

Exploring the future of photosynthetic biogas upgrading process

Laura Vargas-Estrada^{1,2} and Raúl Muñoz^{1,2}



The increasing demand for biomethane in Europe requires the urgent development of sustainable and cost-effective technologies to meet the ambitious target of producing 35 bcm of biomethane by 2030. Biological technologies stand as a promising integrating platform to purify biogas and valorize biogenic CO₂, while producing high-value chemicals. Photosynthetic biogas upgrading has demonstrated to be an attractive platform as microalgae can fix CO₂ while producing biomass that can be further valorized for biofuel, biofertilizer or biostimulant production. However, its high investment cost and the low microalgae biomass productivities have limited commercialization. Recently, the supplementation of nanoparticles obtained from olive-mill wastewater, has demonstrated to be a promising technique to improve biomass productivity, CO₂ removals, and the overall stability of the system. This review summarizes the current trends and future outlooks of this sustainable platform aiming at the development of novel integrated microalgae biorefineries devoted to biogas upgrading and high-value compounds production.

Addresses

¹ Institute of Sustainable Processes, University of Valladolid, Dr. Mergelina s/n., Valladolid 47011, Spain

² Department of Chemical Engineering and Environmental Technology, School of Industrial Engineering, University of Valladolid, Dr. Mergelina s/n., Valladolid 47011, Spain

Corresponding author: Muñoz, Raúl (raul.munoz.torre@uva.es)

Current Opinion in Environmental Sustainability 2026, 79:101605

This review comes from a themed issue on **Accelerating Transition to Circular Economy in the Water Sector**

Edited by **Souryabrata Mohapatra, Bénédicte Rulleau, Dukhabandhu Sahoo and Konstantinos Tsagarakis**

Available online xxxx

Received: 4 November 2025; Revised: 30 December 2025;
Accepted: 3 January 2026

<https://doi.org/10.1016/j.cosust.2026.101605>

1877-3435/© 2026 The Author(s). Published by Elsevier B.V. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction

The ambitious 2030 and 2050 environmental policies targeting net-zero emissions, in addition to the current

war conflicts in Europe, have accelerated the use of renewables, particularly biogas [1].

Biogas produced via anaerobic digestion (AD) stands as a cost-effective energy vector mainly composed of CH₄ (65–70%, v/v) and CO₂ (45–30%, v/v), and other traces of contaminants including H₂S, O₂, N₂, and volatile organic compounds in minor concentrations. To improve biogas energy power, CO₂ and H₂S must be removed to generate a high-quality biomethane analogous to natural gas.

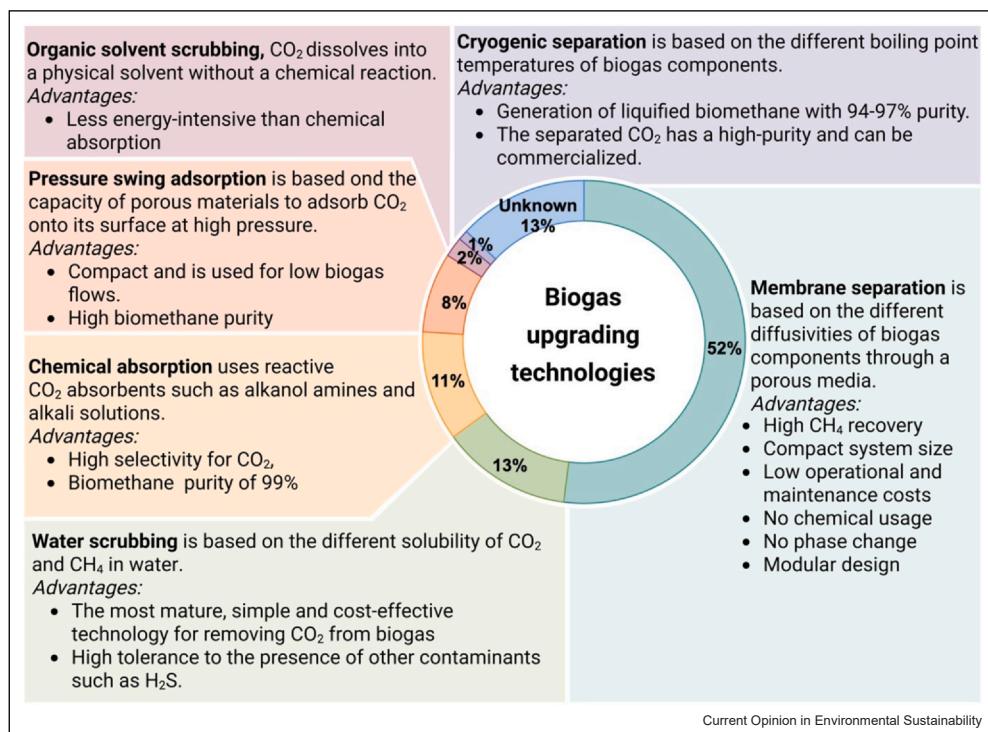
From the year 2023 to 2024, the European investments in the biogas market accounted for €345 million, and future investments of €28.4 billion for developments inside and outside Europe are foreseen by 2030 [2]. New plants producing biomethane will take 85% of the future investments, while the rest will be devoted to upgrading existing biogas-CHP plants to biomethane plants. This intense demand for biomethane plants fosters the urgent development of sustainable biogas upgrading technologies.

This work presents the current trends in biogas upgrading technologies with particular emphasis on photosynthetic biogas upgrading (PBU) process. The 50 peer-reviewed studies with the highest impact in the field, mainly from 2020 to 2025, are presented and discussed to support an equilibrated opinion regarding PBU. We first address the state-of-the-art of commercial biogas upgrading technologies, their position in the market, and the need for sustainable biogas upgrading technologies to meet the ambitious targets of the European Commission to produce 35 bcm of biomethane by 2030. Then, the trending biological biogas upgrading technologies are addressed, discussed, and compared to PBU. Thereafter, the fundamentals of PBU, process limitations, and opportunities to improve the system sustainability by relying on the use of nutrient-rich effluents are discussed. In addition, the current use of nanoparticle supplementation and its role in high-value compounds to improve PBU technoeconomic feasibility is addressed. Finally, the concluding remarks are presented.

Overview of biogas upgrading technologies and future market projections

Different physicochemical technologies are currently available in the market to upgrade biogas aiming to comply with international biomethane regulations for gas injection into the natural gas grid or use as autogas

Figure 1



Current biogas upgrading technologies commercially available and their market share in Europe during 2023 [3].

(Figure 1). Biomethane end-use will determine the quality of its composition, but in Europe, most countries are regulated by the EN 16723:2018, which requires a composition of CH₄ ≥90%, CO₂ ≤2%, O₂ ≤1% and a negligible concentration of H₂S for grid injection.

Typically, the upgrading process accounts for up to 40% of the total investment cost of a biomethane plant and 30% of the cost of biomethane, with CO₂ removal being the most expensive step, resulting in an average biomethane production cost between €9 and 25/GJ [2].

Sustainable and competitive upgrading technologies, namely biological upgrading technologies, can reduce biomethane production cost to €7/GJ. However, the coupled production of valuable by-products from biogas and digestate is crucial to increase biomethane competitiveness and circularity.

Current trends in biological biogas upgrading processes

In 2023, with the creation of the Biomethane Industrial Partnership, innovative upgrading technologies, namely, methanation (biological and chemical) and PBU, were projected as fundamental to accelerate the biomethane industry scale-up [3]. Despite biological technologies' great potential, their current high investment and

operational costs will require additional government support and political regulations before commercialization [4]. Hence, a biorefinery approach to concomitantly produce biomethane and high-value chemicals along with wastewater/digestate treatment could potentially improve their commercialization.

Recently, three biological upgrading technologies could potentially gain market weight (Table 1). Undoubtedly, biological methanation (BM) represents a more feasible technology as it can be directly integrated into existing plants by injecting H₂ to the digester or in an external reactor. BM can reduce the need to install further upgrading operational units and can increase biomethane yield without generating any additional subproducts [5–10]. Recently, Jønson et al. [7] demonstrated the potential of ex-situ BM in a pilot-scale biogas plant. After testing different operation configurations, CH₄ yields between 9.4 and 10.6 Nm³_{CH4}/m³_{Reactord} with CH₄ contents between 95.7% and 97.4%, were obtained. The system demonstrated robustness and ability to recover and maintain high BM efficiencies after H₂ supply losses; nonetheless, one of the main bottlenecks to address is the cost-effective H₂ production [7]. Currently, H₂ is commercially produced by water electrolysis, a high-energy-demand process that relies on the use of fossil fuels, which is neither attractive nor competitive

Table 1**Comparison of fundamentals of the trending biological biogas upgrading technologies.**

Biological upgrading technology	Methanation	Organic acid production	PBU
Technology fundamentals	Based on the conversion of H ₂ and CO ₂ to CH ₄ according to the chemical equation: CO ₂ + 4H ₂ → CH ₄ + H ₂ O Can be conducted via chemical and biological pathways External H ₂ is used to convert CO ₂ from biogas to CH ₄	Based on the sugar fermentation and CO ₂ capture from biogas with anaerobic bacteria, resulting in the production of organic acids.	Based on the use of microalgae-bacteria consortia to symbiotically remove CO ₂ , H ₂ S, NH ₃ , and other volatile organic compounds from biogas via metabolic pathways, mainly photosynthesis and nitrification
Microorganisms used	Hydrogenotrophic methanogenic archaea	Anaerobic bacteria such as <i>Actinobacillus succinogenes</i> 130Z, <i>Anaeribiospirillum succiniciproducens</i> , <i>Basfia succiniciproducens</i> , and <i>Mannheimia succiniciproducens</i>	Microalgae and nitrifying bacteria
System configurations	<i>In-situ</i> . Methanation is conducted in the existing anaerobic digester. Can be more cost-effective but bacterial competition within the reactor is higher. <i>Ex-situ</i> . Requires an external and independent reactor. Trickle-bed reactors have been widely used. More selective and can reach higher CO ₂ conversion	Still in early stages of deployment, but requires an anaerobic fermenter.	Consists of a HRAP, where microalgae-bacteria biomass is grown, connected to an absorption column where biogas is sparged to be upgraded to biomethane. A settler is placed in between to maximize biomass production, and to allow the circulation of microalgae biomass from the HRAP to the column
Technology advantages	High CO ₂ conversion In-situ methanation could reduce the need of external upgrading Enhanced biomethane yield	Production of valuable precursors for chemical industry No O ₂ contamination	High-value products production Use of sunlight as energy Nutrient recycling from digestates
Technology drawbacks	Need for a costly hydrogen source Low mass-transfer of H ₂ to microorganisms High H ₂ partial pressure inside the reactor can inhibit hydrolysis and acetogenesis, resulting in VFA accumulation High sensitivity to alkaline pH	Low bacteria selectivity to inorganic carbon. High concentrations of costly sugars are required	The use of microalgae increases the risk of O ₂ contamination in the upgraded biomethane. Land and water use High investment costs
Investment cost	CAPEX is around 20–200 €/MWh and OPEX 13 €/MWh (0.4 kWh electricity/Nm ³ biogas)	N.A.	Dependent on plant size, but an average plant. For instance a CAPEX of €6000/(Nm ³ /h) was estimated for a biogas capacity of 300 Nm ³ /h
Energy demand	It is estimated as the energy needed to produce H ₂ via electrolysis, 43–66 kWh/kg _{H2}	It has been estimated as 2–19% of the total manufacturing costs	It is estimated as 22.8 kWh/d for a biogas plant producing 120 m ³ _{biogas} /d
CH ₄ purity	>92%	85%	>92%
Subproduct obtained/yield	CH ₄ is the main product	Succinic acid is the targeted acid, yield of 0.92 g/g _{substrate}	Biomass, productivity up to 22 g/m ² d
Use of wastewater	Yes, centrate	Yes, potato wastewater	Yes, mainly centrate
References	[12,13,15,16,18,19]	[4–8]	[3,9–11,20]

N.A., not applicable. VFA, volatile fatty acids

[9]. Hence, other H₂ supplies route, including the use of renewable energy, must be developed to meet BM H₂ demand.

Conversely, the production of organic acids, namely succinic acid (SA), via CO₂ fermentation is emerging as a promising upgrading technology entailing a high purity biomethane > 90% [4,11–14]. Yet, this technology is still in its early stages and has not reached pilot-scale

demonstration. Moreover, the low selectivity of anaerobic bacteria to inorganic carbon has been a major drawback for its implementation [15], particularly as high CO₂ removal efficiencies are targeted during biogas upgrading. Key research advances have been made to increase bacterial CO₂ affinity, namely MgCO₃ supplementation, while sustaining high SA yields of 0.6 g_{SA}/g_{substrate}. However, the upgraded biomethane did not meet the EN 16723:2018 thresholds [15].

In this regard, PBU stands as a more feasible technology since non-expensive energy carriers to produce biomethane are needed. Microalgae robustness, ability to adapt to extreme climate conditions, and high affinity for CO₂, result in a biomethane that meets the EN 16723:2018 standard, highlighting the benefits and scalability of PBU. PBU is a more developed technology that has been extensively studied, optimized, and validated at pilot and demo-scale [16]. During PBU, the upgraded biomethane can be potentially contaminated with O₂, unlike the other biological technologies, but strategies to reduce O₂ in biomethane have been implemented to reduce O₂ to concentrations < 1%. To date, PBU has been limited to small-scale biogas plants mainly due to the large superficial areas needed for the photobioreactors. However, an improved microalgae metabolism and the coupled production of high-value chemicals would facilitate its implementation in medium and large-scale plants.

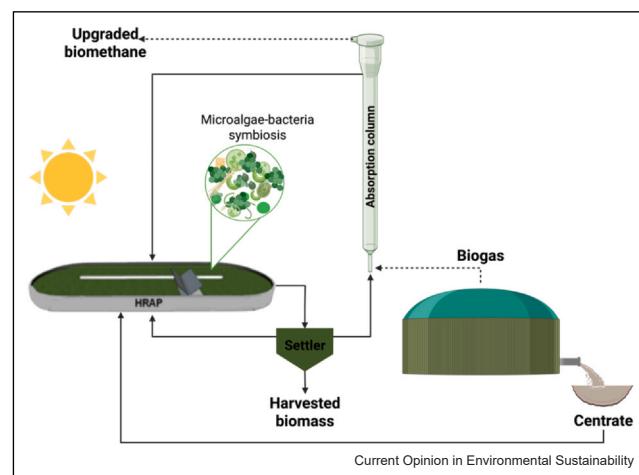
Photosynthetic biogas upgrading

Fundamentals of the process under a circular economy approach

PBU system configuration basically consists of a high rate algal pond (HRAP), where microalgae-bacteria biomass is grown, connected to an absorption column where biogas is sparged to be upgraded to biomethane (Figure 2).

During PBU, microalgae fix CO₂ from biogas via water photolysis and uptake nitrogen and phosphorus, via assimilatory mechanisms, in the so-called photosynthesis process, resulting in an excess of O₂ that promotes the oxidation of H₂S to SO₄²⁻ and NH₃ to NO₃⁻ by bacteria [9]. The intense demand of O₂ to remove contaminants from biogas results in a high-quality biomethane with

Figure 2



Schematic representation of the PBU system: a HRAP interconnected to an absorption column by a settler.

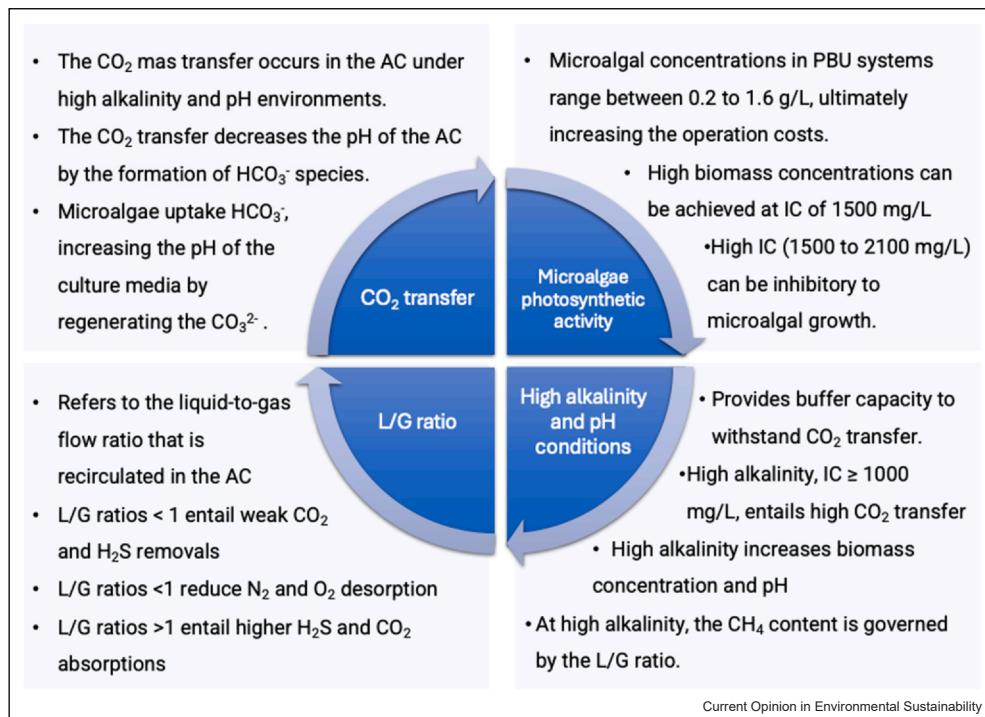
CO₂ contents as low as 0.2%, successfully meeting the EN 16723:2018 thresholds.

Indeed, microalgae cultivation requires a high water demand, increasing concerns for water security and agricultural competition, but the use of nutrient-rich effluents, namely centrate, the liquid fraction of digestates, could improve the system circularity [10]. Centrates have a high ammonia nitrogen content and require a posttreatment before being discharged [20]. Moreover, their high buffer capacity and low heavy metal concentration can trigger their use for PBU [10,21]. Hence, an integrated PBU process results in a dual environmental benefit by recycling nutrients to produce biomass and biomethane.

To date, centrates derived from the AD of food waste, pig farms, and sewage treatment plants have been used during PBU. Even if the nutrient removal efficiency depends on the centrate nutrient load and microalgae species, total nitrogen and phosphorus removals ranging between 70–100%, and 50–100% have been successfully achieved, respectively [22–25]. This intense nutrient uptake has resulted in microalgae biomass productivities of 0.2 g/Ld or 22 g/m²d, which still remain low for commercial exploitation [22].

The use of centrates during PBU has been limited to robust microalgae strains, usually tolerant to high ammonia concentration. Typically, microalgae species with poor commercial value dominate these cultivation broths, that is, *Chlorella* sp., *Chlorococcum* sp., *Desmodesmus* sp., *Scenedesmus* sp. and some cyanobacteria in a lower share, namely, *Anabaena* sp., and *Leptolyngbya* sp. [9]. This microalgae population dominance has directed biomass commercialization to biofertilizers or biostimulants [26–28]. However, wastewater/centrate cultivated microalgae fertilizers entail divergent opinions by the presence of heavy metals, antibiotics, and pathogens [29]. Nonetheless, García-Galán et al. [30], demonstrated that microalgae biomass cultivated in wastewater fulfilled the thresholds set by the European legislation for fertilizers and did not entail a potential risk for the cultivated crops. Similarly, Álvarez-González et al. [31], demonstrated that microalgae biomass cultivated in wastewater was successfully used as fertilizer for lettuce crops. Both microalgae biomass and the cultivated crops fulfilled the thresholds set by the European legislation for fertilizers during pathogens and contaminants of emerging concern analysis. Interestingly, Cd was detected in all the cultivated crops, suggesting Cd was present in the freshwater used for watering rather than in the microalgae biomass. Nevertheless, the use of centrates derived from distillery and food waste represent a less controversial nutrient source as a result of their low concentration of heavy metals and chemicals [27,32].

Figure 3



Key operational parameters limiting the process of PBU.

Process limitations and opportunities

PBU still faces some operational drawbacks and limitations for further implementation on a medium and large scale. Indeed, the composition of the upgraded biomethane depends on the mass transfer of CO₂ to the culture broth, which is related to operational parameters such as alkalinity, pH, liquid-to-gas (L/G) ratio within the column, and on the photosynthetic activity of microalgae (Figure 3).

The diurnal variability of microalgae metabolism, as well as the seasonal change across the year, significantly impact O₂ content in biomethane and microalgae growth. Different strategies have been implemented to decrease O₂ in the upgraded biomethane namely modifying L/G ratio in the absorption column, enhancing mixing in the reactor to foster O₂ stripping, the addition of a O₂ stripping column, and operating under mixotrophic cultivation [33]. However, in most cases, the CO₂ content in the upgraded biomethane is increased, and higher alkalinity and an improved microalgae metabolism would be necessary to effectively remove CO₂. Moreover, during winter, microalgae growth is reduced, compromising the overall system operation. Even if the use of a greenhouse has been proposed to increase the temperature during cold winters [34], light availability and the high investment costs of greenhouses could limit both microalgae growth and the economic feasibility of PBU.

Additionally, microalgae cultivation requires high superficial areas for HRAP deployment. For instance, a biogas plant producing 300 Nm³_{biogas}/h would require an illuminated area of 13.4 ha; hence, 1 Nm³_{biogas}/h requires 621 m² [17]. Inherently, larger biogas plants would require HRAPs with higher superficial area, increasing concerns with food competition. Even if installing the HRAP inside the biogas plant could potentially reduce its footprint, PBU would not be appropriate in biogas plants with limited land space.

Indeed, like most upgrading technologies, PBU still has drawbacks and limitations, but an integrated approach for centrate treatment, CO₂ biofixation, and high-value compounds production can overcome the above-mentioned limitations. Thereby, strategies to enhance CO₂ mass-transfer to the liquor broth and microalgae metabolism to efficiently uptake CO₂, are mandatory to reduce the size of the plants, boosting the feasibility of PBU and accelerating commercialization.

Position in the market

Today, PBU has a TRL of 6–7 and has been successfully demonstrated at semi-industrial scale [16]. It has a low-energy demand, and its operating and maintenance costs are estimated < €0.1/Nm³ of the upgraded biomethane. The investment cost of a standard PBU plant with a biogas production capacity of 300 Nm³/h can reach up to

€6000/Nm³, including the concentration and drying of the microalgal biomass. The commercialization of microalgae biomass for fertilizer production can generate revenues within five years, but the production of high-value compounds, namely pigments or nutraceuticals, could decrease the time to revenues.

In addition, meeting the ambitious biomethane production of 35 bcm in 2030 will result in 46 Mt of biogenic CO₂ that could be potentially exploited. Hence, PBU could directly use the biogenic CO₂ to produce high-value chemicals without requiring any additional separation or compression step for storage and transportation [35].

To date, a demo-scale PBU biorefinery composed of a 300 m² HRAP and a 0.5 m³ column is operating with a capacity to treat 13 m³_{biogas}/d entailing high biomass productivity and complete H₂S removal [16]. However, process scaling is needed to consolidate the feasibility of this technology.

Improving photosynthetic metabolism via nanoparticle addition

The addition of nanoparticles (NPs) to improve microalgae biotechnology has gained particular attention to boost microalgae growth and lipid accumulation [36]. Specifically, NPs synthesized via hydrothermal carbonization (HTC) of olive-mill wastewater (OMWW) have demonstrated great biocompatibility and outstanding biostimulant effect during microalgae growth.

HTC process is a well-known technology to produce a solid fertilizer, better known as hydrochar, and a nutrient-rich liquid, better known as tar, from sludge [37]. While tar has demonstrated to improve biogas yield during AD, both hydrochar and tar have been used to improve plant growth mainly due to the presence of essential micro-nutrients and bioactive compounds [38,39].

The addition of carbon-coated iron NPs synthesized via HTC has demonstrated an outstanding performance during PBU, increasing biomass productivities by two-fold while simultaneously fostering CO₂ removal efficiencies. This boosted effect has been identified at doses of 70 mg/L in *Chlorella sorokiniana* and mixed microalgae-bacteria batch cultures [40]. The scale-up of the process in a 180-L pilot-laboratory-scale system supplemented with 70 mg/L NPs entailed a two-fold increase in biomass concentration (> 3.5 g/L) and an increased CO₂ removal efficiency from 86% to 92%. Increasing NP dosage to 140 mg/L resulted in an improved biomass productivity up to 48 g/m²d with CO₂ removal efficiencies > 98% [41].

The effect of tar was also tested during PBU, and the addition of 1 mL_{tar}/L_{centratefed} presented similar

stimulant effects on microalgae growth and CO₂ removals as the NPs. The long-term effect of tar at daily supplementation of 2 mL_{tar}/L_{centratefed} supported a biomass productivity of 90 g/m²d and CO₂ removal efficiencies > 95%, despite harsh operational changes within the system [42]. Interestingly, the suppressed supplementation of tar reduced the CO₂ removal efficiency to 82% and the biomass concentration to 1.6 g/L underscoring the potential of tar to maintain a stable PBU operation.

The use of NPs could represent a potential risk for biodiversity and compromise the environment integrity. Toxic effects have been observed with TiO₂, CuO, ZnO, Cr₂O, AgNPs, CoNPs, and some iron oxide NPs [43–45]. Indeed, NP toxicity depends on their crystal phase and surface physicochemical properties, but their biocompatibility can be increased by adding a carbon layer of OMWW [46]. The size of OMWW-covered NPs typically ranges between 70 and 100 nm, preventing their internalization to microalgae cells, thereby the NPs can be recovered during the harvesting of biomass. However, if high-value compounds production from microalgae is intended, extraction methods are highly selective, that is, pigments, and the NPs would remain in the residual biomass. Hence, either the produced microalgae biomass or the residual biomass containing the NPs could be further used as plant fertilizers or AD boosters [37]. Nonetheless, the high stability of NPs would result in an expanded lifespan, and further studies regarding the final fate of NPs are necessary before scaling this technology.

Conversely, NP synthesis via HTC is a high-energy-demand process. Thus, the valorization of the produced syngas during HTC and the production of biofuels and/or high-value compounds, along with the production of the microalgal stimulating tar, are necessary to counteract the production costs of the overall PBU process [38].

Photosynthetic biogas upgrading under a biorefinery concept

The addition of NPs to microalgae cultures has diversified the valorization of biomass to the production of biofuels, nutraceuticals, pigments, and environmental applications, including efficient nutrient uptake and CO₂ capture [47]. Nevertheless, pigment production stands as a more cost-effective and attractive pathway to improve PBU profitability as the market value of microalgae-based pigments is projected to grow to €7.2 billion by 2034 [48].

Typically, pigment-producing microalgae have slow growth rates and low biomass productivities, resulting in high production costs. Hence, NP supplementation

stands as a promising technique to improve their growth and pigment yield. Recently, Au, OMWW-based, MoS₂, and ZnO NPs have improved the pigment content of *Chlorella zoingiensis*, *Coelastrella thermophila* var. *gobulina*, *Dunaliella salina*, and *Haematococcus lacustris* [48–50].

Despite the limited literature regarding integrated PBU and pigment production, the supplementation of 1 mL/L of tar derived from OMWW to *Coelastrella thermophila* var. *gobulina* cultures devoted to PBU increased the total carotenoid (TC) content to 838 µg/g [48]. Increasing tar concentration to 3 mL/L mediated a TC content of 1116 µg/g while maintaining a biomass productivity of 1.2 g/Ld. This confirmed the effect of tar on stimulating growth rate, which was supported by the accelerated nutrient consumption and a drastic decline in the photosynthetic activity, leading to an early endogenous phase where *C. thermophila* can synthesize carotenoids [48].

Certainly, the validation of these batch results in a continuous system using real centrate as culture media is needed to assess the feasibility of the integration of pigment production during PBU.

Conclusions

PBU has been successfully demonstrated at a semi-industrial scale. However, there are still some constraints to address, namely process stability and microalgal metabolism. Recently, nanoparticle supplementation has emerged as a cost-effective technique to stimulate microalgae metabolism, triggering stable system operation and resulting in a high-quality biomethane that meets the European standard for grid injection. In addition, the improved biomass production and valuable chemical products, that is, pigments, mediated by nanoparticle supplementation, could accelerate PBU commercialization. Nonetheless, the potential risks of nanoparticle supplementation needs to be studied before reaching industrial commercialization.

Data Availability

No data were used for the research described in the article.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This work was supported by the Department of Education of the Regional Government of Castilla y León and co-financed by the European Union through the European Regional Development Fund (ERDF) (Reference:

CLU-2025-2-06). Laura Vargas-Estrada acknowledges the Marie Skłodowska-Curie Individual Fellowship Grant Agreement No. 101148763.

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. European Commission. Biomethane. RePowerEU plan [Internet]. 2022. Available from: <https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:52022SC0230&from=EN>.
2. IEA. **World Energy Outlook Special Report Outlook for Biogas and Biomethane: A global geospatial assessment** [Internet]. 2025. Available from: www.iea.org.
3. BIP Europe. Task Force 5.1 launches report on the current state of the art of biomethane innovative technologies. 2023 [cited 2025 Dec 8]; Available from: <https://bip-europe.eu/2023/12/07/the-biomethane-industrial-partnership-launches-report-on-the-current-state-of-the-art-of-biomethane-innovative-technologies/>.
4. Carlos López J, Monsonís R, López de los Mozos E, Heredia F, Gómez-Pérez P: **Simultaneous biosuccinic production and biogas upgrading: exploring the potential of sugar-based confectionery waste within a biorefinery concept**. *Bioresour Technol* 2023, **384**:129362 Available from: <https://www.sciencedirect.com/science/article/pii/S0960852423007885>.
5. Feghhipour SE, Hatamipour MS, Amiri H, Nosrati M: **Continuous biogas production and ex-situ biomethanation in a trickling bed bioreactor under mesophilic and thermophilic conditions**. *Process Saf Environ Prot* 2024, **190**:1440–1449. Available from: <https://www.sciencedirect.com/science/article/pii/S0957582024009406>.
- This publication assessed ex situ biological methanation at pilot-scale using real biogas and hydrogen. Different operational configurations were studied to improve the purity of the upgraded biomethane. Interestingly, due to unintended process intermittences, the system showed robustness and ability to recover and maintain high biomethanation efficiencies.
6. Vinardell S, Feickert Fenske C, Heimann A, Cortina JL, Valderrama C, Koch K: **Exploring the potential of biological methanation for future defossilization scenarios: techno-economic and environmental evaluation**. *Energy Convers Manag* 2024, **307**:118339 Available from: <https://www.sciencedirect.com/science/article/pii/S0196890424002802>.
- This publication presents the techno-economic evaluation of biological methanation using H₂ produced via electrolysis and demonstrates that the main bottleneck of this technology is the production of H₂. However, biomethanation has a lower global warming impact than natural gas, positioning biomethane as a promising energy vector to reach economy decarbonization in the coming years.
7. Jønson BD, Tsapekos P, Tahir Ashraf M, Jeppesen M, Ejbye Schmidt J, Bastidas-Oyanedel JR: **Pilot-scale study of biomethanation in biological trickle bed reactors converting impure CO₂ from a Full-scale biogas plant**. *Bioresour Technol* 2022, **365**:128160 Available from: <https://www.sciencedirect.com/science/article/pii/S0960852422014936>.
8. Yörükü HC, Kamravamanesh D, Körkülü EO, Patel GH, Havukainen J, Karjunen H, et al.: **A comprehensive review on biological methanation processes: from gaseous feedstocks to biomethane**. *Energy Convers Manag* 2025, **341**:120075 Available from: <https://www.sciencedirect.com/science/article/pii/S0196890425005990>.
9. Abdelrahman AM, Abdelrazeq N, AlSayed A, Kakar FL, Muller C, Bell KY, et al.: **From process fundamentals to engineering perspectives: a technical review on in-situ biogas upgrading via the hydrogenotrophic methanogenesis pathway**. *Biomass Bioenergy* 2025, **200**:108043 Available from: <https://www.sciencedirect.com/science/article/pii/S0961953425004544>.
10. Ray S, Kuppam C, Pandit S, Kumar P: **Biogas Upgrading by Hydrogenotrophic Methanogens: An Overview**. *Waste Biomass*

Valoriz 2023, **14**:537-552, <https://doi.org/10.1007/s12649-022-01888-6>

11. Yang Z, Wu W, Zhao Q, Angelidaki I, Arhin SG, Hua D, et al.: **Enhanced direct gaseous CO₂ fixation into higher bio-succinic acid production and selectivity.** *J Environ Sci* 2024, **143**:164-175 (<https://www.sciencedirect.com/science/article/pii/S1001074223002346>).

12. Escanciano IA, Santos VE, Blanco Á, Ladero M: **Bioproduction of succinic acid from potato waste. Kinetic modeling.** *Ind Crops Prod* 2023, **203**:117124 (<https://www.sciencedirect.com/science/article/pii/S0926669023008890>).

This publication studied the use of potato waste as sustainable sugar source to carry out the process of CO₂ fermentation for the production of succinic acid. High SA yields up to 92% were obtained.

13. Lithourgidis AA, Kotsopoulos TA, Kalamaras SD, Skiadas IV, Kuglarz M, Vigato F, et al.: **Bio-succinic acid production, up to pilot scale, by fermentation of industrial candy waste with *Actinobacillus succinogenes* 130Z and its downstream purification process.** *J Environ Chem Eng* 2023, **11**:110920 (Available from, (<https://www.sciencedirect.com/science/article/pii/S2213343723016597>).

14. Kim H, Sang BI, Tsapekos P, Angelidaki I, Alvarado-Morales M: **Techno-economic analysis of succinic acid production from sugar-rich wastewater.** *Energy* 2023, **16**:3227 (<https://www.mdpi.com/1996-1073/16/7/3227>).

15. Vigato F, Angelidaki I, Woodley JM, Alvarado-Morales M: **Dissolved CO₂ profile in bio-succinic acid production from sugars-rich industrial waste.** *Biochem Eng J* 2022, **187**:108602 (<https://www.sciencedirect.com/science/article/pii/S1369703x22002716>).

16. URBIOFIN Urban Biorefinery [Internet]. 2025 [cited 2025 Dec 11]. Available from: (<https://www.urbiofin.eu>).

17. Rodero M del R, Ángeles R, García-Depraet O, Lebrero R, Muñoz R: **Chapter 5 - Recent advances on photosynthetic biogas upgrading to biomethane.** In *Biogas to Biomethane*. Edited by Yousaf A, Melville L. Woodhead Publishing; 2024:117-140. Available from: (<https://www.sciencedirect.com/science/article/pii/B9780443184796000107>).

18. Yang W, Li S, Qv M, Dai D, Liu D, Wang W, et al.: **Microalgal cultivation for the upgraded biogas by removing CO₂, coupled with the treatment of slurry from anaerobic digestion: a review.** *Bioresour Technol* 2022, **364**:128118 Available from: (<https://www.sciencedirect.com/science/article/pii/S0960852422014511>).

19. Bose A, Lin R, Rajendran K, O'Shea R, Xia A, Murphy JD: **How to optimise photosynthetic biogas upgrading: a perspective on system design and microalgae selection.** *Biotechnol Adv* 2019, **37**:107444 (<https://www.sciencedirect.com/science/article/pii/S0734975019301442>).

20. Ruiz Palomar C, García Álvaro A, Muñoz R, Repáraz C, Ortega MF, de Godos I: **Pre-commercial demonstration of a photosynthetic upgrading plant: investment and operating cost analysis.** *Processes* 2024, **12**, (<https://www.mdpi.com/2227-9717/12/12/2794>).

21. EBA. Exploring digestate's contribution to healthy soils [Internet]. 2024 [cited 2025 Dec 9]. Available from: (https://www.europeanbiogas.eu/wp-content/uploads/2024/03/Exploring-digestate-contribution-to-health-soils_EBA-Report.pdf).

22. Huang L, Liu J, Li Q, Wang C, Wu K, Wang C, et al.: **A review of biogas slurry treatment technology based on microalgae cultivation.** *Curr Opin Environ Sci Health* 2022, **25**:100315 Available from: (<https://www.sciencedirect.com/science/article/pii/S2468584421000878>).

23. Zhang H, Xu B, Zhao C, Liu J, Zhao Y, Sun S, et al.: **Simultaneous biogas upgrading and biogas slurry treatment by different microalgae-based technologies under various strigolactone analog (GR24) concentrations.** *Bioresour Technol* 2022, **351**:127033 Available from: (<https://www.sciencedirect.com/science/article/pii/S0960852422003625>).

24. Zhang J, Zhao C, Sun S, Zhao Y, Liu J: **Performance of different microalgae-based technologies in nutrient removal and biogas upgrading in response to various GR24 concentrations.** *Int Biodeterior Biodegrad* 2021, **158**:105166 Available from: (<https://www.sciencedirect.com/science/article/pii/S0964830520310970>).

25. Gao S, Hu C, Sun S, Xu J, Zhao Y, Zhang H: **Performance of piggery wastewater treatment and biogas upgrading by three microalgal cultivation technologies under different initial COD concentration.** *Energy* 2018, **165**:360-369 (<https://www.sciencedirect.com/science/article/pii/S0360544218319686>).

26. Herold C, Ishika T, Nwoba EG, Tait S, Ward A, Moheimani NR: **Biomass production of marine microalga *Tetraselmis suecica* using biogas and wastewater as nutrients.** *Biomass Bioenergy* 2021, **145**:105945 Available from: (<https://www.sciencedirect.com/science/article/pii/S0961953420304773>).

27. Almeida KM, Marangon BB, Ribeiro VJ, Castro J de S, da Silva J, Mattiello EM, et al.: **Microalgae cultivated in industrial wastewater as agricultural bioinputs: technical and life cycle assessment to support sustainable Production.** *ACS Omega* 2025, **10**:59208-59218, <https://doi.org/10.1021/acsomega.5c08477>

28. Slinksiene R, Sendzikiene E, Mikolaitiene A, Makareviciene V, Paleckiene R, Ragauskaite D: **Use of microalgae biomass for production of granular nitrogen biofertilizers.** *Green Chem Lett Rev* 2022, **15**:416-426, <https://doi.org/10.1080/17518253.2022.2071593>

29. Suleiman AKA, Lourenço KS, Clark C, Luz RL, da Silva GHR, Vet LEM, et al.: **From toilet to agriculture: fertilization with microalgal biomass from wastewater impacts the soil and rhizosphere active microbiomes, greenhouse gas emissions and plant growth.** *Resour Conserv Recycl* 2020, **161**:104924 Available from: (<https://www.sciencedirect.com/science/article/pii/S0921344920302421>).

30. García-Galán MJ, Matamoros V, Uggetti E, Díez-Montero R, García J: **Removal and environmental risk assessment of contaminants of emerging concern from irrigation waters in a semi-closed microalgae photobioreactor.** *Environ Res* 2021, **194**:110278 Available from: (<https://www.sciencedirect.com/science/article/pii/S0013935120311750>).

31. Álvarez-González A, Uggetti E, Serrano L, Gorchs G, Escolà Casas M, Matamoros V, et al.: **The potential of wastewater grown microalgae for agricultural purposes: contaminants of emerging concern, heavy metals and pathogens assessment.** *Environ Pollut* 2023, **324**:121399 Available from: (<https://www.sciencedirect.com/science/article/pii/S0269749123004013>).

This publication studied the contaminant content of wastewater cultivated microalgae, particularly heavy metal content, contaminants of emerging concern, and pathogens. The produced microalgal biomass was further used as fertilizers for the cultivation of lettuce. The same contaminant analyses were conducted to lettuce and the results showed that the cultivated crops met the European standards for fertilizers, except for Cd, but this was present in the control, suggesting that this contaminant was present in the freshwater. Hence, this study supported the feasible use of wastewater microalgal biomass as safe fertilizers.

32. Ollo E, Mückschel F, Velten H, Heyde BJ, Siemens J, Kämpfer P, et al.: **Fertilization with microalgal biomass of wastewater treatment high-rate algae ponds (HRAP): effects on the wheat root microbiome.** *Total Environ Microbiol* 2025, **1**:100033 Available from: (<https://www.sciencedirect.com/science/article/pii/S3050641725000333>).

33. Franco-Morgado M, Tabaco-Angoa T, Ramírez-García MA, González-Sánchez A: **Strategies for decreasing the O₂ content in the upgraded biogas purified via microalgae-based technology.** *J Environ Manag* 2021, **279**:111813, <https://doi.org/10.1016/j.jenvman.2020.111813>

34. Méndez L, García D, Pérez E, Blanco S, Muñoz R: **Photosynthetic upgrading of biogas from anaerobic digestion of mixed sludge in an outdoors algal-bacterial photobioreactor at pilot scale.** *J Water Process Eng* 2022, **48**:102891, <https://doi.org/10.1016/j.jwpe.2022.102891>

35. EBA. Statistical Report [Internet]. 2024 [cited 2025 Dec 8]. Available from: (https://www.europeanbiogas.eu/wp-content/uploads/2024/12/EBA_stats_report_complete_241204_preview.pdf).

36. Rana MS, Bhushan S, Sudhakar DR, Prajapati SK: **Effect of iron oxide nanoparticles on growth and biofuel potential of**

Chlorella spp. Algal Res 2020, 49:101942, <https://doi.org/10.1016/j.algal.2020.101942>

37. Kossińska N, Krzyżyska R, Ghazal H, Jouhara H: **Hydrothermal carbonisation of sewage sludge and resulting biofuels as a sustainable energy source.** *Energy* 2023, **275**:127337 Available from: <https://www.sciencedirect.com/science/article/pii/S0360544223007314>.
38. Hämäläinen A, Kokko M, Kinnunen V, Hilli T, Rintala J: **Hydrothermal carbonisation of mechanically dewatered digested sewage sludge—energy and nutrient recovery in centralised biogas plant.** *Water Res* 2021, **201**:117284 Available from: <https://www.sciencedirect.com/science/article/pii/S0043135421004826>.
39. Kossińska N, Grosser A, Kwapińska M, Kwapiński W, Ghazal H, Jouhara H, et al.: **Co-hydrothermal carbonization as a potential method of utilising digested sludge and screenings from wastewater treatment plants towards energy application.** *Energy* 2024, **299**:131456 Available from: <https://www.sciencedirect.com/science/article/pii/S0360544224012295>.
40. Vargas-Estrada L, Hoyos EG, Sebastian P, Muñoz R: **Influence of mesoporous iron based nanoparticles on Chlorella sorokiniana metabolism during photosynthetic biogas upgrading.** *Fuel* 2023, **333**:126362, <https://doi.org/10.1016/j.fuel.2022.126362>
41. Hoyos EG, Amo-Duodu G, Gulsum Kiral U, Vargas-Estrada L, Lebrero R, Muñoz R: **Influence of carbon-coated zero-valent iron-based nanoparticle concentration on continuous photosynthetic biogas upgrading.** *Fuel* 2024, **356**:129610 Available from: <https://linkinghub.elsevier.com/retrieve/pii/S001623612302224X>.
42. de Mello BS, Sarti A, Muñoz R: **Long-term effects of liquid nanoparticles on algal growth and photosynthetic biogas upgrading.** *Renew Energy* 2025, **251**:123467 Available from: <https://www.sciencedirect.com/science/article/pii/S0960148125011292>.
43. Nguyen MK, Moon JY, Lee YC: **Microalgal ecotoxicity of nanoparticles: an updated review.** *Ecotoxicol Environ Saf* 2020, **201**:110781 Available from: <https://www.sciencedirect.com/science/article/pii/S0147651320306205>.
44. Chen X, Zhang C, Tan L, Wang J: **Toxicity of Co nanoparticles on three species of marine microalgae.** *Environ Pollut* 2018, **236**:454-461 [cited 2020 Feb 12] <https://www.sciencedirect.com/science/article/pii/S0269749117337946>.
45. Baker TJ, Tyler CR, Galloway TS: **Impacts of metal and metal oxide nanoparticles on marine organisms.** *Environ Pollut* 2014, **186**:257-271, <https://doi.org/10.1016/j.envpol.2013.11.014>
46. Rizvi M, Gerengi H, Gupta P: **Functionalization of nanomaterials: synthesis and characterization.** Functionalized Nanomaterials for Corrosion Mitigation: Synthesis, Characterization, and Applications. American Chemical Society; 2022:1-26, <https://doi.org/10.1021/bk-2022-1418.ch001>
47. Ji CC, Deng SK, Xu XH, Cheng LH: **Integrated influence of sulfide modified nanoscale zero-valent iron on selective removal towards algal extracellular polymeric substances.** *J Environ Chem Eng* 2025, **13**:117592 Available from: <https://www.sciencedirect.com/science/article/pii/S2213343725022882>.
48. de Mello BS, Vargas-Estrada L, Sarti A, Muñoz R: **Enhancing pigment accumulation in *Coelastrella thermophila* var. *globulina* via olive-mill waste biostimulant addition during photosynthetic biogas upgrading.** *J Appl Phycol* 2025, **37**, <https://doi.org/10.1007/s10811-025-03684-w>.
This study presented the integrated PBU process with pigment production from microalgae. To improve pigment content, olive-mill wastewater tar and nanoparticles were added. The results showed that the supplementation of tar significantly increased total carotenoids content.
49. Li X, Sun H, Mao X, Lao Y, Chen F: **Enhanced photosynthesis of carotenoids in microalgae driven by light-harvesting gold nanoparticles.** *ACS Sustain Chem Eng* 2020, **8**:7600-7608, <https://doi.org/10.1021/acssuschemeng.0c00315>
50. Aizpuru A, González-Sánchez A: **Traditional and new trend strategies to enhance pigment contents in microalgae.** *World J Microbiol Biotechnol* 2024, **40**:272, <https://doi.org/10.1007/s11274-024-04070-3>