Contents lists available at ScienceDirect



Science of the Total Environment

journal homepage: www.elsevier.com/locate/scitotenv



Soil fungal communities under plantations of different *Eucalyptus* species in Ethiopia: Insights for evidence-based management



Gonfa Kewessa^{a,b}, Tatek Dejene^{a,c}, Pablo Martín-Pinto^{a,*}

^a Sustainable Forest Management Research Institute, University of Valladolid, Spain

^b School of Natural Resources, Guder Mamo Mezemir Campus, Ambo University, Ambo, Ethiopia

^c Ethiopian Forestry Development, Addis Ababa, Ethiopia

• Soil fungal communities studied in Ethiopian *Eucalyptus* plantations

Fungal communities differ between

E. globulus and E. camaldulensis stands.

Elevation, precipitation and temperature influence fungal composition.
Plantations provide products and income, reducing pressure on native

• A mosaic *Eucalyptus* species provide fungal diversity at the landscape level.

HIGHLIGHTS

forests.

G R A P H I C A L A B S T R A C T



ARTICLE INFO

Editor: Manuel Esteban Lucas-Borja

Keywords: Eucalyptus plantations Soil fungal diversity Fungal community dynamics Land use impact Sustainable forest management Ethiopia

ABSTRACT

The rapid expansion of *Eucalyptus* plantations in Ethiopia is driven by the increasing demand for woody products, raising concerns about their ecological impact. While conserving native forests remains a priority, Eucalyptus plantations provide alternative sources of forest products and income, helping to reduce pressure on native ecosystems. However, the ecological implications of these plantations, particularly their impact on soil fungal communities, key players in nutrient cycling and ecosystem functioning, remain poorly understood. This study investigates soil fungal community dynamics in Eucalyptus globulus and Eucalyptus camaldulensis plantations across diverse environmental gradients in Ethiopia. Soil samples were collected from 24 plots, and fungal DNA was extracted and sequenced using Illumina MiSeq technology, targeting the ITS2 region. Taxonomic classification and functional guild assignment were performed. Although both plantation types supported a high level of fungal richness and diversity, fungal community composition significantly varied by the Eucalyptus species. Environmental factors, including elevation, precipitation, and temperature, were linked to variations in fungal community composition, creating distinct ecological niches. The main indicator taxa under E. camaldulensis were the species Yurkovia mendeliana, Fusarium oxysporum, Talaromyces solicola, and Westerdykella reniformis, as well as an unidentified member of the class Chytridiomycetes. Under E. globulus, the main indicator taxa were the species Saitozyma podzolica, Brachiosphaera tropicalis, Pseudoacremonium sacchari, and Preussia flanaganii, along with an unidentified member of the order Hypocreales. Although the species Archaeorhizomyces finlayi and members of

* Corresponding author at: Avda. Madrid sn Palencia, Spain. *E-mail address:* pmpinto@uva.es (P. Martín-Pinto).

https://doi.org/10.1016/j.scitotenv.2025.179663

Received 26 March 2025; Received in revised form 8 May 2025; Accepted 11 May 2025 Available online 17 May 2025

0048-9697/© 2025 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY-NC license (http://creativecommons.org/licenses/by-nc/4.0/).

the families Hydnangiaceae and Chaetomiaceae and the order Sordariales were present in both plantation types, their relative abundances differed significantly between the two species. Our findings highlight that expanding *Eucalyptus* plantations support soil fungal diversity. A mosaic landscape combining the two species at the landscape level could enhance fungal biodiversity and ecosystem functionality. Understanding these fungal associations provides valuable insights for evidence-based plantation management and sustainable forestry practices in Ethiopia.

1. Introduction

Ethiopia has a long history of deforestation and land degradation (Mosisa et al., 2024), which has led to severe environmental challenges, particularly in regions where indigenous forests were once abundant. These challenges have been exacerbated by rapid population growth, agricultural land expansion, and the overexploitation of natural resources (Zerga et al., 2021) (Sisay et al., 2024; Tekalign et al., 2018). The extensive deforestation and degradation of natural vegetation have resulted in loss biodiversity loss and declining ecosystem services (EBI, 2022). To meet the increasing demand for forest products while mitigating deforestation, large-scale plantations of fast-growing exotic tree species, particularly *Eucalyptus*, have been widely established across Ethiopia. *Eucalyptus* was first introduced to Ethiopia in 1895 (Belachew and Minale, 2025) and has since become the most dominant plantation species in the country (Abebe and Tadesse, 2014).

More than 0.5 million ha of *Eucalyptus* plantations exist in Ethiopia due to its rapid growth, adaptability to various environmental conditions, and short rotation cycles (Bekele, 2011; Jaleta et al., 2016). More than 55 Eucalyptus species are grown in Ethiopia across diverse ecological zones, with E. globulus and E. camaldulensis being the most widely planted. These species provide essential forest products, contribute to afforestation and soil conservation, and serve as a crucial source of income for local communities (Belachew, 2025; Gil et al., 2010). Additionally, Eucalyptus plantations help reduce pressure on native forests by serving as alternative wood sources (Belachew and Minale, 2025). Despite these perspectives, the knowledge and status of soil fungal communities in Eucalyptus plantations across different agroecological zones in Ethiopia remain largely unexplored. Understanding these fungal dynamics is crucial for developing sustainable plantation forest management strategies, which can support the integration of fungal biodiversity conservation and mushroom production in Eucalyptus-dominated landscapes through a mycosilvicultural approach (Castaño et al., 2019; Dejene et al., 2017a). This approach could also foster ecological resilience while enhancing the economic output of plantation forests in the country (Dejene et al., 2017b).

Despite their economic benefits, the rapid expansion of *Eucalyptus* plantations has raised debates over their ecological effects (Belay et al., 2025; Molla et al., 2023) (Aklilu et al., 2019; Birara Dessie et al., 2019; Getnet et al., 2024; Gil et al., 2010; Kassa et al., 2019; Zerga, 2015). While some studies report negative impacts, including reduced biodiversity, altered soil properties and excessive water drainage, others highlight their potential to support ecological functions when properly managed. A growing body of research suggests that *Eucalyptus* plantations can sustain biodiversity, particularly soil microbial communities, which play fundamental roles in nutrient cycling, organic matter decomposition, and tree resilience (Bekele, 2011; Jaleta et al., 2016; Zhu et al., 2024). However, the influence of different plantations of *Eucalyptus* species on soil fungal communities across different environmental gradients in Ethiopia remains poorly understood.

Soil fungi are key drivers of ecosystem stability and forest productivity, influencing soil nutrient dynamics and plant health (Lombard et al., 2011). The soil fungal composition is strongly shaped by the type of tree species, edaphic and environmental variables which make fungi as an important indicators of ecosystem health and functioning (Tedersoo et al., 2014). Understanding fungal dynamics in *Eucalyptus* plantations is therefore essential for informed management practices. Studies suggest that Eucalyptus grandis plantations in Ethiopia enhance fungal diversity and alter community composition, contributing to belowground biodiversity (Castaño et al., 2019; Dejene et al., 2017b). Moreover, soil fungal communities in plantation forests serve as key indicators of soil quality assessment, as higher fungal diversity is often associated with well-functioning ecosystems (Wu et al., 2019). Despite this, the complex interactions between soil fungi, tree species, and environmental factors remain underexplored, particularly in comparison to aboveground ecosystem components. While some research highlights the role of Eucalyptus plantations in biodiversity rehabilitation (Moges et al., 2010), the extent to which different Eucalyptus species influence fungal diversity and composition is largely unknown. Studying the soil fungal diversity can provide valuable insights into the overall health and sustainability of plantation forests (Hereira-Pacheco et al., 2025). Addressing this gap is crucial for ensuring the sustainable management of plantation forests in Ethiopia (Dejene et al., 2017a; Kewessa et al., 2022; Moges et al., 2010).

Given the ecological importance of soil fungi and the increasing expansion of *Eucalyptus* plantations in Ethiopia, it is essential to explore how Eucalyptus species and environmental factors influence soil fungal communities across different site conditions. We aim to address this gap by examining the composition and diversity of soil fungi associated with two species of Eucalyptus that are commonly planted in Ethiopia: E. globulus and E. camaldulensis. We hypothesize that: (i) both Eucalyptus species support similar levels of fungal diversity owing to shared ecological traits, such as plantation structure and leaf litter production. The overall diversity levels are likely to be comparable owing to similar resource availability and plantation management practices. Despite this, we anticipate that: (ii) fungal community composition among Eucalyptus species varies because the composition is driven by species-specific traits, such as leaf chemistry, root architecture, or nutrient cycling. Finally, we hypothesize that: (iii) environmental factors such as mean annual precipitation (MAP), mean annual temperature (MAT), and site variables (i.e., elevation, longitude, and latitude) shape fungal communities by affecting soil parameters. Thus, the specific objectives of this study are: (i) to determine the soil fungal diversity associated with E. globulus and E. camaldulensis; (ii) to analyze and compare the composition of soil fungal taxa in areas planted with E. globulus and E. camaldulensis; and (iii) to elucidate environmental variables shaping the fungal communities associated with these Eucalyptus plantations. Such a comprehensive assessment of soil fungal communities in Eucalyptus plantations should contribute to a deeper understanding of the ecological implications of planting Eucalyptus plantations in Ethiopia. Our findings should inform sustainable forest management practices and guide policymakers in the development of strategies to mitigate potential negative impacts on soil health and biodiversity while maximizing the ecological and economic benefits of Eucalyptus plantations.

2. Materials and methods

2.1. The study area

This study was conducted in *Eucalyptus*-dominated plantations in Ethiopia, selected for their ecological and economic significance. A preliminary survey identified areas with extensive *Eucalyptus* plantations, leading to the selection of *E. globulus* and *E. camaldulensis* as the focus species. The chosen plantations share similar climatic conditions,

minimizing environmental variability and allowing for meaningful comparisons of fungal communities.

Detailed site-specific characteristics, including geographic coordinates, elevation, climate, and soil properties, are provided in Table 1, while the study area map (Fig. 1) highlights the exact locations of the sampled sites, offering a visual representation of the study's spatial context.

2.2. Sampling procedure and environmental data collection

Eight plantations (i.e., three E. camaldulensis plantations and five *E. globulus* plantations) were selected for soil fungal sampling, with three transects established in each plantation. Each transect was spaced 200 m apart, and rectangular plots (2 m \times 50 m) were laid out systematically at 100-m intervals to avoid spatial confounding (Dejene et al., 2017b; Hijesalu et al., 2017; Rudolph et al., 2018). A total of 24 plots (three plots per plantation) were established for soil sampling. Soil samples were collected between August 24 and 26, 2023. The litter layer was removed before sampling to focus on soil fungal communities (Vořiškova et al., 2014). Five soil cores (0-15 cm depth) were collected from the corners and center of each plot using a soil corer (radius, 2 cm; depth, 20 cm) as described in previous studies (Alem et al., 2020; Castaño et al., 2019; Chen et al., 2022; Dejene et al., 2017a; Kewessa et al., 2022; Liu et al., 2021; Zhang et al., 2021). These cores were pooled to form composite samples comprising approximately 500 g of soil per plot. The systematic sampling design ensured consistency while accounting for site-specific variations.

The composite soil samples were dried, sieved (2-mm mesh), and ground into a fine powder for chemical analysis and were frozen immediately and kept at -20 °C until DNA was extracted. Key soil properties, including organic carbon (OC), total nitrogen (N), available phosphorus (P), and pH, were analyzed using standard methods. Specifically, soil OC was assessed using wet digestion (Walkley and Black, 1934). N was measured using the Kjeldahl method (Kim et al., 2005). P was calculated following the standard procedure described by Tan (2005). A pH meter was used to analyze a soil: water (1:2.5) suspension to determine the pH of the soil samples (Reeuwijk, 2002).

Climate data, including temperature and precipitation, were obtained from nearby meteorological stations, while geographical coordinates (latitude and longitude), elevation, and aspect were recorded in the field using a GPS device at each sampled plot (Table 1). These climate variables, combined with site-specific topographical data,

Table 1

The mean values of environmental data and soil characteristics of eac	h study site
---	--------------

Plantation species

provided valuable insights into the environmental factors shaping fungal community dynamics.

2.3. DNA extraction and metagenomic sequencing

To analyze the soil fungal communities, DNA was extracted from the collected composite soil samples using the DNeasy PowerSoil kit (Qiagen) and following the manufacturer's protocol. For each extraction, 0.25 g of soil was used. The extracted DNA was subsequently used for fungal metagenomic analysis, focusing on the amplification of the Internal Transcribed Spacer 1 (ITS1) region, which is a widely accepted marker for fungal community studies, ensuring comprehensive coverage of diverse fungal taxa. Metagenomic sequencing was conducted on the Illumina iSeq 100 platform, utilizing a 2×150 bp configuration. The ITS1 region was amplified using a set of forward and reverse primers designed for this region, which allowed for robust amplification of diverse fungal taxa. The forward primer set consisted of the following sequences:

- ITS_fwd_1 with the sequence CTTGGTCATTTAGAGGAAGTAA;
- ITS_fwd_2 with the sequence CTCGGTCATTTAGAGGAAGTAA;
- ITS fwd 3 with the sequence CTTGGTCATTTAGAGGAACTAA;
- ITS fwd 4 with the sequence CCCGGTCATTTAGAGGAAGTAA;
- ITS fwd 5 with the sequence CTAGGCTATTTAGAGGAAGTAA;
- ITS_fwd_6 with the sequence CTTAGTTATTTAGAGGAAGTAA;
- ITS_fwd_7 with the sequence CTACGTCATTTAGAGGAAGTAA;
- ITS_fwd_8 with the sequence CTTGGTATTTAGAGGTCGTAA.

The reverse primer set consisted of the following sequences:

- ITS_rev_1 with the sequence GCTGCGTTCTTCATCGATGC;
- ITS_rev_2 with the sequence GCTGCGTTCATCGATGG;
- ITS_rev_3 with the sequence GCTACGTTCTTCATCGATGC;
- ITS_rev_4 with the sequence GCTGCGTTCTTCATCGATGT;
- ITS_rev_5 with the sequence ACTGTGTTCTTCATCGATGC;
- ITS rev 6 with the sequence GCTGCGTTCTTCATCGTTGC;
- ITS rev 7 with the sequence GCGTTCTTCATCGATGC.

Following initial amplification, a second PCR was performed to incorporate Illumina sequencing adapters and dual-index barcodes using Nextera[™] XT v2 indices. AMPure XP beads were used after both PCR steps to clean the amplicons, ensuring high-quality DNA libraries.

	Eucalyptus globulus					Eucalyptus camaldulensis		
Site	Gafarsa	Ciri	Dabana 1	Dabana 2	Sidisa	Birbirsa	Odabari	Kality
Elevation (m asl)	2604.33	2397.33	1883.00	1649.67	2010.00	2156.33	2006.00	2174.67
Latitude (N)	9°04′	9°03′	8°41′	8° 42′	8°45′	8°54′	8°57′	8°54′
Longitude (E)	38° 37′	38°25′	36°30′	36°42′	36°34′	37°44′	37°45′	38° 46′
MAP (mm)	1188.00	1100.00	1100.00	1100.00	1100.00	950.00	950.00	1874.00
MAT (°C)	14.00	18.50	15.39	15.39	15.39	19.50	19.50	15.60
Sand (%)	61.00	59.67	66.33	69.00	56.33	47.00	43.67	55.00
Silt (%)	25.67	28.33	19.67	19.00	29.67	34.33	35.67	27.67
Clay (%)	13.33	12.00	14.00	12.00	14.00	18.67	20.67	17.33
рН	5.76	5.82	5.58	5.42	5.16	5.52	5.13	5.48
EC (ppm)	0.66	0.88	0.79	1.31	0.92	0.53	0.43	0.54
Na (ppm)	2.07	2.21	1.98	2.31	2.13	2.29	2.48	2.10
K (ppm)	0.69	0.66	0.98	0.67	0.66	0.85	0.59	0.53
Ca (ppm)	12.74	13.97	13.52	13.13	7.77	11.12	9.21	7.83
Mg (ppm)	5.40	5.65	5.21	5.12	4.25	5.75	4.42	3.90
CEC (ppm)	44.24	44.27	41.13	40.47	41.20	47.13	34.95	32.33
OC (ppm)	2.35	2.61	2.61	2.57	2.13	2.12	1.19	2.16
N (ppm)	0.16	0.19	0.23	0.19	0.16	0.14	0.07	0.13
P (ppm)	7.86	7.55	5.84	6.44	8.48	8.81	8.17	1036

Abbreviations: MAP, mean annual precipitation; mean annual temperature, MAT; EC, exchange capacity; CEC, cation exchange capacity; OC, organic carbon.



Fig. 1. Map of Ethiopia showing the location of the study areas.

Libraries were quantified, normalized, and pooled before sequencing. To enhance data quality, 20 % of PhiX sequencing controls were added to the pooled libraries. Sequencing was performed using the iSeq v2 reagent kit (Illumina, CA, USA) at Biotecnología Forestal Aplicada, Palencia, Spain, following the manufacturer's instructions for 300-cycle paired-end sequencing.

2.4. Taxonomic classification and functional group assignment

For taxonomic identification, the Illumina BaseSpace 16S Metagenomics App was utilized. This platform employs the UNITE fungal ITS sequence database (v7.2) for taxonomic classification, ensuring the accurate assignment of sequences to taxonomic groups. Sequences were classified by pairwise similarity searches against curated fungal ITS sequences, with taxonomic assignments based on Species Hypothesis (SH) groups, as defined by Kõljalg et al. (2013). The UNITE fungal database includes identified fungal sequences and ecological functions, facilitating robust taxonomic classification and functional group analysis. Operational Taxonomic Units (OTUs) with \geq 90 % similarity to fungal SHs with known ecological functions were assigned to functional groups using the FungalTraits database (Põlme et al., 2020). The assignment process was facilitated via the PlutoF web workbench (Abarenkov et al., 2010).

2.5. Statistical analysis

We conducted all statistical analyses utilizing the sequence count for each OTU as the abundance value and following the method by Danzeisen et al. (2011) for non-singleton fungal communities. The data used for relative abundance and Shannon diversity index analyses met the parametric criteria of normality and homoscedasticity and, thus, no transformations were necessary. Evenness was log-transformed, and richness was Box-Cox optimum transformed. Differences among treatments were evaluated using linear mixed-effects (LME) models (Pinheiro et al., 2012), with the plot designated as a random effect and the plantation type as a fixed effect. The adequacy of these mixed models was verified through graphical inspections and the Shapiro–Wilk test to confirm the normality and homoscedasticity of residuals. All data analyses were performed using R version 4.2.2 (R Core Team, 2022).

We performed a Permutational Multivariate Analysis of Variance (PerMANOVA) using the adonis2 function to determine the effect of Eucalyptus species on the distribution of the soil fungal community composition, which was determined based on the Bray-Curtis dissimilarity after excluding singleton OTUs. We used a Hellinger transformed community matrix and environmental scaled data for the analysis. Prior to PerMANOVA, we used betadisper to avoid misinterpretation of potential differences observed. The distribution was visualized using Nonmetric Multidimensional Scaling (NMDS) and the analysis of correlation with environmental data was performed using the envfit function in R (R Core Team 2022). We performed NMDS analyses for total fungi and fungal guilds. Finally, we used multilevel pattern analysis using the multipatt function in R to assign taxa that were significantly associated with each treatment. Fungal community composition was further visualized through a taxonomic hierarchy using Krona charts (Ondov et al., 2011). We used the ordisurf function in R to fit a Generalized Additive Model with a Gaussian error family. This allowed us to represent, within

NMDS, variables that were most statistically correlated with the fungal community composition.

3. Results

3.1. Taxonomic identification

In total, 841,553 high-quality filtered reads that were classified as fungi were obtained from the sequenced soil samples. Between 15,437 and 67,943 high-quality reads were obtained from each soil sample. Across the eight plantations, 3824 fungal species belonging to six distinct fungal phyla were detected. The majority of the identified fungal species belonged to either Ascomycota (58 %) or Basidiomycota (26 %). Other phyla included Mortierellomycota (10 %) and Kickxellomycota (3 %). However, approximately 3 % of the total recorded fungal species could not be assigned to a specific fungal phylum (Fig. 2).

Within the phylum Ascomycota, several genera were particularly abundant, with *Microidium* accounting for 11 % of the community, followed by *Penicillium* (6 %), *Chaetomium* (5 %), and *Trichoderma* (5 %) (Fig. 3). Other notable genera, such as *Fusarium* (4 %), were abundant across the eight plantations. The chart also shows the presence of a large number of lesser-known fungal taxa, many of which remained unidentified (4 %), reflecting the unexpectedly high diversity and complexity of fungal species associated with *Eucalyptus* plantations.

Laccaria was the most abundant genus within the Basidiomycota phylum (Fig. 4), accounting for 24 % of the Basidiomycota community, followed by the genus *Saitozyma* (18 %). The hierarchal analysis also

highlighted other diverse types of genera, including *Descomyces* (5 %) and *Boletus* (3 %) (Fig. 5), suggesting a complex fungal community with various ecological roles. However, 17 % of the Basidiomycota taxa remained unidentified (Fig. 4).

3.2. Effect of Eucalyptus species on the abundance and diversity of soil fungal taxa

The soil fungal communities detected in *E. globulus* and *E. camaldulensis* plantations did not differ significantly in terms of fungal relative abundance (p = 0.108), the Shannon–Wiener index, (H') (p = 0.906), richness (p = 0.279), evenness (p = 0.352) (Fig. 5), or fungal guilds (Fig. 6).

3.3. Effect of Eucalyptus species on the soil fungal community composition

The overall fungal community composition detected in *E. globulus* plantations was significantly different from that in *E. camaldulensis* plantations (F = 1.8752, $R^2 = 0.07854$, p = 0.013) (Fig. 7A). The low-stress value obtained in the NMDS analysis indicated a relatively good fit of the data. Elevation, latitude, longitude, MAP, potassium (K), and cation exchange capacity (CEC) were significantly correlated with fungal composition, while phosphorus (P) exhibited a marginal correlation. By contrast, MAT was not significantly correlated with fungal community composition (Table 2; Fig. 7A).

The distribution of guilds in E. globulus plantations did not differ



Fig. 2. Taxonomic hierarchy of the soil fungal community associated with plantations of *Eucalyptus* species across different landscapes in Ethiopia. The Krona chart represents the fungal community as a taxonomic hierarchy up to the genus level. The circles and sizes are proportional to the number of fungal species belonging to each phylum, and colors represent absolute taxon abundance (sum of read number).



Fig. 3. Taxonomic hierarchy of the soil fungal community belonging to the phylum Ascomycota associated with plantations of *Eucalyptus* species across different landscapes in Ethiopia. The Krona chart represents the Ascomycota fungal community composition grouped up to genus level. The circles and sizes are proportional to the number of fungal species within each order, and colors represent absolute taxon abundance (sum of read number).

significantly from that in *E. camaldulensis* plantations ($R^2 = 0.0349$, F = 0.7959, p = 0.528). The low-stress value obtained in the NMDS analysis indicated a good fit of guild data (Fig. 7B), with stress type 1 and β dispersion (F = 0.000, p = 0.977). K and CEC were found to be significantly correlated with fungal guilds in *E. globulus* and *E. camaldulensis* plantations (Table 1). Geographic and environmental variables, such as latitude, longitude, and elevation, emerged as dominant drivers of fungal community composition, explaining nearly half (49.4 %) of the variation in fungal community structure. Moreover, climatic factors, including MAP and MAT, also contributed (26.3 %) to fungal community composition,

3.4. Indicator fungal species and guilds

In total, 3003 OTUs were associated with *E. globulus* plantations, of which 40 showed significant associations with this plantation type. By contrast, among the 2656 indicator fungal taxa linked to *E. camaldulensis* plantations, 147 were significantly associated with this plantation type (Table S1). The ten most contributing taxa for each type of *Eucalyptus* plantation are shown in Fig. 8. However, none of the guilds were significantly associated with either *E. globulus* or *E. camaldulensis* plantations. Although *Archaeorhizomyces finlayi* and species of

Hydnangiaceae, Sordariales, and Chaetomiaceae were present in both plantation types, their relative abundances differed significantly between the two species.

4. Discussion

Sequencing analysis revealed a high diversity of fungal species in soil samples collected from plantations of Eucalyptus tree species. Furthermore, the estimated range of high-quality reads suggests variability in fungal abundance, which could be influenced by both soil conditions and the Eucalyptus tree species in the study areas. This finding in general aligns with studies performed by Dejene et al. (2017a) and Castaño et al. (2019), who reported that Ethiopian Eucalyptus grandis plantations support a diverse range of fungal species, based on their analyses of sporocarp diversity and soil fungal profiles, respectively. Our findings further highlight the role of Eucalyptus plantations in fostering fungal biodiversity, challenging the traditional perspective that Eucalyptus has a negative ecological impact on these ecosystems (FAO, 2011; Jaleta et al., 2016; Teketay, 2000). Although concerns about Eucalyptus plantations remain, there is a growing body of evidence indicating that they may have some unexpected ecological benefits. For example, Eucalyptus plantations have been reported to support diverse herbaceous species



Fig. 4. Taxonomic hierarchy of the soil fungal composition associated with *Eucalyptus* plantations belonging to the phylum Basidiomycota across different landscapes in Ethiopia. The Krona chart represents the Basidiomycota community composition grouped up to genus level. The circles and sizes are proportional to the number of fungal species within each order, and colors represent absolute taxon abundance (sum of read number).



Fig. 5. Total abundance and diversity of the total fungal community detected in *Eucalyptus globulus* and *Eucalyptus camaldulensis* plantations. Box-and-whisker plots indicate the data range; horizontal lines indicate the median.

(Moges et al., 2010), facilitate native flora regeneration (Lemenih and Teketay, 2005; Yirdaw and Luukkanen, 2003), and harbor a rich diversity of fungal species, including economically valuable mushroom species (Castaño et al., 2019; Dejene et al., 2017b). Such findings call for a reconsideration of the role of *Eucalyptus* in Ethiopia's forestry landscape, revealing its potential as a multifunctional resource. Thus, by integrating mycosilvicultural practices into the forest management of *Eucalyptus* plantations, *Eucalyptus* species could be used as multifunctional landscape species that simultaneously support timber production,

biodiversity conservation, and the sustainable cultivation of valuable edible mushrooms (Castaño et al., 2019; Dejene et al., 2017b). This approach would not only address environmental concerns but also create economic opportunities for local communities, promoting more balanced and sustainable forestry management strategies (Castellano and Molina, 1989; Trappe, 1977).

Laccaria was the most abundant ectomycorrhizal species detected in this study. The dominance of *Laccaria* in our study forests is particularly advantageous because *Laccaria* enhances plant survival and growth on



Fig. 6. The relative abundance of the total fungal guilds detected in *Eucalyptus globulus* and *Eucalyptus camaldulensis* plantations. Box-and-whisker plots indicate the range of the data; horizontal lines indicate the median; and dots indicate outliers.

nutrient-poor and degraded soils, which are common in Ethiopia and, therefore, could promote the rehabilitation and restoration of degraded forest lands. Furthermore, as edible fungi, *Laccaria* species have significant economic potential, thereby increasing the value of *Eucalyptus* plantations, and are becoming an important source of rural income (Abate, 2008; Boa, 2004).

The soil fungal communities detected in *E. globulus* plantations did not differ significantly from those detected in *E. camaldulensis* plantations in terms of fungal abundance, diversity, richness, or evenness when analyzing the total community or when grouping by fungal guilds. However, the dominance of some fungal species in the soil of both types of plantations could also contribute to the lack of significant variation in their diversity indices (Dahlberg, 2002, Kipfer et al., 2010). In addition, factors other than *Eucalyptus* species, such as soil conditions, environmental variables, or forest management practices might have a greater effect on the richness and diversity of soil fungal communities (Drenovsky et al., 2004, Lauber et al., 2009). Therefore, further research is needed to better understand the dynamics and characteristics of these soil fungal communities.

However, as expected, the overall soil fungal community composition of E. globulus plantations did differ significantly from that of E. camaldulensis plantations. This difference could be due to differences in environmental variables and landscape heterogeneity (Bahram et al., 2015; Ferrari et al., 2016; Peay et al., 2010; Tedersoo et al., 2014). The ordination analysis showed that spatial factors such as elevation and longitude, as well as climate and edaphic variables, highly influenced the soil fungal assembly in Eucalyptus plantations. This may be an indication that the combined effects of these variables (Li et al., 2020) affected the spatial variation of the fungal community (Chen et al., 2015) in our study areas. Climate conditions and edaphic variables, such as temperature, rainfall, humidity, and elevation, influence fungal distribution because different fungi have specific environmental preferences and some species may not exist in certain plantations (Li et al., 2020). In addition, the physiology of Eucalyptus species, including root exudates and leaf chemistry, affects fungal associations, with some fungi exhibiting host specificity. Different species, in our case Eucalyptus, secrete distinct profiles of root exudates, including sugars, which selectively attract or inhibit specific fungal taxa (Sasse et al., 2018;



Fig. 7. The relationship between the distribution of the total fungal community (A) and fungal guilds (B) and correlated environmental factors in plantations of *Eucalyptus globulus* or *Eucalyptus camaldulensis*. Visualization is based on nonmetric multidimensional scaling (NMDS) using Euclidean distance. The polygon represents the 95 % confidence intervals surrounding each group. Vector names represent significantly correlated environmental factors. Green lines represent a Generalized Additive Model with a Gaussian error family for the most statistically correlated variables. Mean annual precipitation (MAP) for the total community in A, and elevation for fungal guilds in B. CEC, cation exchange capacity.

 Table 2

 Variables significantly correlated with the soil fungal community composition.

Community	Study site variables	NMDS1	NMDS2	r ²	<i>p</i> -Value
Total fungi	Elevation	0.6635	0.7482	0.3437	0.015
	Latitude	0.5760	0.8175	0.4249	0.006
	Longitude	0.2476	0.9689	0.3963	0.006
	MAP	-0.7410	0.6715	0.2696	0.011
	К	0.4236	-0.9035	0.0268	0.047
	CEC	0.9659	-0.2586	0.3122	0.027
	Р	-0.4652	0.8852	0.2219	0.067
Guilds	Elevation	0.8527	0.5224	0.2258	0.063
	K	0.3107	-0.9505	0.2327	0.061
	CEC	0.8449	0.5350	0.2453	0.061

Abbreviations: MAP, mean annual precipitation; CEC, cation exchange capacity; K, potassium; P, phosphorus; NMDS, non-metric multidimensional scaling. Bold: significant correlation.

Walker et al., 2003). These chemical differences might be leading to the difference in fungal community composition even if overall diversity metrics are similar. Furthermore, given that the study sites were in

different regions, geographic isolation and dispersal limitations would likely further influence fungal composition given that spore movement and colonization potential varies across landscapes (Tedersoo et al., 2014).

More than three times the number of fungal species were significantly correlated with *E. camaldulensis* plantations than with *E. globulus* plantations. On the one hand, this could reflect the distinct ecological niches created by each of the *Eucalyptus* species. On the other hand, many fungal taxa were detected in both *E. camaldulensis* and *E. globulus* plantations. Thus, given the importance of *Eucalyptus* species for fungal diversity promotion, planting different *Eucalyptus* species at separate sites and then combining them at the landscape level would enhance fungal richness and biodiversity.

In this study, the edaphic element of the Potassium (K) was found to correlate with the overall soil fungal community, suggesting that soil cation concentrations may influence fungal community development. Because cations, including the cation exchange capacity (CEC), are involved in various physicochemical processes such as photosynthesis (Shi et al., 2014), which can impact plant photosynthesis and,



Fig. 8. The number of total soil fungal taxa and the ten most contributing taxa associated with plantations of Eucalyptus globulus and Eucalyptus camaldulensis.

Science of the Total Environment 982 (2025) 179663

consequently, the carbon available to soil fungi (Shi et al., 2014). Also, the correlation between phosphorus (P) and fungal taxa aligns with previous studies by Kranabetter et al. (2009); Reverchon et al. (2010), which highlighted the influence of P on fungal distribution patterns. In this study also observed that fungal communities often specialized based on guilds in soils with higher potassium (K) levels. This could be because K-rich environments offer more favorable conditions for certain fungal species that thrive in nutrient-rich soils, allowing them to dominate or establish stable communities (Trudell and Edmonds, 2004). Furthermore, potassium's presence may affect soil pH and other factors that contribute to the specialization of fungal communities in such areas (Li et al., 2020).

5. Conclusions

In this study we have found that *Eucalyptus globulus* and *Eucalyptus camaldulensis* plantations support high and comparable levels of soil fungal diversity in Ethiopia. However, despite similarities in overall abundance, richness, and evenness, the composition of fungal communities differed significantly between the two species. These differences are likely influenced by species-specific factors such as variation in root exudate chemistry, in addition to the environmental variables such as the elevation, temperature, and precipitation. *E. globulus* stands were dominated by distinct fungal taxa compared to *E. camaldulensis* stands, indicating that each tree stands fosters a unique fungal assemblage in the rhizosphere. At the landscape level, managing both species can enhance overall fungal diversity, contributing to soil health, ecosystem resilience, and sustainable plantation management.

While the conservation of native forests remains a priority, *Eucalyptus* plantations, when strategically managed, can contribute to fungal biodiversity and ecosystem health. To maximize biodiversity benefits while reducing pressure on native forests, a mosaic landscape approach should be used, where different *Eucalyptus* species are planted at separate sites and integrated at the landscape level. Moreover, given the importance of soil fungi in maintaining soil fertility and forest productivity, sustainable plantation management should consider fungal dynamics when planning afforestation and reforestation programs. To ensure the long-term productivity of *Eucalyptus* plantations, further research is needed to explore their ecological impacts over extended periods. Understanding these dynamics will provide crucial insights into optimizing plantation forestry for both economic and ecological sustainability, ultimately contributing to resilient forest ecosystems in Ethiopia and beyond.

Supplementary data to this article can be found online at https://doi.org/10.1016/j.scitotenv.2025.179663.

CRediT authorship contribution statement

Gonfa Kewessa: Writing – review & editing, Writing – original draft, Methodology, Investigation, Formal analysis. Tatek Dejene: Writing – review & editing, Project administration, Conceptualization. Pablo Martín-Pinto: Writing – review & editing, Methodology, Investigation, Funding acquisition, Formal analysis, Conceptualization.

Funding

This research was supported by the project SUSTFUNGI_ET III: 2022/ ACDE/000201 funded by the Spanish Agency for International Development and Cooperation. The first author received research funds through the research contracts of the UVa 2021, co-financed by Banco Santander.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

We would like to express our gratitude to the people who, in one way or another, contributed to the success of this work.

Data availability

Data will be made available on request.

References

- Abarenkov, K., Tedersoo, L., Nilsson, R.H., Vellak, K., Saar, I., Veldre, V., 2010. PlutoF a web based workbench for ecological and taxonomic research, with an online implementation for fungal ITS sequences. Evol. Bioinform. 6, 189–196. https://doi. org/10.4137/EBO.S6271.
- Abate, D., 2008. Wild mushooms in Ethiopia and our eating habit. In: National Mushroom Conference, Faculty of Science, Addis Ababa University, May 14–15. Addis Ababa, Ethiopia.
- Abebe, M., Tadesse, W., 2014. Eucalyptus in Ethiopia: risk or opportunity? http://www. eiar.gov.et.
- Aklilu, B.M., Bekele, L., Merkineh, M.M., Barana, 2019. Is the expansion of Eucalyptus tree a curse or an opportunity? Implications from a dispute on the trees ecological and economic impact in Ethiopia: a review. J. Ecol. Nat. Environ. 11, 75–83. https:// doi.org/10.5897/jene2019.0765.
- Alem, D., Dejene, T., Oria-de-Rueda, J.A., Geml, J., Martín-Pinto, P., 2020. Soil fungal communities under *Pinus patula* Schiede ex Schltdl. & Cham. plantation forests of different ages in Ethiopia. Forests 11 (10), 1109. https://doi.org/10.3390/ f11101109.
- Bahram, M., Peay, K.G., Tedersoo, L., 2015. Local-scale biogeography and spatiotemporal variability in communities of mycorrhizal fungi. New Phytol. 205, 1454–1463. https://doi.org/10.1111/nph.13206.
- Bekele, M., 2011. Forest plantation and woodlots in Ethiopia. African forest forum working paper series. African Forest Forum, Nairobi, Kenya. Afr. For. Forum Work. Pap. Ser. 12 (1), 1–56.
- Belachew, K.G., 2025. Socioeconomic and Environmental Impacts of Eucalyptus Plantations in Ethiopia: An Evaluation of Benefits, Challenges, and Sustainable Practices 2025. https://doi.org/10.1155/tswj/1780293.
- Belay, F., Mulugeta, M., Makonnen, T., 2025. *Eucalyptus*-based livelihoods: enhancing household food security and resilience in Northwest Ethiopia. Front. Sustain. Food Syst. 9. https://doi.org/10.3389/fsufs.2025.1496756.
- Birara Dessie, A., Mire Abate, T., Melese Mekie, T., 2019. Eucalyptus: the popular exotic tree crop in Ethiopia. Acta Sci. Agric. 3, 50–56. https://doi.org/10.31080/ asag.2019.03.0607.
- Boa, E., 2004. Wild Edible Fungi: A Global Overview of Their Use and Importance to People, Non-wood Forest Products. FAO, Rome, p. 17.
- Castaño, C., Dejene, T., Mediavilla, O., Geml, J., Andres, J., Oria-de-Rueda, Martín-Pinto, P., 2019. Changes in fungal diversity and composition along a chronosequence of *Eucalyptus grandis* plantations in Ethiopia. Fungal Ecol. 39, 328–335. https://doi. org/10.1016/j.funeco.2019.02.003.
- Castellano, M., Molina, R., 1989. Mycorrhizae. In: Landis, T.D., Tinus, R.W., McDonald, S.E., Barnett, J.P. (Eds.), The Biological Component: Nursery Pests and Mycorrhizal. Vol. 5. The Container Tree Nursery Manual. Agriculture Handbook. USDA, Forest Service, Washington, DC, p. 674, 171 p.
- Chen, J., Shi, Z., Liu, S., Zhang, M., Cao, X., Chen, M., Xu, G., Xing, H., Li, F., Feng, Q., 2022. Altitudinal variation influences soil fungal community composition and diversity in alpine–gorge region on the eastern Qinghai–Tibetan plateau. J. Fungi 8 (8), 807. https://doi.org/10.3390/jof8080807.
- Chen, Z., Yu, G., Ge, J., Wang, Q., Zhu, X., Xu, Z., 2015. Roles of climate, vegetation and soil in regulating the spatial variations in ecosystem carbon dioxide fluxes in the northern hemisphere. PLoS One 10, e0125265. https://doi.org/10.1371/journal. pone.0125265.
- Dahlberg, A., 2002. Effects of fire on ectomycorrhizal fungi in fennoscandian boreal forests. Silva Fenn 36, 69–80. https://doi.org/10.14214/sf.551.
- Danzeisen, J.L., Kim, H.B., Isaacson, R.E., Tu, Z.J., Johnson, T.J., 2011. Modulations of the chicken cecal microbiome and metagenome in response to anticoccidial and growth promoter treatment. PloS One 6 (11), e27949.
- Dejene, T., Oria-de-rueda, J.A., Martín-Pinto, P., 2017a. Fungal diversity and succession following stand development in *Pinus patula* Schiede ex Schltdl. & Cham. plantations in Ethiopia. For. Ecol. Manag. 395, 9–18. https://doi.org/10.1016/j. foreco.2017.03.032.
- Dejene, T., Oria-de-Rueda, J.A., Martín-Pinto, P., 2017b. Fungal diversity and succession under *Eucalyptus grandis* plantations in ethiopia. For. Ecol. Manag. 405, 179–187. https://doi.org/10.1016/j.foreco.2017.08.050.
- Drenovsky, R., Vo, D., Graham, K., Scow, K., 2004. Soil water content and organic carbon availability are major determinants of soil microbial community composition. Microb. Ecol. 48, 424–430.
- EBI, 2022. National Ecosystem Assessment of Ethiopia: Syntheses of the Status of Biodiversity and Ecosystem Services, and Scenarios of Change.

- FAO. 2011. Eucalyptus in East Africa, socio-economic and environmental issues, by Dessie, Gessesse, Erkossa, Teklu. Planted Forests and Trees Working Paper 46/E, Forest Management Team, Forest Management Division. FAO, Rome.
- Ferrari, B.C., Bissett, A., Snape, I., van Dorst, J., Palmer, A.S., Ji, M., Siciliano, S.D., Stark, J.S., Winsley, T., Brown, M.V., 2016. Geological connectivity drives microbial community structure and connectivity in polar, terrestrial ecosystems. Environ. Microbiol. 18, 1834–1849. https://doi.org/10.1111/1462-2920.13034.
- Getnet, K., Zegeye, R., Azene, F., Sharma, D., Ayele, T. 2024. Impacts of *Eucalyptus globulus* plantation on soil properties in the northwestern highlands of Ethiopia. 10. 20944/preprints202405.1336.v1.
- Gil L. W T. E T. R L. 2010 *Eucalyptus* species management, history, status and trends in Ethiopia. Proceedings from the Congress Held in Addis, *Eucalyptus* Species Management, History, Status and Trends in ethiopia.
- Hereira-Pacheco, S., Razo, I.A., Miranda-carrazco, A., Dendooven, L., 2025. Metagenomic analysis of fungal assemblages at a regional scale in high-altitude temperate forest soils: alternative methods to determine diversity, composition and environmental drivers. PeerJ 13, e18323. https://doi.org/10.7717/peerj.18323.
- Hiiesalu, I., Bahram, M., Tedersoo, L., 2017. Plant species richness and productivity determine the diversity of soil fungal guilds in temperate coniferous forest and bog habitats. Mol. Ecol. 26, 4846–4858. https://doi.org/10.1111/mec.14246.
- Jaleta, D., Mbilinyi, B., Mahoo, H., Lemenih, M., 2016. Eucalyptus expansion as relieving and provocative tree in Ethiopia. J. Agric. Ecol. Res. Int. 6, 1–12. https://doi.org/ 10.9734/JAERI/2016/22841.
- Kassa, G., Molla, E., Abiyu, A., 2019. Effects of Eucalyptus camaldulensis tree plantation on indigenous trees and soil properties in North Western Ethiopia. Abyssinia J. Sci. Technol. 4, 1–10.
- Kewessa, G., Dejene, T., Alem, D., Tolera, M., Martín-Pinto, P., 2022. Forest type and site conditions influence the diversity and biomass of edible macrofungal species in Ethiopia. J. Fungi 8, 1023. https://doi.org/10.3390/jof8101023.
- Kim, J., Kreller, C.R., Greenberg, M.M., 2005. Preparation and analysis of oligonucleotides containing the C4'-oxidized abasic site and related mechanistic probes. J. Organomet. Chem. 70, 8122–8129. https://doi.org/10.1021/jo0512249.
- Kipfer, T., Egli, S., Ghazoul, J., Moser, B., Wohlgemuth, T., 2010. Susceptibility of ectomycorrhizal fungi to soil heating. Fungal Biol. 114, 467–472. https://doi.org/ 10.1016/j.funbio.2010.03.008.
- Kõljalg, U., Nilsson, R.H., Abarenkov, K., Tedersoo, L., Taylor, A.F.S., Bahram, M., Bates, S.T., Bruns, T.D., Bengtsson-Palme, J., Callaghan, T.M., Douglas, B., Drenkhan, T., Eberhardt, U., Dueñas, M., Grebenc, T., Griffith, G.W., Hartmann, M., Kirk, P.M., Kohout, P., Larsson, E., Lindahl, B.D., Lücking, R., Martín, M.P., Matheny, P.B., Nguyen, N.H., Niskanen, T., Oja, J., Peay, K.G., Peintner, U., Peterson, M., Pöldmaa, K., Saag, L., Saar, I., Schüßler, A., Scott, J.A., Senés, C., Smith, M.E., Suija, A., Taylor, D.L., Telleria, M.T., Weiss, M., Larsson, K.H., 2013. Towards a unified paradigm for sequence-based identification of fungi. Mol. Ecol. https://doi.org/10.1111/mec.12481.
- Kranabetter, J.M., Durall, D.M., MacKenzie, W.H., 2009. Diversity and species distribution of ectomycorrhizal fungi along productivity gradients of a southern boreal forest. Mycorrhiza 19, 99–111. https://doi.org/10.1007/s00572-008-0208-z.
- Lauber, C.L., Hamady, M., Knight, R., Fierer, N., 2009. Pyrosequencing based assessment of soil pH as a predictor of soil bacterial community structure at the continental scale. Appl. Environ. Microbiol. 75, 5111–5120.
- Lemenih, M., Teketay, D., 2005. Effect of prior land use on the recolonization of native woody species under plantation forests in the highlands of Ethiopia. For. Ecol. Manag. 218, 60–73. https://doi.org/10.1016/j.foreco.2005.07.010.
 Li, P., Li, W., Dumbrell, A.J., Liu, M., Li, G., Wu, M., Jiang, C., Li, Z., 2020. Spatial
- Li, P., Li, W., Dumbrell, A.J., Liu, M., Li, G., Wu, M., Jiang, C., Li, Z., 2020. Spatial variation in soil fungal communities across paddy fields in subtropical China. mSystems 5 (1), e00704-19. https://doi.org/10.1128/mSystems.00704-19.
- Liu, B., Qu, Z., Ma, Y., Xu, J., Chen, P., Sun, H., 2021. *Eucalyptus* plantation age and species govern soil fungal community structure and function under a tropical monsoon climate in China. Front. Fungal Biol. 2, 1–13. https://doi.org/10.3389/ ffunb.2021.703467.
- Lombard, N., Prestat, E., van Elsas, J.D., Simonet, P., 2011. Soil-specific limitations for access and analysis of soil microbial communities by metagenomics. FEMS Microbiol. Ecol. 78, 31–49. https://doi.org/10.1111/j.1574-6941.2011.01140.x.
- Moges Y. Eshetu Z. Nune S. 2010 Ethiopian forest resources: current status and future management options in view of access to carbon finances. A report submitted to Ethiopian climate resarch and networking and UNDP. Addis Ababa, Ethiopia.
- Molla, G., Addisie, M.B., Ayele, G.T., 2023. Expansion of *Eucalyptus* plantation on fertile cultivated lands in the north-western highlands of Ethiopia. Remote Sens. 15, 661. https://doi.org/10.3390/rs15030661.
- Mosisa, G.B., Tassie, N., Adula, M., 2024. Current and future distribution of *Eucalyptus globulus* under changing climate in Ethiopia: implications for forest management. Environ. Syst. Res. 13, 4. https://doi.org/10.1186/s40068-024-00332-z.
- Ondov, B.D., Bergman, N.H., Phillippy, A.M., 2011. Interactive metagenomic visualization in a web browser. BMC Bioinformatics 12, 385. https://doi.org/ 10.1186/1471-2105-12-385.
- Peay, K., Garbelotto, M., Bruns, T., 2010. Evidence of dispersal limitation in soil microorganisms: isolation reduces species richness on mycorrhizal tree islands. Ecology 91, 3631–3640.
- Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D., 2012. Nonlinear mixed-effects models. R package version 3, 1–89.
- Pölme, S., Abarenkov, K., Henrik Nilsson, R., Lindahl, B.D., Clemmensen, K.E., Kauserud, H., Nguyen, N., Kjøller, R., Bates, S.T., Baldrian, P., Frøslev, T.G., Adojaan, K., Vizzini, A., Suija, A., Pfister, D., Baral, H.O., Järv, H., Madrid, H., Nordén, J., Liu, J.K., Pawlowska, J., Põldmaa, K., Pärtel, K., Runnel, K., Hansen, K., Larsson, K.H., Hyde, K.D., Sandoval-Denis, M., Smith, M.E., Toome-Heller, M., Wijayawardene, N.N., Menolli, N., Reynolds, N.K., Drenkhan, R.,

Maharachchikumbura, S.S.N., Gibertoni, T.B., Læssøe, T., Davis, W., Tokarev, Y., Corrales, A., Soares, A.M., Agan, A., Machado, A.R., Argüelles-Moyao, A., Detheridge, A., de Meiras-Ottoni, A., Verbeken, A., Dutta, A.K., Cui, B.K., Pradeep, C. K., Marín, C., Stanton, D., Gohar, D., Wanasinghe, D.N., Otsing, E., Aslani, F. Griffith, G.W., Lumbsch, T.H., Grossart, H.P., Masigol, H., Timling, I., Hiiesalu, I., Oja, J., Kupagme, J.Y., Geml, J., Alvarez-Manjarrez, J., Ilves, K., Loit, K., Adamson, K., Nara, K., Küngas, K., Rojas-Jimenez, K., Bitenieks, K., Irinyi, L., Nagy, L.L., Soonvald, L., Zhou, L.W., Wagner, L., Aime, M.C., Öpik, M., Mujica, M.I., Metsoja, M., Ryberg, M., Vasar, M., Murata, M., Nelsen, M.P., Cleary, M., Samarakoon, M.C., Doilom, M., Bahram, M., Hagh-Doust, N., Dulya, O., Johnston, P., Kohout, P., Chen, Q., Tian, Q., Nandi, R., Amiri, R., Perera, R.H., dos Santos Chikowski, R., Mendes-Alvarenga, R.L., Garibay-Orijel, R., Gielen, R., Phookamsak, R., Jayawardena, R.S., Rahimlou, S., Karunarathna, S.C. Tibpromma, S., Brown, S.P., Sepp, S.K., Mundra, S., Luo, Z.H., Bose, T., Vahter, T., Netherway, T., Yang, T., May, T., Varga, T., Li, W., Coimbra, V.R.M., de Oliveira, V. R.T., de Lima, V.X., Mikryukov, V.S., Lu, Y., Matsuda, Y., Miyamoto, Y., Kõljalg, U., Tedersoo, L., 2020. FungalTraits: a user-friendly traits database of fungi and funguslike stramenopiles. Fungal Divers. 105. https://doi.org/10.1007/s13225-020-00466-2

- R Core Team, 2022. R: A language and environment for statistical computing (Versión 4.2.2) [Software]. R Foundation for Statistical Computing. https://www.R-project. org/.
- Reeuwijk, L., 2002. Procedures for Soil Analysis, 6th ed. International Soil Reference and Information Centre, Wageningen, The Netherlands.
- Reverchon, F., Del Ortega-Larrocea, P.M., Pérez-Moreno, J., 2010. Saprophytic fungal communities change in diversity and species composition across a volcanic soil chronosequence at Sierra del Chichinautzin, Mexico. Ann. Microbiol. 60, 217–226. https://doi.org/10.1007/s13213-010-0030-7.
- Rudolph, S., Maciá-Vicente, J.G., Lotz-Winter, H., Schleuning, M., Piepenbring, M., 2018. Temporal variation of fungal diversity in a mosaic landscape in Germany. Stud. Mycol. 89, 95–104. https://doi.org/10.1016/j.simyco.2018.01.001.
- Sasse, J., Martinoia, E., Northen, T., 2018. Feed your friends: do plant exudates shape the root microbiome? Trends Plant Sci. 23, 25–41. https://doi.org/10.1016/j. tplants.2017.09.003.
- Shi, L., Mortimer, P., Ferry, S.J., et al., 2014. Variation in forest soil fungal diversity along a latitudinal gradient. Fungal Divers. 64, 305–315.
- Sisay, G., Gessesse, B., Kassie, M., Kebede, B., de Aza, C.H., 2024. Exploring drivers of land use/land cover transformations in Goang watershed Ethiopia: integrating local community perceptions with remote sensing data. Environ. Challenges 17, 101043. https://doi.org/10.1016/j.envc.2024.101043.
- Tan, K.H., 2005. Soil Sampling, Preparation and Analysis, second ed. CRC Press, Boca Raton.
- Tedersoo, L., Bahram, M., Põlme, S., Kõljalg, U., Yorou, N.S., Wijesundera, R., Ruiz, L.V., Vasco-Palacios, A.M., Thu, P.Q., Suija, A., Smith, M.E., Sharp, C., Saluveer, E., Saitta, A., Rosas, M., Riit, T., Ratkowsky, D., Pritsch, K., Põldmaa, K., Piepenbring, M., Phosri, C., Peterson, M., Parts, K., Pärtel, K., Otsing, E., Nouhra, E., Njouonkou, A.L., Nilsson, R.H., Morgado, L.N., Mayor, J., May, T.W., Majuakim, L., Lodge, D.J., Lee, S., Larsson, K.H., Kohout, P., Hosaka, K., Hiiesalu, I., Henkel, T.W., Harend, H., Guo, L.D., Greslebin, A., Grelet, G., Geml, J., Gates, G., Dunstan, W., Dunk, C., Drenkhan, R., Dearnaley, J., De Kesel, A., Dang, T., Chen, X., Buegger, F., Brearley, F.Q., Bonito, G., Anslan, S., Abell, S., Abarenkov, K., 2014. Global diversity and geography of soil fungi. Science 346 (6213), 1256688. https://doi.org/10.1126/ science.1256688.
- Tekalign, M., Flasse, C., Frankl, A., Van Rompaey, A., Poesen, J., Nyssen, J., Muys, B., 2018. Forest cover loss and recovery in an East African remnant forest area: understanding its context and drivers for conservation and sustainable ecosystem service provision. Appl. Geogr. 98, 133–142. https://doi.org/10.1016/j. apgeog.2018.07.014.
- Teketay, D., 2000. The ecological effects of *Eucalyptus*: ground for making wise and informed decision. In: The "*Eucalyptus* Dilemma" November 15. Addis Ababa, Ethiopia.
- Trappe, J., 1977. Selection of fungi for ectomycorrhizal inoculation in nurseries. Annu. Rev. Phytopathol. 15, 203–222.
- Trudell, S. a, Edmonds, R.L., 2004. Macrofungus communities correlate with moisture and nitrogen abundance in two old-growth conifer forests, Olympic National Park, Washington, USA. Can. J. Bot. 82, 781–800. https://doi.org/10.1139/b04-057.
- Vořiškova, J., Brabcová, V., Cajthaml, T., Baldrian, P., 2014. Seasonal dynamics of fungal communities in a temperate oak forest soil. New Phytol. 201, 269–278. https://doi. org/10.1111/nph.12481.
- Walker, T.S., Bais, H.P., Grotewold, E., Vivanco, J.M., 2003. Root exudation and rhizosphere biology. Plant Physiol. 132, 44–51. https://doi.org/10.1104/ pp.102.019661.
- Walkley, A., Black, I.A., 1934. An examination of the digestion method for determining soil organic matter and a proposed modification of the chromic acid titration method. Soil Sci. 34, 29–38.
- Wu, B., Hussain, M., Zhang, W., Stadler, M., Liu, X., Xiang, M., 2019. Current insights into fungal species diversity and perspective on naming the environmental DNA sequences of fungi. Mycology 10, 127–140. https://doi.org/10.1080/ 21501203.2019.1614106.
- Yirdaw, E., Luukkanen, O., 2003. Indigenous woody species diversity in *Eucalyptus globulus* Labill. ssp. globulus plantations in the Ethiopian highlands. Biodivers. Conserv. 12, 567–582. https://doi.org/10.1023/A:1022483700992.
- Zerga, B., Warkineh, B., Teketay, D., Woldetsadik, M., Sahle, M., 2021. Land use and land cover changes driven by expansion of eucalypt plantations in the Western Gurage watersheds, Central-South Ethiopia. Trees For. People 5, 100087. https://doi.org/ 10.1016/j.tfp.2021.100087.

G. Kewessa et al.

Zerga Belay 2015 Ecological impacts of eucalyptus plantation in Eza, Int. Interv. J. Agric. Soil Sci. 3, 47–51.

- Zhang, P., Luan, M., Li, X., Lian, Z., Zhao, X., 2021. The distribution of soil fungal communities along an altitudinal gradient in an alpine meadow. Glob. Ecol. Conserv. 31, e01838. https://doi.org/10.1016/j.gecco.2021.e01838.
- Zhu, P., Hu, X., Zou, Q., Yang, X., Jiang, B., Zuo, J., Bai, X., Song, J., Wu, N., Hou, Y., 2024. Shifts in fungal community diversity and potential function under natural forest succession and planted forest restoration in the Kunyu Mountains, East China. Ecol. Evol. 14, 1–14. https://doi.org/10.1002/ece3.70055.