is a promising strategy in order to improve the selective action of unspecific drugs and, in fact, most encapsulated drugs in novel nanodevices are clinical chemotherapeutic agents, such as doxorubicin, paclitaxel and docetaxel (Han et al., 2017; Kushwah et al., 2018; Saw et al., 2017). This fact is due not only to the high incidence of cancer, but also because tumors present a characteristic physiology which is a huge challenge for biomedical research and demands therapeutic agents to have special features (Spencer et al., 2015). Thus, nanotechnology has explored over the past several decades different approaches in order to achieve better encapsulation devices, such as liposomes, nanoparticles, micelles or dendrimers (Brannon-Peppas and Blanchette, 2004; Howes et al., 2014; MacEwan and Chilkoti, 2017; Minelli et al., 2010; Shi et al., 2017). Furthermore, some of these promising nanodevices have been used in clinical trials. In fact, the chemotherapeutic drug doxorubicin, which is currently used for the treatment of breast cancer among others, is administered with liposomal nature. As most clinically used chemotherapeutic agents are highly hydrophobic, drug encapsulation inside nanocarriers allow us to achieve higher concentrations within tumor cells (Behzadi et al., 2017; Kushwah et al., 2018; Luginbuhl et al., 2017; Yousefpour et al., 2019).

**Table 1.** A summary of different types of biomedical nanodevices for cancer therapy described in this review.

Туре	Example	Chemotherapeutic agent	Targeted nanosystem	Indication	
Liposome	Doxil®	Doxil® Doxorubicin		AIDS-related Kaposi's sarcoma, Acute myeloid leukemia and ovarian cancer	
	MbP-426	Oxaliplatin	Yes	Gastric cancer	
	SGT-53	p53 gene	Yes	Solid tumors	
	MCC-465	Doxorubicin	Yes	Colorectal and stomach cancer	
PEGylated protein	Oncaspar®	PEG-1-asparaginase,	No	Acute lymphocytic leukemia, Lymphoblastic leukemia, Lymphoma	

Polymer-drug conjugate		PK2	Doxorubicin	Yes	Liver cancer
Particle	Albumin- based	Abraxane®	Paclitaxel	No	Breast Cancer, Non-Small Cell Lung Cancer, Pancreatic Cancer
	Polymer	CAlAA-01	siRNA	Yes	Solid tumors

### 4. Clinically approved therapeutics

Table 1 summarizes the uses and content of nanosystems for drug delivery purposes described in this review. Doxil® is a representative example of non-targeted nanotherapeutic agent, as it has been used in the clinic for over two decades (Green and Rose, 2006; O'Brien, 2008). This small PEG-liposome (100 nm) containing the cytotoxic drug doxorubicin was first approved against AIDS-related Kaposi's sarcoma and is now approved for clinical use in ovarian cancer and multiple myeloma (Chanan-Khan and Lee, 2007). Although Doxil® also has important advantages for the clinic when compared to free doxorubicin, such as a 100-times longer half-life inside the body circulation and a reduction in cardiotoxicity, on the other hand it provokes skin toxicity that is not observed after administration of the free drug (Uziely *et al.*, 1995).

Along this same line, Abraxane<sup>®</sup>, a 120 nm size albumin nanoparticle containing paclitaxel, was designed and developed to retain the therapeutic activity of paclitaxel without the toxicity associated with the emulsifier Cremophor El<sup>®</sup> involved in the paclitaxel formulation (Taxol<sup>®</sup>) (Sparreboom *et al.*, 2005). A phase III study involving 454 patients with metastatic breast cancer compared Abraxane<sup>®</sup> and Taxol<sup>®</sup> (Gradishar *et al.*, 2005). Results showed up to 80% higher maximally tolerated dose (MTD) of Abraxane<sup>®</sup> and significantly greater response rates compared to patients treated with Taxol<sup>®</sup>. Furthermore, the Abraxane<sup>®</sup> group showed significantly lower incidence of

grade 4 neutropenia compared to patients treated with Taxol®. Therefore, the clinical advantage associated with Abraxane® is not only due to its "nano" form but also to the lack in the formulation of toxic Cremophor El® (Henderson and Bhatia, 2007). Interestingly, and as a direct consequence of this, Abraxane® does not require the use of pre-medications that are normally required for Taxol® treatment.

Multiple nanotherapeutic agents based on PEGylated proteins have been also approved for clinical use. PEGylation strategy is widely used as it increases the solubility, prevents rapid renal clearance, reduces immunogenicity and increases circulation times (Harris and Chess, 2003; Posey *et al.*, 2005; Rowinsky *et al.*, 2003). For these reasons, different proteins, such as enzymes, cytokines and monoclonal antibody Fab fragments, have been typically PEGylated. The US Food and Drug Administration approved Oncaspar®, a PEG-1-asparaginase, in 1994 to treat acute lymphoblastic leukaemia (Graham, 2003). Although the naked form of the drug accurately depletes asparagine and is active against acute lymphoblastic leukaemia and lymphoma, a hypersensitivity reaction and antibody production are induced, thereby leading to rapid clearance from the circulation. Oncaspar® was shown to have increased plasma half-life compared to the naked 1asparaginase and reduced the hypersensitivity reaction in patients with refractory lymphoma (Abshire *et al.*, 2000; Agrawal *et al.*, 2003; Ho *et al.*, 1986).

Furthermore, cancer cells are characterized by higher expression of multiple proteins, not only cytoplasmic proteins, but also cell membrane receptors (Byrne *et al.*, 2008). These cancer markers are of huge interest, as we can use different targets depending on the type of tumor and we can even differentiate primary tumors from distant metastases (Byrne *et al.*, 2008). The surface of nanoparticles can therefore be decorated with molecules as

targeting systems in order to specifically drive these therapeutic nanodevices to cancer cells and thereby avoid undesired effects in healthy cells (Shi *et al.*, 2017). Thus, nanotechnology can take advantage of cancer markers in order to develop advanced nanocarriers that allow us to achieve personalized biomedical therapeutics (Eskandari *et al.*, 2020; Henderson and Shankar, 2017).

PK2, a HPMA polymer-doxorubicin conjugate, was the first targeted nanotherapeutic agent to reach the clinic. The nanoparticle includes galactosamine to target the asialoglycoprotein receptor (ASGPR), which is typically expressed in hepatocytes (Reshitko *et al.*, 2020). As ASGPR is expressed in both healthy hepatocytes and primary liver cancer cells, the targeted nanoparticles get accumulated in normal liver cells as well as in the tumor (Hopewel et al., 2001). Concentrations of the drug in the liver were 15-20% of the administered dose after 24 hours (Julyan et al., 1999) and the concentrations within the tumor tissue were up to 50-fold higher than would have been achieved through free doxorubicin. Currently, the only targeted nanoparticles in the clinic are specifically driven to the transferrin receptor, which is known to be upregulated in many types of cancer (Gatter et al., 1983). Thus, we can find MbP-426, a liposome containing the cytotoxic platinum-based drug oxaliplatin (Sankhala et al., 2009); SGT-53, which contains a plasmid encoding for the tumor suppressor p53 in a liposome (Pirollo et al., 2016; Senzer et al., 2013); and CAIAA-01, a 70 nm polymer-based nanoparticle containing small interfering RNA (siRNA) (Davis, 2009; Eifler and Thaxton, 2011, Zuckerman et al., 2014). MCC-465, an immunoliposome-encapsulated doxorubicin with encouraging results in clinical trials against colorectal and stomach cancer (Fernandes et al., 2015; Hamaguchi et al., 2004; Matsumura et al., 2004), is not in use yet. Other examples of clinically approved nanodrugs are summarized in Table 2.

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Product	Туре	Chemotherapeutic agent	Size (nm)	Indication	Approved by	Biologic License Application
Adagen®	Polymeric nanoparticle	Bovine pegademase	n/a	Severe Combined Immunodeficiency (SCID)	FDA	019818
Bexxar®	Antibody- radioactive element conjugate	iodine-131	n/a	Non-Hodgkin's lymphoma	FDA	125011
DaunoXome®	Liposome	Daunorubicin	45	AIDS-related Kaposi's sarcoma, metastatic ovarian cancer, metastatic breast cancer, multiple myeloma	FDA	050704
DepoCyt®	Liposome	Cytarabine	n/a	Lymphomas or leukemia with meningeal spread add neoplastic meningitis	FDA	21-041
Eligard®	Polymeric nanoparticle	Leuprolide acetate	n/a	Prostate cancer	FDA	021731
Kadcyla®	Antibody- drug conjugate	DM1	n/a	Metastatic breast cancer	FDA	125427/0
Marqibo®	Liposome	Vincristine	163	Acute lymphoid leukemia	FDA	202497
Mepact®	Liposome	Mifamurtide	n/a	Osteosarcoma	EMA	EMEA/H/C/000802
Myocet®	Liposome	Doxorubicin	150	Breast	EMA	EMEA/H/C/000297
Mylotarg®	Antibody- drug conjugate	Calicheamicin	n/a	Acute myeloid leukemia.	FDA	761060
Onivyde®	Liposome	Irinotecan	n/a	Pancreatic cancer	FDA	207793
Ontak®	Fusion protein	Denileukin diftito	n/a	Cutaneous T-cell lymphoma	FDA; EMA	103767; EU/3/01/075
Vyxeos®	Liposome	Daunorubicin and Cytarabine	n/a	Acute Myeloid Leukemia	FDA	209401
Zevalin®	Antibody- radioactive element conjugate	yttrium-90	n/a	Non-Hodgkin's lymphoma	FDA; EMA	125019; EMEA/H/C/000547

**Table 2.** Examples of clinically approved biomedical nanodevices for cancer therapy.

# 5. Challenges

Table 3 shows different nanoformulations currently used in clinical trials. One of the main disappointing obstacles is the stark contrast between the high amount of successful nanotherapeutics in preclinical studies in the laboratory at both *in vitro* and *in vivo* level and the outcomes from clinical trials. Despite multiple animal models are currently available, from cell line-based subcutaneous and orthotopic xenografts or patient-derived xenografts (PDXs) to genetically engineered mouse models (GEMMs), there is a lack of tumour models able to fully reproduce all aspects of human cancer (Choi *et al.*, 2014; Sharpless and Depinho, 2006). Furthermore, it is of great importance to highlight the major contribution of tumor metastases to cancer mortality, whereas models of human tumor metastasis are not valid for the nanotherapeutic penetration within metastatic tissues compared to primary tumors. Therefore, the translation of therapeutic nanosystems could be improved by the development of novel animal models able to mimic the heterogeneity and special physiology of human tumors (Hubbard *et al.*, 2016; Lin *et al.*, 2014; Rongvaux *et al.*, 2014).

Moreover, the manufacture and the escalating complexity are important challenges for clinical development of therapeutic nanosystems. The transition from preclinical to clinical development and subsequent commercialization requires predetermined standards of quality, controls and good manufacturing practice, which can be achieved by means of manufacturing unit operations already available in the pharmaceutical industry.

Type of nanomaterial	Product	Chemotherapeutic agent	Indication	Status	Clinical trial reference
Dendrimer	AZD4320	AZD4320	Advanced solid tumors; lymphoma; multiple myeloma; Hematologic malignancies	Phase I	NCT04214093
Gold nanoparticle	Aurimmune	TNF	Solid tumor	Phase I	NCT00356980
	CPX1	Irinotecan	Colorectal cancer	Phase II	NCT00361842
	LE-SN38	Sn-38	Neoplasms	Phase II	NCT00046540
Liposome	Lipoplatin <sup>TM</sup>	Cisplatin	Non-small cell lung cancer; breast cancer; gastric cancer	Phase III	NCT02702700
	LNDDP	Cisplatin	Malignant mesothelioma	Phase II	NCT00004033
	2B3-101	Doxorubicin	Glioma	Phase I,II	NCT01386580
Polymeric conjugate	XMT1001	Camptothecin	Small cell lung cancer; Non-small cell lung cancer;	Phase I	NCT00455052
	Cpc634	Docetaxel	Ovarian cancer	Phase II	NCT03742713
	CRLX101- Olaparib	Olaparib	Urothelial cancer; Small cell lung cancer; Non-small cell lung cancer; prostate cancer	Phase I; Phase II	NCT02769962
Polymeric micelle	Nk105	Paclitaxel	Breast cancer nor metastatic recurrent	Phase III	NCT01644890
	Nanoxel® M	Docetaxel	Head and neck squamous cell carcinoma	Phase II	NCT02639858

**Table 3.** Examples of biomedical nanosytems in clinical trials for cancer therapy.

	Lipusu®	Paclitaxel	Breast cancer	Phase IV	NCT02142790
	Nanoplatin	Gemcitabine	Locally advanced and metastatic pancreatic cancer	Phase I; Phase II	NCT00910741
	E7389-e044- 112	Eribulin-lf	Solid tumors	Phase I	NCT01945710
	Nc6004	Cisplatin	Head and neck cancer	Phase II	NCT03771820
	Paclical	Paclitaxel	Ovarian cancer	Phase III	NCT00989131
Targeted minicell	TargomiRs	miRNA mimic	Malignant pleural mesothelioma; Non-small cell lung cancer	Phase I	NCT02369198

## 6. Conclusion

Despite the numerous advances in oncology, chemotherapeutic drugs currently used in the clinic present important limitations, such as low specificity of therapeutic agents with important side effects on healthy tissues. Thus, novel strategies for achieving an accurate action on targeted cells are needed. Nanotechnology and medicine have been combined thereby constituting the nanomedicine field, which consists of the development of therapeutic devices at the "nano" scale aiming to tackle cancer cells specifically. Even though key steps are still needed to improve its accuracy, nanomedicine is becoming more and more promising, as demonstrated by the increasing number of clinical trials involving nanodrugs.

Nanomedicine is continuously evolving and more sophisticated multifunctional nanotherapeutics are reaching the clinical practice. Even though many challenges still exist for the translation of nanodevices to clinically approved products, their potential advantages should lead to their successful development. Moreover, the continuing need

of novel classes of anticancer therapies mandates scientific research to achieve improved nano-based approaches. Ideally, nanomedicine will contribute to improve patient survival in the foreseeable future.

## **Conflict of Interest**

The authors declare no conflict of interest, financial or otherwise.

## Funding

The authors are grateful for financial support from the CIBER-BBN and the MICIUN

(DTS19/00162 and PID2019-106386RB-I00).

## References

Aamodt, J.M., Grainger, D.W., 2016. Extracellular matrix-based biomaterial scaffolds and the host response. Biomaterials. 86, 68-82. https://doi.org/10.1016/j.biomaterials.2016.02.003

Abshire, T.C., Pollock, B.H., Billett, A.L., Bradley, P., Buchanan, G.R., 2000. Weekly polyethylene glycol conjugated L-asparaginase compared with biweekly dosing produces superior induction remission rates in childhood relapsed acute lymphoblastic leukemia: a Pediatric Oncology Group Study. Blood. 96, 1709-1715.

Aggarwal, P., Hall, J.B., McLeland, C.B., Dobrovolskaia, M.A., McNeil, S.E., 2009. Nanoparticle interaction with plasma proteins as it relates to particle biodistribution, biocompatibility and therapeutic efficacy. Adv. Drug Deliv. Rev. 61, 428-437. https://doi.org/10.1016/j.addr.2009.03.009

Agrawal, N.R., Bukowski, R.M., Rybicki, L.A., Kurtzberg, J., Cohen, L.J., Hussein, M.A., 2003. A Phase I-II trial of polyethylene glycol-conjugated L-asparaginase in patients with multiple myeloma. Cancer. 98, 94-99. https://doi.org/10.1002/cncr.11480

Allen, T.M., Cullis, P.R., 2013. Liposomal drug delivery systems: from concept to clinical applications. Adv. Drug Deliv. Rev. 65, 36-48. https://doi.org/10.1016/j.addr.2012.09.037

Astete, C.E., Sabliov, C.M., 2006. Synthesis and characterization of PLGA nanoparticles. J. Biomater. Sci. Polym. Ed. 17, 247-289. https://doi.org/10.1163/156856206775997322

Barenholz, Y., 2012. Doxil<sup>®</sup>--the first FDA-approved nano-drug: lessons learned. J. Control. Release. 160, 117-134. https://doi.org/10.1016/j.jconrel.2012.03.020

Behzadi, S., Serpooshan, V., Tao, W., Hamaly, M.A., Alkawareek, M.Y., Dreaden, E.C., Brown, D., Alkilany, A.M., Farokhzad, O.C., Mahmoudi, M., 2017. Cellular uptake of nanoparticles: journey inside the cell. Chem. Soc. Rev. 46, 4218-4244. https://doi.org/10.1039/c6cs00636a

Beri, P., Matte, B.F., Fattet, L., Kim, D., Yang, J., Engler, A.J., 2018. Biomaterials to model and measure epithelial cancers. Nat. Rev. Mater. 3, 418-430.

Biggs, M., Zeugolis, D., Pandit, A., 2018. Foreword to special issue on two-dimensional biomaterials in regenerative medicine. Nanomedicine. 14, 2351-2353. https://doi.org/10.1016/j.nano.2017.10.005

Bobo, D., Robinson, K.J., Islam, J., Thurecht, K.J., Corrie, S.R., 2016. Nanoparticle-Based Medicines: A Review of FDA-approved materials and clinical trials to date. Pharm. Res. 33, 2373-2387. https://doi.org/10.1007/s11095-016-1958-5

https://doi.org/10.2147/ijn.s139687

Brannon-Peppas, L., Blanchette, J.O., 2004. Nanoparticle and targeted systems for cancer therapy. Adv. Drug Deliv. Rev. 56, 1649-1659. https://doi.org/10.1016/j.addr.2004.02.014

Brouillard, A., Deshpande, N., Kulkarni, A. A., 2021. Engineered multifunctional nano- and biological materials for cancer immunotherapy. Adv. Healthc. Mater. 1680-1692. https://doi.org/ 10.1002/adhm.202001680

https://doi.org/10.1016/j.addr.2008.08.005

Chanan-Khan, A.A., Lee, K., 2007. Pegylated liposomal doxorubicin and immunomodulatory drug combinations in multiple myeloma: rationale and clinical experience. Clin. Lymphoma Myeloma. 7 Suppl 4, S163-169. https://doi.org/10.3816/clm.2007.s.018

Choi, S.M., Chaudhry, P., Zo, S.M., Han, S.S., 2018. Advances in protein-based materials: from origin to novel biomaterials. Adv. Exp. Med. Biol. 1078, 161-210. https://doi.org/10.1007/978-981-13-0950-2\_10

Choi, S.Y., Lin, D., Gout, P.W., Collins, C.C., Xu, Y., Wang, Y., 2014. Lessons from patient-derived xenografts for better in vitro modeling of human cancer. Adv. Drug Deliv. Rev. 79-80, 222-237. https://doi.org/10.1016/j.addr.2014.09.009

Creixell, M., Peppas, N.A., 2012. Co-delivery of siRNA and therapeutic agents using nanocarriers to overcome cancer resistance. Nano Today. 7, 367-379. https://doi.org/10.1016/j.nantod.2012.06.013

Dahlman, J.E., Barnes, C., Khan, O., Thiriot, A., Jhunjunwala, S., Shaw, T.E., Xing, Y., Sager, H.B., Sahay, G., Speciner, L., Bader, A., Bogorad, R.L., Yin, H., Racie, T., Dong, Y., Jiang, S., Seedorf, D., Dave, A., Sandu, K.S., Webber, M.J., Novobrantseva, T., Ruda, V.M., Lytton-Jean, A.K.R., Levins, C.G., Kalish, B., Mudge, D.K., Perez, M., Abezgauz, L., Dutta, P., Smith, L., Charisse, K., Kieran,

Schroeder, A., Panigrahy, D., Kotelianski, V., Langer, R., Anderson, D.G., 2014. In vivo endothelial siRNA delivery using polymeric nanoparticles with low molecular weight. Nat. Nanotechnol. 9, 648-655. https://doi.org/10.1038/nnano.2014.84

Daniel, D., Crawford, J., 2006. Myelotoxicity from chemotherapy. Semin. Oncol. 33, 74-85. https://doi.org/10.1053/j.seminoncol.2005.11.003

Davis, M.E., 2009. The first targeted delivery of siRNA in humans via a self-assembling, cyclodextrin polymer-based nanoparticle: from concept to clinic. Mol. Pharm. 6, 659-668. https://doi.org/10.1021/mp900015y

Davis, M.E., Chen, Z.G., Shin, D.M., 2008. Nanoparticle therapeutics: an emerging treatment modality for cancer. Nat. Rev. Drug Discov. 7, 771-782. https://doi.org/10.1038/nrd2614

DeBerardinis, R.J., Chandel, N.S., 2016. Fundamentals of cancer metabolism. Sci. Adv. 2, e1600200. https://doi.org/10.1126/sciadv.1600200

Eifler, A.C., Thaxton, C.S., 2011. Nanoparticle therapeutics: FDA approval, clinical trials, regulatory pathways, and case study. Methods Mol. Biol. 726, 325-338. https://doi.org/10.1007/978-1-61779-052-2\_21

Eskandari, Z., Bahadori, F., Celik, B., Onyuksel, H., 2020. Targeted nanomedicines for cancer therapy, from basics to clinical trials. J. Pharm. Pharm. Sci. 23, 132-157. https://doi.org/10.18433/jpps30583

Fernandes, E., Ferreira, J.A., Andreia, P., Luís, L., Barroso, S., Sarmento, B., Santos, L.L., 2015. New trends in guided nanotherapies for digestive cancers: A systematic review. J. Control. Release. 209, 288-307. https://doi.org/10.1016/j.jconrel.2015.05.003

Gatter, K.C., Brown, G., Trowbridge, I.S., Woolston, R.E., Mason, D.Y., 1983. Transferrin receptors in human tissues: their distribution and possible clinical relevance. J. Clin. Pathol. 36, 539-545. https://doi.org/10.1136/jcp.36.5.539

https://doi.org/10.1056/NEJMoa1113205

https://doi.org/10.2147/ijn.s102385

Gonzalez-Valdivieso, J., Borrego, B., Girotti, A., Moreno, S., Brun, A., Bermejo-Martin, J.F., Arias, F.J., 2020. A DNA vaccine delivery platform based on Elastin-Like Recombinamer nanosystems for Rift Valley fever virus. Mol. Pharm. 17, 1608-1620. https://doi.org/10.1021/acs.molpharmaceut.0c00054

Gonzalez-Valdivieso, J., Girotti, A., Muñoz, R., Rodriguez-Cabello, J.C., Arias, F.J., 2019. Self-Assembling ELR-Based Nanoparticles as smart drug-delivery systems modulating cellular growth via Akt. Biomacromolecules. 20, 1996-2007. https://doi.org/10.1021/acs.biomac.9b00206

Gradishar, W.J., Tjulandin, S., Davidson, N., Shaw, H., Desai, N., Bhar, P., Hawkins, M., O'Shaughnessy, J., 2005. Phase III trial of nanoparticle albumin-bound paclitaxel compared with polyethylated castor oil-based paclitaxel in women with breast cancer. J. Clin. Oncol. 23, 7794-7803. https://doi.org/10.1200/jco.2005.04.937

Graham, M.L., 2003. Pegaspargase: a review of clinical studies. Adv. Drug Deliv. Rev. 55, 1293-1302. https://doi.org/10.1016/s0169-409x(03)00110-8

Green, A.E., Rose, P.G., 2006. Pegylated liposomal doxorubicin in ovarian cancer. Int. J. Nanomedicine. 1, 229-239.

Gref, R., Lück, M., Quellec, P., Marchand, M., Dellacherie, E., Harnisch, S., Blunk, T., Müller, R.H., 2000. 'Stealth' corona-core nanoparticles surface modified by polyethylene glycol (PEG): influences of the corona (PEG chain length and surface density) and of the core composition on phagocytic uptake and plasma protein adsorption. Colloids Surf. B Biointerfaces. 18, 301-313. https://doi.org/10.1016/s0927-7765(99)00156-3

Grothey, A., 2003. Oxaliplatin-safety profile: neurotoxicity. Semin. Oncol. 30, 5-13. https://doi.org/10.1016/s0093-7754(03)00399-3

https://doi.org/10.1016/j.biomaterials.2004.10.012

Hamaguchi, T., Matsumura, Y., Nakanishi, Y., Muro, K., Yamada, Y., Shimada, Y., Shirao, K., Niki, H., Hosokawa, S., Tagawa, T., Kakizoe, T., 2004. Antitumor effect of MCC-465, pegylated liposomal doxorubicin tagged with newly developed monoclonal antibody GAH, in colorectal cancer xenografts. Cancer Sci. 95, 608-613. https://doi.org/10.1111/j.1349-7006.2004.tb02495.x

Han, W., Chilkoti, A., López, G.P., 2017. Self-assembled hybrid elastin-like polypeptide/silica nanoparticles enable triggered drug release. Nanoscale. 9, 6178-6186. https://doi.org/10.1039/c7nr00172j

Harris, J.M., Chess, R.B., 2003. Effect of pegylation on pharmaceuticals. Nat. Rev. Drug Discov. 2, 214-221. https://doi.org/10.1038/nrd1033

Henderson, I.C., Bhatia, V., 2007. Nab-paclitaxel for breast cancer: a new formulation with an improved safety profile and greater efficacy. Expert Rev. Anticancer Ther. 7, 919-943. https://doi.org/10.1586/14737140.7.7.919

Henderson, L.A., Shankar, L.K., 2017. Clinical translation of the National Institutes of Health's Investments in nanodrug products and devices. Aaps j. 19, 343-359. https://doi.org/10.1208/s12248-016-9995-x

Heng, P.W.S., 2018. Controlled release drug delivery systems. Pharm. Dev. Technol. 23, 833. https://doi.org/10.1080/10837450.2018.1534376

Hinshaw, D.C., Shevde, L.A., 2019. The tumor microenvironment innately modulates cancer progression. Cancer Res. 79, 4557-4566. https://doi.org/10.1158/0008-5472.can-18-3962

Ho, D.H., Brown, N.S., Yen, A., Holmes, R., Keating, M., Abuchowski, A., Newman, R.A., Krakoff, I.H., 1986. Clinical pharmacology of polyethylene glycol-L-asparaginase. Drug Metab. Dispos. 14, 349-352.

Hopewel, J.W., Duncan, R., Wilding, D., Chakrabarti, K., 2001. Preclinical evaluation of the cardiotoxicity of PK2: a novel HPMA copolymer-doxorubicin-galactosamine conjugate antitumour agent. Hum. Exp. Toxicol. 20, 461-470. https://doi.org/10.1191/096032701682693017

Howes, P.D., Chandrawati, R., Stevens, M.M., 2014. Bionanotechnology. Colloidal nanoparticlesasadvancedbiologicalsensors.Science.346,1247390.https://doi.org/10.1126/science.1247390

Hubbard, G.K., Mutton, L.N., Khalili, M., McMullin, R.P., Hicks, J.L., Bianchi-Frias, D., Horn, L.A., Kulac, I., Moubarek, M.S., Nelson, P.S., Yegnasubramanian, S., De Marzo, A.M., Bieberich, C.J., 2016. Combined MYC activation and Pten loss are sufficient to create genomic instability and lethal metastatic prostate cancer. Cancer Res. 76, 283-292. https://doi.org/10.1158/0008-5472.can-14-3280

Huebsch, N., Mooney, D.J., 2009. Inspiration and application in the evolution of biomaterials. Nature. 462, 426-432. https://doi.org/10.1038/nature08601

Hühn, D., Kantner, K., Geidel, C., Brandholt, S., De Cock, I., Soenen, S.J., Rivera Gil, P., Montenegro, J.M., Braeckmans, K., Müllen, K., Nienhaus, G.U., Klapper, M., Parak, W.J., 2013. Polymer-coated nanoparticles interacting with proteins and cells: focusing on the sign of the net charge. ACS Nano. 7, 3253-3263. https://doi.org/10.1021/nn3059295

Jain, R.K., Stylianopoulos, T., 2010. Delivering nanomedicine to solid tumors. Nat. Rev. Clin. Oncol. 7, 653-664. https://doi.org/10.1038/nrclinonc.2010.139

Julyan, P.J., Seymour, L.W., Ferry, D.R., Daryani, S., Boivin, C.M., Doran, J., David, M., Anderson, D., Christodoulou, C., Young, A.M., Hesslewood, S., Kerr, D.J., 1999. Preliminary clinical study of the distribution of HPMA copolymers bearing doxorubicin and galactosamine. J. Control. Release. 57, 281-290. https://doi.org/10.1016/s0168-3659(98)00124-2

Karmali, P.P., Simberg, D., 2011. Interactions of nanoparticles with plasma proteins: implication on clearance and toxicity of drug delivery systems. Expert Opin. Drug Deliv. 8, 343-357. https://doi.org/10.1517/17425247.2011.554818

Kibria, G., Hatakeyama, H., Ohga, N., Hida, K., Harashima, H., 2013. The effect of liposomal size on the targeted delivery of doxorubicin to Integrin  $\alpha\nu\beta$ 3-expressing tumor endothelial cells. Biomaterials. 34, 5617-5627. https://doi.org/10.1016/j.biomaterials.2013.03.094

Krieghoff-Henning, E., Folkerts, J., Penzkofer, A., Weg-Remers, S., 2017. Cancer – an overview. Med. Monatsschr. Pharm. 40, 48-54.

Kumar, M., Gupta, D., Singh, G., Sharma, S., Bhat, M., Prashant, C.K., Dinda, A.K., Kharbanda, S., Kufe, D., Singh, H., 2014. Novel polymeric nanoparticles for intracellular delivery of peptide Cargos: antitumor efficacy of the BCL-2 conversion peptide NuBCP-9. Cancer Res. 74, 3271-3281. https://doi.org/10.1158/0008-5472.can-13-2015

Kushwah, V., Katiyar, S.S., Agrawal, A.K., Gupta, R.C., Jain, S., 2018. Co-delivery of docetaxel and gemcitabine using PEGylated self-assembled stealth nanoparticles for improved breast cancer therapy. Nanomedicine. 14, 1629-1641. https://doi.org/10.1016/j.nano.2018.04.009

therapeutics for gastrointestinal disorders. Dig. Liver Dis. 45, 995-1002. https://doi.org/ 10.1016/j.dld.2013.03.019

Lin, D., Wyatt, A.W., Xue, H., Wang, Y., Dong, X., Haegert, A., Wu, R., Brahmbhatt, S., Mo, F., Jong, L., Bell, R.H., Anderson, S., Hurtado-Coll, A., Fazli, L., Sharma, M., Beltran, H., Rubin, M., Cox, M., Gout, P.W., Morris, J., Goldenberg, L., Volik, S.V., Gleave, M.E., Collins, C.C., Wang, Y., 2014. High fidelity patient-derived xenografts for accelerating prostate cancer discovery and drug development. Cancer Res. 74, 1272-1283. https://doi.org/10.1158/0008-5472.can-13-2921-t

Liu, X., Tang, I., Wainberg, Z.A., Meng, H., 2020. Safety considerations of cancer nanomedicine-A key step toward translation. Small. e2000673. https://doi.org/10.1002/smll.202000673

Liu, Z., Robinson, J.T., Sun, X., Dai, H., 2008. PEGylated nanographene oxide for delivery of waterinsoluble cancer drugs. J. Am. Chem. Soc. 130, 10876-10877. https://doi.org/10.1021/ja803688x

Lopes, J.R., Santos, G., Barata, P., Oliveira, R., Lopes, C.M., 2013. Physical and chemical stimuliresponsive drug delivery systems: targeted delivery and main routes of administration. Curr. Pharm. Des. 19, 7169-7184. https://doi.org/10.2174/13816128113199990698

Luginbuhl, K.M., Mozhdehi, D., Dzuricky, M., Yousefpour, P., Huang, F.C., Mayne, N.R., Buehne, K.L., Chilkoti, A., 2017. Recombinant synthesis of hybrid lipid-peptide polymer fusions that selfassemble and encapsulate hydrophobic drugs. Angew. Chem 56, 13979-13984. https://doi.org/10.1002/anie.201704625

MacEwan, S.R., Chilkoti, A., 2017. From composition to cure: A systems engineering approach to anticancer drug carriers. Angew. Chem 56, 6712-6733. https://doi.org/10.1002/anie.201610819

Maeda, H., 2015. Toward a full understanding of the EPR effect in primary and metastatic tumors as well as issues related to its heterogeneity. Adv. Drug Deliv. Rev. 91, 3-6. https://doi.org/10.1016/j.addr.2015.01.002

Maier-Hauff, K., Ulrich, F., Nestler, D., Niehoff, H., Wust, P., Thiesen, B., Orawa, H., Budach, V., Jordan, A., 2011. Efficacy and safety of intratumoral thermotherapy using magnetic iron-oxide nanoparticles combined with external beam radiotherapy on patients with recurrent glioblastoma multiforme. J. Neurooncol. 103, 317-324. https://doi.org/10.1007/s11060-010-0389-0

https://doi.org/10.1016/j.tips.2009.08.004

Matsumura, Y., Gotoh, M., Muro, K., Yamada, Y., Shirao, K., Shimada, Y., Okuwa, M., Matsumoto, S., Miyata, Y., Ohkura, H., Chin, K., Baba, S., Yamao, T., Kannami, A., Takamatsu, Y., Ito, K., Takahashi, K., 2004. Phase I and pharmacokinetic study of MCC-465, a doxorubicin (DXR) encapsulated in PEG immunoliposome, in patients with metastatic stomach cancer. Ann. Oncol. 15, 517-525. https://doi.org/10.1093/annonc/mdh092

Matsumura, Y., Maeda, H., 1986. A new concept for macromolecular therapeutics in cancer chemotherapy: mechanism of tumoritropic accumulation of proteins and the antitumor agent smancs. Cancer Res. 46, 6387-6392.

Mi, P., 2020. Stimuli-responsive nanocarriers for drug delivery, tumor imaging, therapy and theranostics. Theranostics. 10, 4557-4588. https://doi.org/10.7150/thno.38069

Minelli, C., Lowe, S.B., Stevens, M.M., 2010. Engineering nanocomposite materials for cancer therapy. Small. 6, 2336-2357. https://doi.org/10.1002/smll.201000523

Mitchell, E.P., 2006. Gastrointestinal toxicity of chemotherapeutic agents. Semin. Oncol. 33, 106-120. https://doi.org/10.1053/j.seminoncol.2005.12.001

Monopoli, M.P., Aberg, C., Salvati, A., Dawson, K.A., 2012. Biomolecular coronas provide the biological identity of nanosized materials. Nat. Nanotechnol. 7, 779-786. https://doi.org/10.1038/nnano.2012.207

Mura, S., Couvreur, P., 2012. Nanotheranostics for personalized medicine. Adv. Drug Deliv. Rev. 64, 1394-1416. https://doi.org/10.1016/j.addr.2012.06.006

Murphy, S.V., Atala, A., 2014. 3D bioprinting of tissues and organs. Nat. Biotechnol. 32, 773-785. https://doi.org/10.1038/nbt.2958

Narang, A.S., Chang, R.K., Hussain, M.A., 2013. Pharmaceutical development and regulatory considerations for nanoparticles and nanoparticulate drug delivery systems. J. Pharm. Sci. 102, 3867-3882. https://doi.org/10.1002/jps.23691

NIH, 2020. www.clinicaltrials.gov (accessed November 12 2020)

Nikolova, M.P., Chavali, M.S., 2019. Recent advances in biomaterials for 3D scaffolds: A review. Bioact. Mater. 4, 271-292. https://doi.org/10.1016/j.bioactmat.2019.10.005

https://doi.org/10.2217/nnm.15.21

O'Brien, M.E., 2008. Single-agent treatment with pegylated liposomal doxorubicin for metastatic breast cancer. Anticancer Drugs. 19, 1-7. https://doi.org/10.1097/CAD.0b013e3282f14a00

Octavia, Y., Tocchetti, C.G., Gabrielson, K.L., Janssens, S., Crijns, H.J., Moens, A.L., 2012. Doxorubicin-induced cardiomyopathy: from molecular mechanisms to therapeutic strategies. J. Mol. Cell Cardiol. 52, 1213-1225. https://doi.org/10.1016/j.yjmcc.2012.03.006 Oliveira, E.P., Malysz-Cymborska, I., Golubczyk, D., Kalkowski, L., Kwiatkowska, J., Reis, R.L., Oliveira, J.M., Walczak, P., 2019. Advances in bioinks and in vivo imaging of biomaterials for CNS applications. Acta Biomater. 95, 60-72. https://doi.org/10.1016/j.actbio.2019.05.006

Oussoren, C., Zuidema, J., Crommelin, D.J., Storm, G., 1997. Lymphatic uptake and biodistribution of liposomes after subcutaneous injection. II. Influence of liposomal size, lipid compostion and lipid dose. Biochim. Biophys. Acta. 1328, 261-272. https://doi.org/10.1016/s0005-2736(97)00122-3

Owens, D.E., Peppas, N.A., 2006. Opsonization, biodistribution, and pharmacokinetics of<br/>polymeric nanoparticles.Int.J.Pharm.307,93-102.https://doi.org/10.1016/j.ijpharm.2005.10.010

Peppas, N.A., Langer, R., 1994. New challenges in biomaterials. Science. 263, 1715-1720. https://doi.org/10.1126/science.8134835

https://doi.org/10.1016/j.jconrel.2016.04.028

Pierna, M., Santos, M., Arias, F.J., Alonso, M., Rodríguez-Cabello, J.C., 2013. Efficient cell and cellsheet harvesting based on smart surfaces coated with a multifunctional and self-organizing elastin-like recombinamer. Biomacromolecules. 14, 1893-1903. https://doi.org/10.1021/bm400268v

Piña, M.J., Girotti, A., Serrano, S., Muñoz, R., Rodríguez-Cabello, J.C., Arias, F.J., 2020. A double safety lock tumor-specific device for suicide gene therapy in breast cancer. Cancer Lett. 470, 43-53. https://doi.org/10.1016/j.canlet.2019.11.031

Pirollo, K.F., Nemunaitis, J., Leung, P.K., Nunan, R., Adams, J., Chang, E.H., 2016. Safety and efficacy in advanced solid tumors of a targeted nanocomplex carrying the p53 gene used in combination with Docetaxel: A phase 1b study. Mol. Ther. 24, 1697-1706. https://doi.org/10.1038/mt.2016.135

Posey, J.A., Saif, M.W., Carlisle, R., Goetz, A., Rizzo, J., Stevenson, S., Rudoltz, M.S., Kwiatek, J., Simmons, P., Rowinsky, E.K., Takimoto, C.H., Tolcher, A.W., 2005. Phase 1 study of weekly polyethylene glycol-camptothecin in patients with advanced solid tumors and lymphomas. Clin. Cancer Res. 11, 7866-7871. https://doi.org/10.1158/1078-0432.ccr-05-0783

Quader, S., Kataoka, K., 2017. Nanomaterial-enabled cancer therapy. Mol. Ther. 25, 1501-1513. https://doi.org/10.1016/j.ymthe.2017.04.026

Rai, A., Noor, S., Ahmad, S.I., Alajmi, M.F., Hussain, A., Abbas, H., Hasan, G. M., 2021. Recent advances and implication of bioengineered nanomaterials in cancer theranostics. Medicina. 57, https://doi.org/10.3390/medicina57020091

Reshitko, G.S., Yamansarov, E.Y., Evteev, S.A., Lopatukhina, E.V., Shkil, D.O., Saltykova, I.V., Lopukhov, A.V., Kovalev, S.V., Lobov, A.N., Kislyakov, I.V., Burenina, O.Y., Klyachko, N.L., Garanina, A.S., Dontsova, O.A., Ivanenkov, Y.A., Erofeev, A.S., Gorelkin, P.V., Beloglazkina, E.K., Majouga, A.G., 2020. Synthesis and evaluation of new trivalent Lligands for hepatocyte targeting via the asialoglycoprotein receptor. Bioconjugate Chemistry. 31, 1313-1319. https://doi.org/10.1021/acs.bioconjchem.0c00202

Rongvaux, A., Willinger, T., Martinek, J., Strowig, T., Gearty, S.V., Teichmann, L.L., Saito, Y., Marches, F., Halene, S., Palucka, A.K., Manz, M.G., Flavell, R.A., 2014. Development and function of human innate immune cells in a humanized mouse model. Nat. Biotechnol. 32, 364-372. https://doi.org/10.1038/nbt.2858

Rowinsky, E.K., Rizzo, J., Ochoa, L., Takimoto, C.H., Forouzesh, B., Schwartz, G., Hammond, L.A., Patnaik, A., Kwiatek, J., Goetz, A., Denis, L., McGuire, J., Tolcher, A.W., 2003. A phase I and pharmacokinetic study of pegylated camptothecin as a 1-hour infusion every 3 weeks in patients with advanced solid malignancies. J. Clin. Oncol. 21, 148-157. https://doi.org/10.1200/jco.2003.03.143

Sanjay, S.T., Zhou, W., Dou, M., Tavakoli, H., Ma, L., Xu, F., Li, X., 2018. Recent advances of controlled drug delivery using microfluidic platforms. Adv. Drug Deliv. Rev. 128, 3-28. https://doi.org/10.1016/j.addr.2017.09.013

Sankhala, K.K., Mita, A.C., Adinin, R., Wood, L., Beeram, M., Bullock, S., Yamagata, N., Matsuno, K., Fujisawa, T., Phan, A., 2009. A phase I pharmacokinetic (PK) study of MBP-426, a novel liposome encapsulated oxaliplatin. J. Clin. Oncol. 27, 2535-2545. https://doi.org/10.1200/jco.2009.27.15\_suppl.2535

Santana-Armas, M. L., Tros de Ilarduya, C., 2021. Strategies for cancer gene-delivery improvement by non-viral vectors. Int. J. Pharm. 596, 120291-120306. https://doi.org/10.1016/j.ijpharm.2021.120291

Saw, P.E., Yu, M., Choi, M., Lee, E., Jon, S., Farokhzad, O.C., 2017. Hyper-cell-permeable micelles as a drug delivery carrier for effective cancer therapy. Biomaterials. 123, 118-126. https://doi.org/10.1016/j.biomaterials.2017.01.040

Sawant, R.R., Torchilin, V.P., 2012. Challenges in development of targeted liposomal therapeutics. AAPS J. 14, 303-315. https://doi.org/10.1208/s12248-012-9330-0

Senzer, N., Nemunaitis, J., Nemunaitis, D., Bedell, C., Edelman, G., Barve, M., Nunan, R., Pirollo, K.F., Rait, A., Chang, E.H., 2013. Phase I study of a systemically delivered p53 nanoparticle in advanced solid tumors. Mol. Ther. 21, 1096-1103. https://doi.org/10.1038/mt.2013.32

Sharpless, N.E., Depinho, R.A., 2006. The mighty mouse: genetically engineered mouse models in cancer drug development. Nat. Rev. Drug Discov. 5, 741-754. https://doi.org/10.1038/nrd2110

Shemetov, A.A., Nabiev, I., Sukhanova, A., 2012. Molecular interaction of proteins and peptides with nanoparticles. ACS Nano. 6, 4585-4602. https://doi.org/10.1021/nn300415x

https://doi.org/10.2174/0929867324666170830102409

Sparreboom, A., Scripture, C.D., Trieu, V., Williams, P.J., De, T., Yang, A., Beals, B., Figg, W.D., Hawkins, M., Desai, N., 2005. Comparative preclinical and clinical pharmacokinetics of a cremophor-free, nanoparticle albumin-bound paclitaxel (ABI-007) and paclitaxel formulated in Cremophor (Taxol). Clin. Cancer Res. 11, 4136-4143. https://doi.org/10.1158/1078-0432.ccr-04-2291

Spencer, D.S., Puranik, A.S., Peppas, N.A., 2015. Intelligent Nanoparticles for Advanced Drug Delivery in Cancer Treatment. Curr. Opin. Chem. Eng. 7, 84-92. https://doi.org/10.1016/j.coche.2014.12.003

Tang, F., Li, L., Chen, D., 2012. Mesoporous silica nanoparticles: synthesis, biocompatibility and drug delivery. Adv. Mater. 24, 1504-1534. https://doi.org/10.1002/adma.201104763

Tao, W., Kong, N., Ji, X., Zhang, Y., Sharma, A., Ouyang, J., Qi, B., Wang, J., Xie, N., Kang, C., Zhang, H., Farokhzad, O.C., Kim, J.S., 2019. Emerging two-dimensional monoelemental materials (Xenes) for biomedical applications. Chem. Soc. Rev. 48, 2891-2912. https://doi.org/10.1039/c8cs00823j

Taraballi, F., Sushnitha, M., Tsao, C., Bauza, G., Liverani, C., Shi, A., Tasciotti, E., 2018. Biomimetic tissue engineering: Tuning the immune and inflammatory response to implantable biomaterials. Adv. Healthc. Mater. 7, e1800490. https://doi.org/10.1002/adhm.201800490

#### https://doi.org/10.2174/138161210791208992

Tenzer, S., Docter, D., Kuharev, J., Musyanovych, A., Fetz, V., Hecht, R., Schlenk, F., Fischer, D., Kiouptsi, K., Reinhardt, C., Landfester, K., Schild, H., Maskos, M., Knauer, S.K., Stauber, R.H., 2013. Rapid formation of plasma protein corona critically affects nanoparticle pathophysiology. Nat. Nanotechnol. 8, 772-781. https://doi.org/10.1038/nnano.2013.181

Tinkle, S., McNeil, S.E., Mühlebach, S., Bawa, R., Borchard, G., Barenholz, Y.C., Tamarkin, L., Desai, N., 2014. Nanomedicines: addressing the scientific and regulatory gap. Ann. N. Y. Acad Sci. 1313, 35-56. https://doi.org/10.1111/nyas.12403

Tran, S., DeGiovanni, P.J., Piel, B., Rai, P., 2017. Cancer nanomedicine: a review of recent success in drug delivery. Clin. Transl. Med. 6, 44. https://doi.org/10.1186/s40169-017-0175-0

Tsoi, K.M., MacParland, S.A., Ma, X.Z., Spetzler, V.N., Echeverri, J., Ouyang, B., Fadel, S.M., Sykes, E.A., Goldaracena, N., Kaths, J.M., Conneely, J.B., Alman, B.A., Selzner, M., Ostrowski, M.A.,

Uziely, B., Jeffers, S., Isacson, R., Kutsch, K., Wei-Tsao, D., Yehoshua, Z., Libson, E., Muggia, F.M., Gabizon, A., 1995. Liposomal doxorubicin: antitumor activity and unique toxicities during two complementary phase I studies. J. Clin. Oncol. 13, 1777-1785. https://doi.org/10.1200/jco.1995.13.7.1777

Vallejo, R., Gonzalez-Valdivieso, J., Santos, M., Rodriguez-Rojo, S., Arias, F.J., 2021. Production of elastin-like recombinamer-based nanoparticles for docetaxel encapsulation and use as smart drug-delivery systems using a supercritical anti-solvent process. J. Ind. Eng. Chem. 93, 361-374. https://doi.org/ 10.1016/j.jiec.2020.10.013

Vander Heiden, M.G., DeBerardinis, R.J., 2017. Understanding the intersections between metabolism and cancer biology. Cell. 168, 657-669. https://doi.org/10.1016/j.cell.2016.12.039

Vazquez, A., Kamphorst, J.J., Markert, E.K., Schug, Z.T., Tardito, S., Gottlieb, E., 2016. Cancer metabolism at a glance. J. Cell Sci. 129, 3367-3373. https://doi.org/10.1242/jcs.181016

Wagner, V., Dullaart, A., Bock, A.K., Zweck, A., 2006. The emerging nanomedicine landscape. Nat. Biotechnol. 24, 1211-1217. https://doi.org/10.1038/nbt1006-1211

Walkey, C.D., Olsen, J.B., Guo, H., Emili, A., Chan, W.C., 2012. Nanoparticle size and surface chemistry determine serum protein adsorption and macrophage uptake. J. Am. Chem. Soc. 134, 2139-2147. https://doi.org/10.1021/ja2084338

WHO, 2020. http://gco.iarc.fr/today/fact-sheets-cancers (accessed December 5 2020)

WHO, 2020. https://www.who.int/healthinfo/global\_burden\_disease/estimates/en/index1.html (accessed December 5 2020)

Williams, D.F., 2009. On the nature of biomaterials. Biomaterials. 30, 5897-5909. https://doi.org/10.1016/j.biomaterials.2009.07.027

Wolfram, J., Zhu, M., Yang, Y., Shen, J., Gentile, E., Paolino, D., Fresta, M., Nie, G., Chen, C., Shen, H., Ferrari, M., Zhao, Y., 2015. Safety of nanoparticles in medicine. Curr. Drug Targets. 16, 1671-1681. https://doi.org/10.2174/1389450115666140804124808

Wong, X.Y., Sena-Torralba, A., Álvarez-Diduk, R., Muthoosamy, K., Merkoçi, A., 2020. Nanomaterials for nanotheranostics: Tuning their properties according to disease needs. ACS Nano. 14, 2585-2627. https://doi.org/10.1021/acsnano.9b08133

Yang, S.T., Liu, Y., Wang, Y.W., Cao, A., 2013. Biosafety and bioapplication of nanomaterials bydesigningprotein-nanoparticleinteractions.Small.9,1635-1653.https://doi.org/10.1002/smll.201201492

Yousefpour, P., Ahn, L., Tewksbury, J., Saha, S., Costa, S.A., Bellucci, J.J., Li, X., Chilkoti, A., 2019. Conjugate of Doxorubicin to albumin-binding peptide outperforms Aldoxorubicin. Small. 15, e1804452. https://doi.org/10.1002/smll.201804452

Zuckerman, J.E., Gritli, I., Tolcher, A., Heidel, J.D., Lim, D., Morgan, R., Chmielowski, B., Ribas, A., Davis, M.E., Yen, Y., 2014. Correlating animal and human phase Ia/Ib clinical data with CALAA-

01, a targeted, polymer-based nanoparticle containing siRNA. Proc. Natl. Acad. Sci. 111, 11449-11454. https://doi.org/10.1073/pnas.1411393111